

Supplemental material

Within-subject Pooling of Biological Samples to Reduce Exposure Misclassification in Biomarker-based Studies

Flavie Perrier, Lise Giorgis-Allemand, Remy Slama, Claire Philippat

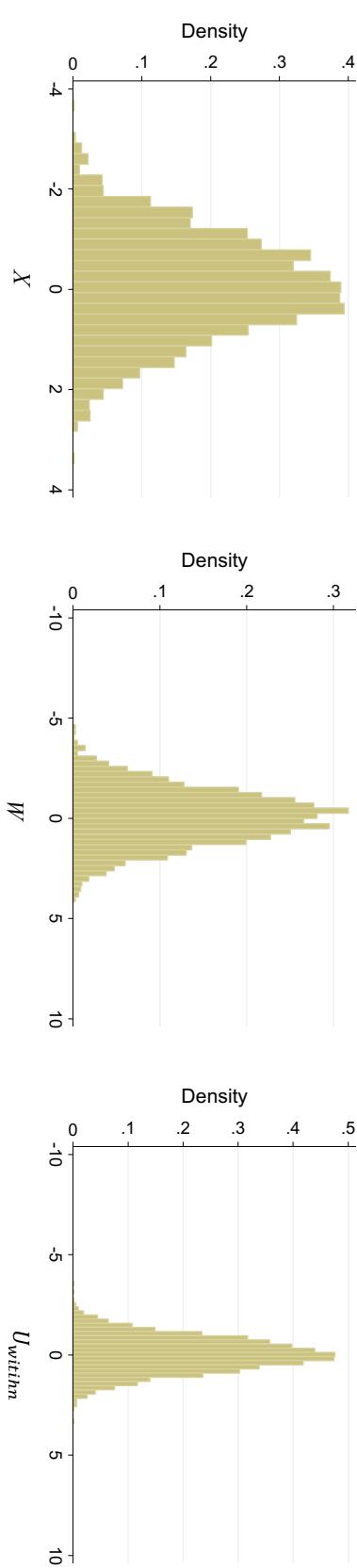
Table of contents

eFigure 1: Distribution of the simulated exposure (X, W) and the measurement error (U_{within}) in one of our simulated studies for chemicals with an ICC of 0.6 and 0.2	3
eFigure 2: Normalized urine concentrations of chemical A (ICC, 0.6) and chemical B (ICC, 0.2), measured with error, for 3 participants of one of our simulated studies	4
eFigure 3: Example of the extrapolation step of Simex using one of our simulated datasets (3000 subjects with 12 biospecimens each; real effect $\beta_1 = -100$ g, lack of between assay error ICC, 0.6)	15
eTable 1: Effect of the between-assay error on the effect estimates and statistical power of studies aiming at characterizing the associations between biomarker-based exposure and a continuous health outcome error. ICC decreased with increasing between-assay error.	5
eTable 2: Effect of the between-assay error on the effect estimates and statistical power of studies aiming at characterizing the associations between biomarker-based exposure and a continuous health outcome. ICC was kept constant whatever the value of the between-assay error.	7
eTable 3: Binary outcome, ICC of 0.6 - Effect estimates, Odds-Ratios and statistical power of the simulated studies aiming at characterizing the association between biomarker-based exposure to chemical A (ICC, 0.6) and a binary health outcome, according to the number of biospecimens collected per subject and the approach used to limit exposure misclassification	9
eTable 4: Binary outcome, ICC of 0.6 - Effect estimates, Odds-Ratios and statistical power of the simulated studies aiming at characterizing the association between a biomarker-based exposure to chemical B (ICC, 0.2) and a binary health outcome, according to the number of biospecimens collected per subject and the approach used to limit exposure misclassification	11
eTable 5: Effect estimates and type I error of studies aiming at characterizing the association between biomarker-based exposure to chemical A (ICC, 0.6) and B (ICC, 0.2) and a continuous outcome, according to the number of biospecimens collected per subject and the approach used to limit the effect of exposure misclassification	12
eTable 6: Effect estimates, Odds Ratios and type I error of studies aiming at characterizing the association between biomarker-based exposure to chemical A (ICC, 0.6) and B (ICC, 0.2) and a binary outcome, according to the number of biospecimens collected per subject and the approach used to limit the effect of exposure misclassification	13

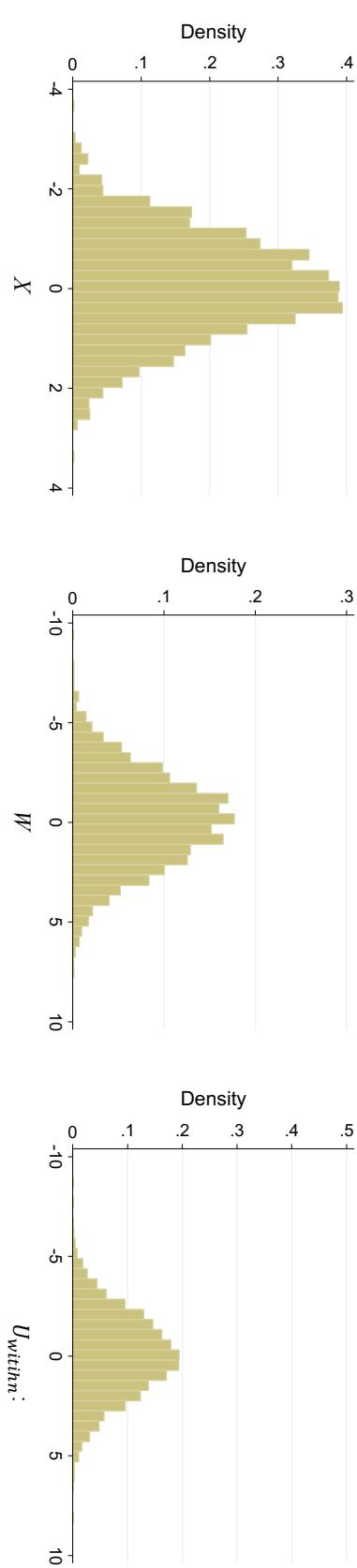
eAppendix 1: Description of the Regression Calibration and SIMEX methods	14
eAppendix 2: Code of our simulations for the continuous outcome	17

eFigure 1: Distribution of the simulated exposure (X, W) and the measurement error (U_{within}) in one of our simulated studies for chemicals with an ICC of 0.6 (A) and 0.2 (B) (lack of between-assay error).

A) ICC of 0.6



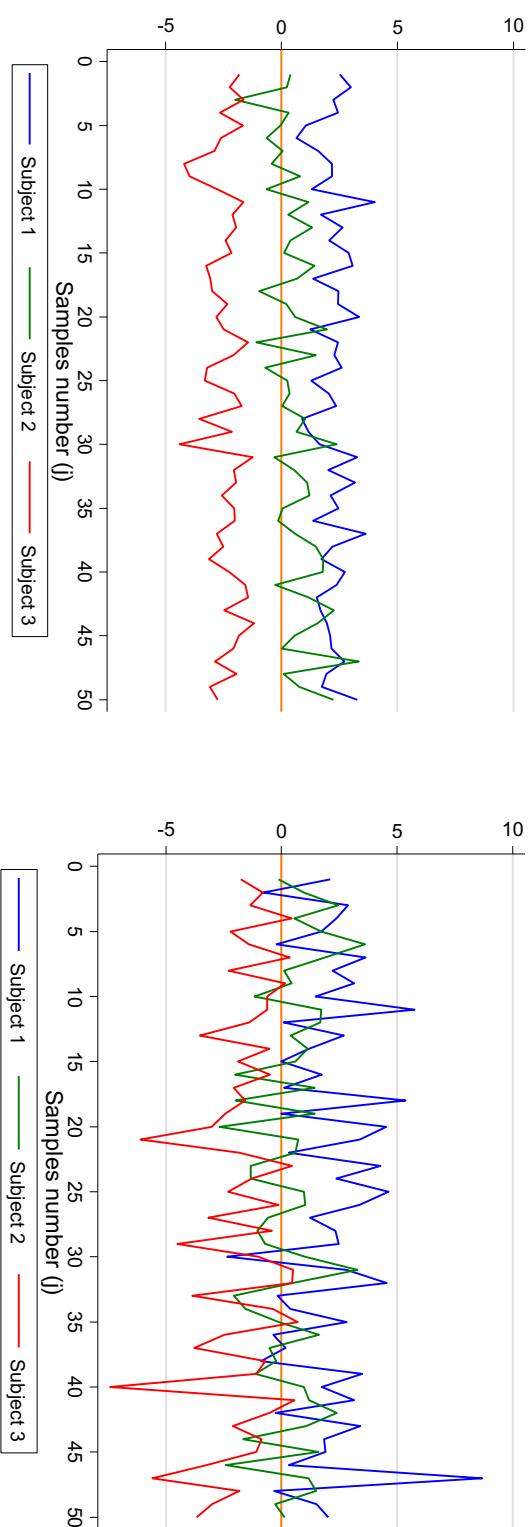
B) ICC of 0.2



Abbreviations: U_{within} : measurement error, W : error-prone variable, X : real exposure measured without error

eFigure 2: Normalized urine concentrations of chemical A (ICC, 0.6) and chemical B (ICC, 0.2), measured with error, for 3 participants of one of our simulated studies (lack of between-assay error variance).

A) Compound A with an ICC of 0.6



B) Compound B with an ICC of 0.2

The horizontal line represents the real unobserved average of urinary concentrations of the considered chemical over a toxicologically relevant exposure window, measured without error and centered around 0.

eTable 1: Effect of the between-assay error on the effect estimates and statistical power of studies aiming at characterizing the associations between biomarker-based exposure and a continuous health outcome (1000 simulations of studies with 3000 subjects each; real effect $\beta_1 = -100$ g). ICC decreased with increasing between-assay error.

		Chemical A						Chemical B							
σ_{Ussay}^2	Number of specimens per subject	Within-subject pooling			Simex calibration			Within-subject pooling			Simex calibration				
		Real effect ^a	ICC ^a	Effect estimate ^b	Power ^c	Bias (%) ^d	Effect estimate ^b	Power ^c	Bias (%) ^d	Effect estimate ^b	Power ^c	Bias (%) ^d	Effect estimate ^b		
0	1	-100	0.60	-60 ^e	0.71	40	-100	0.71	0	0.20	-20 ^e	0.32	80		
	2	-100	0.60	-75	0.82	25	-100	0.82	0	-95	0.85	5	-100	0.86	0
	5	-100	0.60	-89	0.88	11	-100	0.88	0	-100	0.89	0	-101	0.91	1
	10	-99	0.60	-93	0.90	6	-100	0.90	1	-100	0.91	0	-100	0.91	0
	50	-99	0.60	-97	0.90	2	-98	0.90	1	-98	0.92	0	0.20	-91	0.88
0.1	1	-101	0.60	-60 ^e	0.72	40	-100	0.72	1	0.20	-20 ^e	0.33	80		
	2	-99	0.60	-74	0.81	26	-98	0.81	1	-94	0.83	6	-99	0.84	0
	5	-101	0.60	-88	0.88	13	-100	0.88	1	-100	0.89	1	-101	0.9	0
	10	-100	0.60	-93	0.89	7	-99	0.89	1	-100	0.91	0	-100	0.92	0
	50	-99	0.60	-97	0.90	2	-98	0.90	1	-99	0.92	0	0.20	-91	0.88
0.5	1	-101	0.57	-57	0.71	43	-95	0.71	6	0.196	-20 ^e	0.32	80		
	2	-99	0.57	-69	0.79	31	-92	0.79	7	-93	0.82	7	-99	0.83	0
	5	-101	0.57	-82	0.86	19	-93	0.86	8	-100	0.89	1	-101	0.89	0
	10	-100	0.57	-86	0.87	14	-92	0.87	8	-100	0.91	0	-100	0.91	0
	50	-99	0.57	-89	0.88	10	-90	0.88	9	-99	0.92	0	0.196	-84	0.85
10	1	-101	0.46	-47	0.62	54	-78	0.62	23	0.18	-18 ^e	0.29	82		
	2	-99	0.46	-54	0.68	46	-72	0.69	27	-86	0.78	13	-99	0.79	0
	5	-101	0.46	-62	0.75	39	-70	0.75	31	-98	0.87	3	-101	0.87	0
	10	-100	0.46	-64	0.77	36	-69	0.77	31	-100	0.90	0	-100	0.90	0
	50	-99	0.46	-66	0.76	34	-66	0.76	33	-99	0.92	0	0.18	-63	0.74

^a ICC decreased with increasing between-assay error variance.

^b Mean of 1000 effect estimates. SIMEX and regression calibration cannot be used if only one biospecimen per subject is available.

^c Statistical power, estimated as the proportion of studies in which the p-value of the parameter characterizing the association between the error-prone exposure variable (W_{ij}) and the continuous outcome was below 0.05.

^d Difference between the real effect and the effect estimate, divided by the real effect.

^e Corresponds to a situation without pooling.

Note: The between assay error ($U_{ij,assay}$) was randomly drawn from a distribution with mean 0 and variance σ_{uassay}^2 . With regard to the relatively low coefficients of variation reported for short half-life chemicals (for example, coefficients of variation of about 60 replicates in a period of 9 months were between 3% and 10% for phenol biomarkers¹), we expected the between-assay error to be of low amplitude compared to the total variability. We however used a relatively large set of values for the between-assay variance (σ_{uassay}^2), ranging from 0.01, 0.1 to 0.5 (half of the variance of X_i). We hypothesize that each measured concentration (W_{ij}) in the j^{th} biospecimen of subject i was affected by the between-assay error, in addition to the error related to the within-subject error, such that:

$$W_{ij} = X_i + U_{ij,within} + U_{ij,assay} \quad i=1, \dots, n; j=1, \dots, k$$

The concentration measured in the pool of k biospecimens of equal volume from subject i was simulated as follows:

$$W_{ik,pool} = X_i + \frac{\sum_{j=1}^k U_{tj,indiv}}{k} + U_{ik,assay} \quad i=1, \dots, n; \quad k=1, \dots, 50$$

(This formula assumes that no chemical reaction takes place in the biospecimens before or after pooling).

With this approach, ICC decreased with increasing between-assay error, which makes it difficult to differentiate between the effect on bias due to the within-subject error ($U_{ij,within}$) and that due to the between-assay error. An alternative approach in which the ICC was kept constant whatever the value of the between-assay variance is presented in eTable 2.

eTable 2: Effect of the between-assay error on the effect estimates and statistical power of studies aiming at characterizing the associations between biomarker-based exposure and a continuous health outcome (1000 simulations of studies with 3000 subjects each; real effect $\beta_1 = -100$ g). ICC was kept constant whatever the value of the between-assay error.

Chemical A, ICC of 0.6 ^a										Chemical B, ICC of 0.2 ^a										Simex calibration												
σ_{Uassay}^2 of specimens effect ^b per subject	Within-subject pooling					Within-subject dissaturation					Simex calibration					Within-subject pooling					Within-subject dissaturation					Simex calibration						
	Effect estimate ^c	Power ^c	Bias (%) ^d	Effect estimate ^c	Power ^c	Bias (%) ^d	Effect estimate ^c	Power ^c	Bias (%) ^d	Effect estimate ^c	Power ^c	Bias (%) ^d	Effect estimate ^c	Power ^c	Bias (%) ^d	Effect estimate ^c	Power ^c	Bias (%) ^d	Effect estimate ^c	Power ^c	Bias (%) ^d	Effect estimate ^c	Power ^c	Bias (%) ^d	Effect estimate ^c	Power ^c	Bias (%) ^d					
0	1	-100	-60 ^e	0.71	40	-100 ^e	0.71	0	-100	0.82	25	-100	0.82	0	-95	0.85	5	-100	0.86	0	-34	0.50	66	-101	0.50	1	-54	0.52	45	-101	0.53	2
	2	-100	-75	0.82	25	-100	0.82	0	-100	0.88	11	-100	0.88	0	-100	0.89	0	-101	0.91	1	-56	0.71	43	-101	0.71	1	-82	0.72	18	-102	0.74	2
	5	-100	-89	0.88	11	-100	0.88	0	-100	0.90	6	-100	0.90	0	-100	0.91	0	-100	0.91	0	-72	0.80	28	-100	0.80	1	-93	0.82	6	-100	0.83	1
	10	-99	-93	0.90	2	-98	0.90	1	-98	0.92	0	-98	0.92	0	-91	0.88	8	-98	0.88	1	-98	0.89	0	-98	0.90	1	-98	0.90	0	-98	0.90	0
	50	-99	-97	0.90	2	-100 ^e	0.73	40	-100 ^e	0.73	0	-100 ^e	0.73	0	-20 ^e	0.31	80	-100 ^e	0.31	0	-34	0.50	67	-101	0.50	0	-55	0.53	46	-102	0.53	1
0.01	1	-100	-60 ^e	0.73	40	-101	0.82	0	-96	0.84	5	-101	0.85	0	-55	0.68	45	-99	0.68	1	-80	0.72	20	-99	0.73	1	-99	0.83	0	-99	0.83	0
	2	-100	-87	0.88	13	-99	0.88	1	-99	0.90	1	-100	0.90	0	-70	0.80	29	-99	0.80	0	-92	0.82	7	-99	0.83	0	-99	0.83	0	-99	0.83	0
	5	-100	-92	0.89	7	-98	0.89	1	-99	0.91	0	-99	0.91	0	-70	0.80	29	-99	0.80	0	-92	0.82	7	-99	0.83	0	-99	0.83	0	-99	0.83	0
	10	-99	-92	0.89	7	-98	0.89	1	-99	0.91	0	-99	0.91	0	-70	0.80	29	-99	0.80	0	-92	0.82	7	-99	0.83	0	-99	0.83	0	-99	0.83	0
	50	-101	-99	0.91	2	-100	0.91	1	-101	0.93	0	-101	0.92	0	-93	0.89	8	-100	0.89	1	-101	0.90	0	-101	0.91	0	-101	0.91	0	-101	0.91	0
0.1	1	-100 ^e	-60 ^e	0.74	40	-100 ^e	0.74	0	-100 ^e	0.74	0	-100 ^e	0.74	0	-20 ^e	0.31	80	-100 ^e	0.32	0	-33	0.50	67	-100	0.51	1	-55	0.53	46	-102	0.53	1
	2	-101	-73	0.80	27	-98	0.80	3	-96	0.84	5	-101	0.85	0	-53	0.66	47	-95	0.66	5	-80	0.71	20	-99	0.72	1	-99	0.82	0	-99	0.82	0
	5	-100	-82	0.86	18	-93	0.86	7	-99	0.90	1	-100	0.90	0	-67	0.79	33	-93	0.79	6	-92	0.81	7	-99	0.82	0	-99	0.82	0	-99	0.82	0
	10	-99	-86	0.87	13	-92	0.87	7	-99	0.91	0	-99	0.91	0	-67	0.79	33	-93	0.79	6	-92	0.81	7	-99	0.82	0	-99	0.82	0	-99	0.82	0
	50	-101	-91	0.88	10	-92	0.88	9	-101	0.93	0	-101	0.92	0	-86	0.86	15	-93	0.86	8	-101	0.90	0	-101	0.91	0	-101	0.91	0	-101	0.91	0
0.5	1	-100 ^e	-60 ^e	0.74	40	-100 ^e	0.74	0	-100 ^e	0.74	0	-100 ^e	0.74	0	-20 ^e	0.32	80	-100 ^e	0.32	0	-31	0.48	69	-94	0.48	7	-55	0.54	46	-102	0.53	1
	2	-101	-64	0.75	36	-86	0.75	15	-96	0.85	5	-101	0.87	0	-45	0.59	55	-81	0.59	19	-80	0.72	20	-99	0.72	1	-99	0.72	1	-99	0.72	1
	5	-100	-65	0.76	35	-73	0.76	27	-99	0.90	1	-100	0.90	0	-54	0.68	46	-76	0.68	24	-92	0.82	7	-99	0.83	0	-99	0.83	0	-99	0.83	0
	10	-99	-66	0.77	34	-70	0.77	29	-99	0.91	0	-99	0.91	0	-65	0.76	36	-70	0.76	31	-101	0.90	0	-101	0.90	0	-101	0.90	0	-101	0.90	0
	50	-101	-67	0.77	33	-68	0.77	32	-101	0.93	0	-101	0.92	0	-65	0.76	36	-70	0.76	31	-101	0.90	0	-101	0.90	0	-101	0.90	0	-101	0.90	0

^a ICC was constant whatever the value of the between-assay error variance.

^b Mean of 1000 effect estimates. SIMEX and regression calibration cannot be used if only one biospecimen per subject is available.

^c Statistical power, estimated as the proportion of studies in which the p-value of the parameter characterizing the association between the error-prone exposure variable (W_{ij}) and the continuous outcome was below 0.05.

^d Difference between the real effect and the effect estimate, divided by the real effect.

^e Corresponds to a situation without pooling.

Note: The simulation of the impact of between-assay error slightly differed compared to eTable 1. In eTable 1, the between assay error was added to the $U_{ij,within}$, while in eTable 6, we broke up the within-subject error $U_{ij,within}$ in two components: 1) the part of the error related to the individual ($U_{ij,indiv}$), 2) the between-assay error ($U_{ij,assay}$) due to the measurement such that:

$$U_{ij,within} = U_{ij,indiv} + U_{ij,assay} \quad i=1, \dots, n; j=1, \dots, k \quad (2)$$

With this approach, any increase in between-assay error is counterbalanced by a decrease in the error related to the individual ($U_{ij,indiv}$). Thus, the ICC is maintained constant (which was not the case in eTable 1). This might not be realistic but allows discriminating an effect of the between-assay error from an effect of the within-subject variability; it allows to discriminate, for a given ICC, which approach (pooling, SIMEX, regression calibration) is the most efficient to correct bias, according to whether the error predominantly comes from the individual or the assay.

eTable 3: Binary outcome, ICC of 0.6 - Effect estimates, Odds-Ratios and statistical power of the simulated studies aiming at characterizing the association between biomarker-based exposure to chemical A (ICC, 0.6) and a binary health outcome, according to the number of biospecimens collected per subject and the approach used to limit exposure misclassification (1000 simulations of studies with 3000 subjects each; real effect $\beta_1 = 0.262$ corresponding to an OR of 1.30, assuming lack of between-assay error)

Number of biospecimens per subject	Real effect ^b	Within-subject pooling				Within-subject pooling + a posteriori disattenuation				SIMEX				Regression calibration						
		OR ^a	Effect estimate ^b	95% CI ^c	Power ^d	OR ^a	Effect estimate ^b	95% CI ^c	Power ^d	OR ^a	Effect estimate ^b	95% CI ^c	Power ^d	OR ^a	Effect estimate ^b	95% CI ^c	Power ^d			
1	0.264	1.17	[0.03 ; 0.29]	0.69	40	1.31	0.264	[0.05 ; 0.49]	0.69	0	1.29	0.251	[0.06 ; 0.43]	0.77	5	1.31	0.265	[0.06 ; 0.46]	0.79	0
2	0.266	1.22	[0.05 ; 0.34]	0.77	25	1.31	0.265	[0.06 ; 0.45]	0.77	1	1.30	0.258	[0.08 ; 0.43]	0.82	2	1.31	0.263	[0.08 ; 0.43]	0.83	0
3	0.262	1.24	[0.07 ; 0.35]	0.82	18	1.31	0.263	[0.08 ; 0.43]	0.82	0	1.30	0.258	[0.07 ; 0.43]	0.82	2	1.30	0.258	[0.07 ; 0.44]	0.82	1
4	0.260	1.25	[0.06 ; 0.37]	0.81	15	1.30	0.258	[0.07 ; 0.44]	0.81	1	1.30	0.255	[0.07 ; 0.43]	0.82	2	1.30	0.258	[0.07 ; 0.44]	0.82	0
5	0.265	1.27	[0.09 ; 0.39]	0.86	12	1.31	0.265	[0.10 ; 0.44]	0.86	0	1.31	0.263	[0.09 ; 0.44]	0.85	1	1.31	0.265	[0.10 ; 0.44]	0.87	0
6	0.259	1.27	[0.08 ; 0.39]	0.83	10	1.30	0.259	[0.08 ; 0.43]	0.83	0	1.30	0.258	[0.08 ; 0.42]	0.85	1	1.30	0.259	[0.08 ; 0.43]	0.85	0
7	0.261	1.27	[0.07 ; 0.39]	0.84	9	1.30	0.261	[0.08 ; 0.43]	0.84	0	1.30	0.260	[0.08 ; 0.43]	0.85	0	1.30	0.261	[0.08 ; 0.43]	0.86	0
8	0.259	1.28	[0.09 ; 0.40]	0.86	7	1.30	0.260	[0.10 ; 0.43]	0.86	0	1.30	0.259	[0.09 ; 0.43]	0.87	0	1.30	0.260	[0.10 ; 0.43]	0.87	0
9	0.262	1.28	[0.08 ; 0.40]	0.86	7	1.30	0.262	[0.09 ; 0.43]	0.86	0	1.30	0.262	[0.08 ; 0.43]	0.87	0	1.30	0.262	[0.09 ; 0.43]	0.88	0
10	0.261	1.28	[0.08 ; 0.41]	0.85	6	1.30	0.261	[0.08 ; 0.44]	0.85	0	1.30	0.261	[0.08 ; 0.44]	0.86	0	1.30	0.261	[0.08 ; 0.44]	0.87	0
12	0.264	1.29	[0.09 ; 0.41]	0.86	5	1.31	0.264	[0.09 ; 0.44]	0.86	0	1.31	0.264	[0.09 ; 0.44]	0.87	0	1.31	0.264	[0.09 ; 0.44]	0.88	0
15	0.262	1.29	[0.09 ; 0.41]	0.88	4	1.30	0.262	[0.09 ; 0.42]	0.88	0	1.30	0.262	[0.09 ; 0.43]	0.88	0	1.30	0.262	[0.09 ; 0.42]	0.89	0
18	0.265	1.30	[0.09 ; 0.43]	0.88	4	1.31	0.265	[0.09 ; 0.44]	0.88	0	1.31	0.265	[0.09 ; 0.45]	0.88	0	1.31	0.265	[0.09 ; 0.44]	0.89	0
20	0.267	1.30	[0.09 ; 0.42]	0.88	3	1.31	0.266	[0.10 ; 0.43]	0.88	0	1.31	0.266	[0.10 ; 0.43]	0.90	0	1.31	0.266	[0.10 ; 0.43]	0.89	0
25	0.266	1.30	[0.08 ; 0.42]	0.88	3	1.31	0.266	[0.09 ; 0.43]	0.88	0	1.31	0.266	[0.09 ; 0.44]	0.89	0	1.31	0.266	[0.09 ; 0.43]	0.88	0
30	0.260	1.29	[0.09 ; 0.42]	0.87	2	1.30	0.260	[0.09 ; 0.42]	0.87	0	1.30	0.260	[0.09 ; 0.42]	0.88	0	1.30	0.260	[0.09 ; 0.42]	0.88	0
35	0.260	1.30	[0.09 ; 0.42]	0.86	2	1.30	0.260	[0.09 ; 0.43]	0.86	0	1.30	0.260	[0.09 ; 0.43]	0.88	0	1.30	0.260	[0.09 ; 0.43]	0.87	0
40	0.266	1.30	[0.09 ; 0.42]	0.89	2	1.31	0.266	[0.09 ; 0.43]	0.89	0	1.31	0.267	[0.09 ; 0.43]	0.90	0	1.31	0.266	[0.09 ; 0.43]	0.90	0
50	0.264	1.30	[0.09 ; 0.43]	0.88	1	1.31	0.264	[0.09 ; 0.43]	0.88	0	1.31	0.264	[0.09 ; 0.43]	0.88	0	1.31	0.264	[0.09 ; 0.43]	0.89	0

SIMEX and regression calibration cannot be used if only one biospecimen per subject is available

^a Mean of 1000 estimated odds ratio.

^b Mean of 1000 effect estimates

^c Empirical confidence interval, corresponding to the empirical 2.5 and 97.5 percentiles of the health effect estimates over the 1000 simulation runs.

^d Statistical power, proportion of studies in which the p-value of the parameter characterizing the association between the error-prone variables (W_{ij}) and the binary outcome was below 0.05.

Difference between the real effect and the effect estimate divided by the real effect

f Corresponds to a situation without pooling.

Note: The binary outcome (Y') was simulated as follows. First, $P(Y'_i = 1)$ was estimated according to a logistic regression model:

$$P(Y'_i = 1) = \frac{1}{1 + \exp(-(α' + β'_1 X_i))}$$

where $α' = -2.944$ (corresponding to a disease frequency of 5% in subjects with $X = 0$). The value of the parameter ($β'_1$) associated with real exposure X was 0.262, corresponding to an odds-ratio of 1.30 for each increase by 1 in exposure. For each subject, we then generated a random number between 0 and 1 drawn from a uniform distribution; we considered that the disease was present ($Y'_i = 1$) when the value of this random number was below $P(Y'_i = 1)$.

eTable 4: Binary outcome, ICC of 0.2 - Effect estimates, Odds-Ratios and statistical power of the simulated studies aiming at characterizing the association between a biomarker-based exposure to chemical B (ICC, 0.2) and a binary health outcome, according to the number of biospecimens collected per subject and the approach used to limit exposure misclassification (1000 simulations of studies with 3000 subjects each; real effect, $\beta_1 = 0.262$, corresponding to an OR of 1.30, assuming lack of between-assay error)

Number of biospecimens per subject	Real effect ^b	Within-subject pooling				Within-subject pooling + a posteriori dissattenuation				SIMEX				Regression calibration							
		Effect estimate ^b		95% CI ^c	Power ^d	Bias ^e	Effect estimate ^b		95% CI ^c	Power ^d	Bias ^e	Effect estimate ^b		95% CI ^c	Power ^d	Bias ^e	Effect estimate ^b		95% CI ^c	Power ^d	Bias ^e
		OR ^a	estimate ^b	(%)	(%)	(%)	OR ^a	estimate ^b	(%)	(%)	(%)	OR ^a	estimate ^b	(%)	(%)	(%)	OR ^a	estimate ^b	(%)	(%)	(%)
1	0.264	1.06 ^f	0.053	[-0.02 ; 0.13]	0.31	80	1.33 ^f	0.265	[-0.11 ; 0.65]	0.31	0	1.16	0.142	[-0.02 ; 0.31]	0.48	47	1.32	0.265	[-0.04 ; 0.58]	0.47	0
2	0.266	1.09	0.088	[-0.01 ; 0.19]	0.44	67	1.31	0.263	[-0.04 ; 0.56]	0.44	1	1.16	0.142	[-0.02 ; 0.31]	0.48	33	1.31	0.265	[-0.01 ; 0.50]	0.59	1
3	0.262	1.12	0.113	[0.00 ; 0.22]	0.56	57	1.31	0.264	[0.01 ; 0.50]	0.56	1	1.20	0.176	[0.01 ; 0.34]	0.57	26	1.30	0.256	[0.01 ; 0.48]	0.61	2
4	0.260	1.14	0.128	[0.01 ; 0.24]	0.58	51	1.3	0.255	[0.01 ; 0.48]	0.58	2	1.22	0.191	[0.01 ; 0.36]	0.60	20	1.31	0.264	[0.04 ; 0.49]	0.66	0
5	0.265	1.16	0.147	[0.02 ; 0.27]	0.64	45	1.31	0.264	[0.04 ; 0.49]	0.64	0	1.24	0.213	[0.03 ; 0.39]	0.66	15	1.30	0.259	[0.04 ; 0.47]	0.70	0
6	0.259	1.17	0.156	[0.02 ; 0.28]	0.68	40	1.30	0.259	[0.04 ; 0.47]	0.68	0	1.25	0.219	[0.04 ; 0.39]	0.70	15	1.30	0.259	[0.04 ; 0.47]	0.70	0
7	0.261	1.18	0.165	[0.04 ; 0.30]	0.71	37	1.30	0.260	[0.06 ; 0.46]	0.71	1	1.26	0.228	[0.05 ; 0.41]	0.71	13	1.30	0.260	[0.06 ; 0.46]	0.72	1
8	0.259	1.19	0.174	[0.05 ; 0.31]	0.73	33	1.30	0.261	[0.07 ; 0.47]	0.73	1	1.27	0.234	[0.07 ; 0.42]	0.73	10	1.30	0.261	[0.07 ; 0.47]	0.74	1
9	0.262	1.20	0.181	[0.03 ; 0.32]	0.76	31	1.31	0.262	[0.05 ; 0.46]	0.76	0	1.28	0.240	[0.05 ; 0.42]	0.76	9	1.31	0.262	[0.05 ; 0.46]	0.77	0
10	0.261	1.21	0.186	[0.04 ; 0.33]	0.76	29	1.30	0.261	[0.05 ; 0.46]	0.76	0	1.28	0.243	[0.04 ; 0.44]	0.78	7	1.30	0.261	[0.05 ; 0.47]	0.78	0
12	0.264	1.22	0.198	[0.06 ; 0.35]	0.78	25	1.31	0.264	[0.08 ; 0.46]	0.78	0	1.29	0.251	[0.07 ; 0.44]	0.77	5	1.31	0.264	[0.08 ; 0.46]	0.80	0
15	0.262	1.23	0.207	[0.06 ; 0.36]	0.80	21	1.31	0.262	[0.08 ; 0.45]	0.80	0	1.29	0.254	[0.07 ; 0.44]	0.81	3	1.31	0.262	[0.08 ; 0.45]	0.82	0
18	0.265	1.25	0.217	[0.06 ; 0.38]	0.83	18	1.31	0.265	[0.08 ; 0.46]	0.83	0	1.30	0.260	[0.08 ; 0.45]	0.84	2	1.31	0.265	[0.08 ; 0.46]	0.85	0
20	0.267	1.25	0.221	[0.07 ; 0.37]	0.84	17	1.31	0.266	[0.08 ; 0.45]	0.84	1	1.30	0.261	[0.08 ; 0.44]	0.84	2	1.31	0.266	[0.08 ; 0.45]	0.85	0
25	0.266	1.26	0.229	[0.07 ; 0.39]	0.83	14	1.31	0.266	[0.08 ; 0.45]	0.83	0	1.31	0.264	[0.08 ; 0.45]	0.85	1	1.31	0.266	[0.08 ; 0.45]	0.85	0
30	0.260	1.26	0.229	[0.07 ; 0.38]	0.84	12	1.30	0.260	[0.07 ; 0.43]	0.84	0	1.30	0.259	[0.07 ; 0.42]	0.85	0	1.30	0.260	[0.08 ; 0.43]	0.84	0
35	0.260	1.27	0.233	[0.07 ; 0.39]	0.83	11	1.30	0.259	[0.08 ; 0.44]	0.83	0	1.30	0.258	[0.08 ; 0.43]	0.84	1	1.30	0.259	[0.08 ; 0.44]	0.85	0
40	0.266	1.28	0.242	[0.08 ; 0.39]	0.87	9	1.31	0.266	[0.09 ; 0.43]	0.87	0	1.31	0.266	[0.09 ; 0.43]	0.87	0	1.31	0.266	[0.08 ; 0.43]	0.88	0
50	0.264	1.28	0.245	[0.09 ; 0.40]	0.85	7	1.31	0.264	[0.09 ; 0.43]	0.85	0	1.31	0.264	[0.09 ; 0.44]	0.86	0	1.31	0.264	[0.09 ; 0.43]	0.87	0

SIMEX and regression calibration cannot be used if only one biospecimen per subject is available.

^aMean of 1000 estimated odds ratio.

^bMean of 1000 effect estimates.

^cEmpirical confidence interval, corresponding to the empirical 2.5 and 97.5 percentiles of the health effect estimates over the 1000 simulation runs.

^dStatistical power, proportion of studies in which the p-value of the parameter characterizing the association between the error-prone variables (W_{ij}) and the binary outcome was below 0.05.

^eDifference between the real effect and the effect estimate, divided by the real effect.

^fCorresponds to a situation without pooling.

Note: The binary outcome (Y) was simulated as explained in the note below eTable 3.

eTable 5: Effect estimates and type I error of studies aiming at characterizing the association between biomarker-based exposure to chemical A ($\text{ICC}_A = 0.6$) and B ($\text{ICC}_B = 0.2$) and a continuous outcome, according to the number of biospecimens collected per subject and the approach used to limit the effect of exposure misclassification (1000 simulations of studies with 3000 subjects each; real effect $\beta_1 = 0 \text{ g}$, assuming lack of between-assay error).

Number of biospecimens per subject	Within-subject pooling			Within-subject pooling + dissattenuation			SIMEX			Regression calibration		
	Effect estimate ^a	95% CI ^b	Type I error ^c	Effect estimate ^a	95% CI ^b	Type I error ^c	Effect estimate ^a	95% CI ^b	Type I error ^c	Effect estimate ^a	95% CI ^b	Type I error ^c
Compound A, ICC of 0.6												
1	0 ^d	[45; 45]	5%	0 ^d	[-76; 75]	5%						
2	0	[-52; 52]	5%	0	[-69; 70]	5%	0	[-66; 67]	7%	0	[-69; 70]	7%
5	0	[-56; 57]	6%	0	[-64; 64]	6%	-1	[-66; 63]	8%	0	[-64; 64]	7%
10	0	[-56; 56]	4%	0	[-60; 60]	4%	0	[-59; 61]	6%	0	[-60; 60]	6%
50	2	[-56; 59]	4%	2	[-56; 60]	4%	2	[-58; 58]	7%	2	[-56; 60]	7%
Compound B, ICC of 0.2												
1	0 ^d	[-25; 27]	5%	0 ^d	[-124; 136]	5%						
2	0	[-34; 35]	5%	-1	[-103; 105]	5%	0	[-56; 58]	6%	-1	[-99; 110]	6%
5	-1	[-49; 42]	5%	-1	[-88; 76]	5%	-1	[-72; 62]	7%	-2	[-88; 76]	7%
10	0	[-47; 48]	4%	0	[-66; 67]	4%	0	[-61; 65]	5%	0	[-66; 67]	5%
50	2	[-53; 58]	5%	2	[-57; 63]	5%	2	[-57; 64]	7%	2	[-57; 63]	7%

^aMean of 1000 effect estimates.

^bEmpirical confidence interval, corresponding to the empirical 2.5 and 97.5 percentiles of the health effect estimates over the 1000 simulation runs.

^cType I error (false positive association), proportion of studies in which the p-value of the parameter characterizing the association between the error-prone variables and health outcome was below 0.05.

^dCorresponds to a situation without pooling.

eTable 6: Effect estimates, Odds Ratios and type I error of studies aiming at characterizing the association between biomarker-based exposure to chemical A (ICC, 0.6) and B (ICC, 0.2) and a binary outcome, according to the number of biospecimens collected per subject and the approach used to limit the effect of exposure misclassification (1000 simulations of studies with 3000 subjects each; real effect $\beta_1 = 0$ g, assuming lack of between-assay error).

Number of biospecimens per subject	Within-subject pooling			Within-subject pooling + dissattenuation			SIMEX			Regression calibration		
	OR ^a	Effect estimate ^b	95% CI ^c	OR ^a	Effect estimate ^b	95% CI ^c	OR ^a	Effect estimate ^b	95% CI ^c	OR ^a	Effect estimate ^b	95% CI ^c
Compound A, ICC of 0.6												
1	1.00 ^e	-0.002 ^e	[-0.13; 0.12]	5%	1.00 ^e	-0.003	[-0.22; 0.20]	5%	1.01	0.005	[-0.18; 0.19]	6%
2	1.01	0.004	[-0.14; 0.15]	5%	1.01	0.006	[-0.19; 0.21]	5%	1.00	-0.002	[-0.18; 0.17]	6%
5	1.00	-0.001	[-0.15; 0.16]	4%	1.00	-0.002	[-0.17; 0.18]	4%	1.00	0.000	[-0.17; 0.17]	6%
10	1.00	-0.001	[-0.16; 0.16]	5%	1.00	-0.001	[-0.17; 0.17]	5%	1.00	-0.001	[-0.17; 0.17]	6%
50	1.01	0.001	[-0.17; 0.16]	5%	1.01	0.001	[-0.17; 0.16]	5%	1.01	0.001	[-0.17; 0.17]	6%
Compound B, ICC of 0.2												
1	1.00 ^e	-0.001 ^e	[-0.08; 0.07]	5%	1.01 ^e	-0.007	[-0.39; 0.36]	5%	1.01	0.004	[-0.15; 0.16]	7%
2	1.00	0.003	[-0.09; 0.10]	6%	1.02	0.008	[-0.28; 0.30]	6%	1.01	-0.003	[-0.18; 0.17]	6%
5	1.00	-0.002	[-0.13; 0.12]	5%	1.00	-0.003	[-0.23; 0.21]	5%	1.00	-0.003	[-0.18; 0.18]	6%
10	1.00	-0.002	[-0.14; 0.14]	5%	1.00	-0.003	[-0.20; 0.19]	5%	1.00	-0.003	[-0.18; 0.17]	6%
50	1.00	0.001	[-0.17; 0.16]	5%	1.00	0.001	[-0.18; 0.17]	5%	1.01	0.001	[-0.18; 0.17]	6%

^a Mean of 1000 estimated odds ratio.

^b Mean of 1000 effect estimates.

^c Empirical confidence interval, corresponding to the empirical 2.5 and 97.5 percentiles of the health effect estimates over the 1000 simulation runs.

^d Type I error (false positive association), proportion of studies in which the p-value of the parameter characterizing the association between the error-prone variables and health outcome was below 0.05.

^e Corresponds to a situation without pooling

eAppendix 1: Description of the Regression Calibration and SIMEX methods²

Regression Calibration:

Regression calibration algorithm consists in estimating the unobserved exposure X_i (real exposure measured without error) using the repeated error-prone measurements W_{ij} and using the predicted values of X_i in the regression model estimating the association with the outcome:

$$Y_i = \beta_0 + \hat{\beta}_1 \widehat{E}(X_i | \bar{W}_i)$$

SIMEX:

SIMEX measurement error model includes a simulation and an extrapolation step.³ The simulation step consists of adding random pseudo error ($\sqrt{\gamma}U_{S,i,j}$) to the original values of the error-prone variable W_{ij} and fitting the regression model (continuous outcome):

$$Y_i = \alpha + \beta_{S,\gamma} (W_{ij} + \sqrt{\gamma}U_{S,i,j}) + \varepsilon_i$$

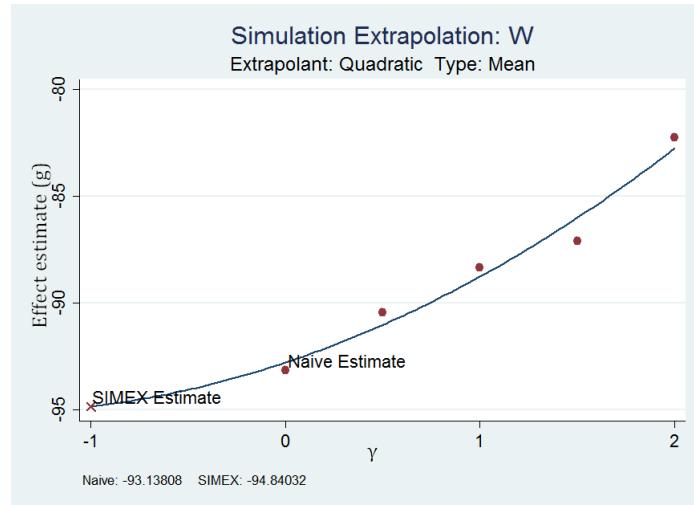
50 simulations with varying values of $U_{S,i,j}$ (normal random variable with mean, 0; SD; σ_{U_S}) are performed and the mean ($\bar{\beta}_\gamma$) of the biased estimators $\beta_{S,\gamma}$ is computed. The procedure is repeated for each value of γ in (0, 0.5, 1, 1.5 and 2).

The extrapolation step consists of modelling $\bar{\beta}_\gamma$ as a function of the measurement error variance:

$$\bar{\beta}_\gamma = \frac{\sigma_x^2}{\sigma_x^2 + \sigma_u^2(1+\gamma)} \hat{\beta}_1$$

and extrapolating to the case with no measurement error ($\gamma = -1$) in order to obtain the unbiased estimator (see eFigure 3).

eFigure 3: Example of the extrapolation step of Simex using one of our simulated datasets (3000 subjects with 12 biospecimens each; real effect $\beta_1 = -100$ g, $\sigma_{Uassay}^2 = 0$; ICC, 0.6)



References of the Supplemental Material

1. Philippat C, Botton J, Calafat AM, Ye X, Charles MA, Slama R, Group ES. Prenatal exposure to phenols and growth in boys. *Epidemiology* 2014;25(5):625-35.
2. Carroll RJ, Ruppert D, Stefanski LA, eds. Measurement Error in Nonlinear Models, A Modern Perspective, Second Edition London: Chapman & Hall., 1995.
3. Cook JR, Stefanski A. Simulation-Extrapolation estimation in parametric measurement error models. . *J Am Stat Assoc* 1994;89:1314–1328.

```

Codes for balanced desing_continuous outcome_ICC_0.2_0.6_2015_07_15 - Printed on 7/15/2015 11:37:56 AM
1  ****
2  /* Exposure measurement error in biomarker based studies
3   Codes for the continous outcome, balanced desing
4   (Codes for the binary outcome are available upon request) */
5  ****
6
7  -----
8  Creation : 22/11/13 (F.Perrier)
9  Modification : 21/10/14 (C.Philippat)
10 Modification : 25/03/15 (F.Perrier)
11 Modification : 28/05/2015 (F. Perrier)
12 Software used: STATA/SE, version 13 (StataCorp, College Station, TX, USA)
13
14 Package that need to be install before running the program :
15 net from http://www.stata.com/merror
16 net install merror // package needed in order to use simex and rcal fonctions
17
18 -----
19 * version 12.1
20
21 cap program drop _all
22 clear all
23 *set matsize 6000
24 set more off
25
26 timer on 1
27
28 /* Definitions of the paths to folders in which we will store the
29 simulated datasets and the results of the simulation */
30 ****
31 cd "D:\Erreur de mesure"
32 global simulation = "Simulations\reg_lin_test 25-03-15" /*dataset storage */
33 global results = "Resultats\reg_lin_test 25-03-15" /*result storage */
34
35 /* Definition of the parameters used in the simulation */
36 ****
37 global b0 = 14900          /* Intercept of the regression model */
38 global b1 = -100 /*0*/      /* Effect of the real exposure (X) on the outcome (Y) */
39 global nb_indiv = 3000     /*500 1000 2000 4000*/ /* Number of subjects in the simulated population */
40 global nb_simul = 2        /* Number of repetitions / simulations */
41 global corr_list = "0.6 0.2" /* Intraclass correlation coefficient observed between repeated measurements */
42 global sample_list = "1 2 3 4 5 6 7 8 9 10 12 15 18 20 25 30 35 40 50" /* List containing the different
43 numbers of biospecimens selected for each participants to study the association with the health outcome */
44 global var_Uassay_list "0 0.01 0.1 0.5" /* Variance of between-assay error */
45
46 -----
47 * Folder creation (needed datasets and results storage)
48 * mkdir "...."
49 /*
50 * datasets
51 *****
52 foreach c in $corr_list{
53   mkdir "$simulation\Correlation `c'"
54 }
55
56 foreach b in $b1 0 {
57   foreach c in $corr_list {
58     mkdir "$simulation\Correlation `c'\beta `b'"
59   }
60 }
61
62 foreach n in 500 1000 2000 3000 4000 {
63   foreach b in $b1 0 {
64     foreach c in $corr_list {
65       mkdir "$simulation\Correlation `c'\beta `b'\n `n'"
66     }
67   }
68 }
69
70 * Results
71 *****
72
73 foreach c in $corr_list {
74   mkdir "$results\Correlation `c'"
75 }
76
77 foreach b in $b1 0 {
78   foreach c in $corr_list {
79     mkdir "$results\Correlation `c'\beta `b'"
80   }
81 }
82
83 foreach n in 500 1000 2000 3000 4000 {
84   foreach b in $b1 0 {
85     foreach c in $corr_list {
86       mkdir "$results\Correlation `c'\beta `b'\n `n'"
87     }
88 }
89

```

```

88 }
89 }
90 */
91 */
93 -----
94 */
95 /* ****
96 DATA SET SIMULATION
97 In this section we simulated the datasets
98 * Notations:
99 X: real exposure
100 Uwithin: within-subject error
101 Uassay: between-assay error
102 W: Exposures measured with error
103 Y: Continuous health outcome
104 *****/
105
106 foreach var_Ua in $var_Uassay_list{
107     global var_Uassay=`var_Ua` /* Variance of the between-assay error */
108     di in green "variance between-assay : $var_Uassay"
109
110    foreach corr in $corr_list{
111        set seed 18102013
112        global c = `corr' /* ICC */
113        di in green " Data set simulation for beta:$b1 ICC:$c"
114
115        forvalues i = 1/$nb_simul {
116            di in yellow "Simulation number : " `i' " / " $nb_simul
117
118            foreach j in $sample_list{
119                /* Simulation of X (True exposure) */
120                global fixed_mean_X = 0 // mean
121                global fixed_var_X = 1 // variance
122                mat m_X = $fixed_mean_X
123                mat sd_X = sqrt($fixed_var_X)
124                qui drawnorm X, mean(m_X) sds(sd_X) n($nb_indiv)
125                qui sum X
126                global var_X_true = r(Var)
127
128
129                /* Simulation of Uassay (inter-assay error) */
130                mat m_Uassay = J(1, `j', 0)
131                mat sd_Uassay = J(1, `j', sqrt($var_Uassay))
132                if `j'==1{
133                    qui drawnorm Uassay_1, mean(m_Uassay) sds(sd_Uassay) n($nb_indiv)
134                }
135                else {
136                    qui drawnorm Uassay_1-Uassay_`j', mean(m_Uassay) sds(sd_Uassay) n($nb_indiv)
137                }
138
139                /* Simulation of Uwithin (within-subject error) */
140                global var_Uwithin = $var_X_true * (1/$c -1) // variance of error Uwithin
141                mat m_Uwithin = J(1, `j', 0)
142                mat sd_Uwithin = J(1, `j', sqrt($var_Uwithin))
143                if `j'==1{
144                    qui drawnorm Uwithin_1, mean(m_Uwithin) sds(sd_Uwithin) n($nb_indiv)
145                }
146                else {
147                    qui drawnorm Uwithin_1-Uwithin_`j', mean(m_Uwithin) sds(sd_Uwithin) n($nb_indiv)
148                }
149
150                /* Computation of the concentration measured in the pool */
151                qui egen mean_Uwithin = rowmean(Uwithin_1-Uwithin_`j') /* Mean of the within-subject error
152                error */
153                qui gen pool = X + mean_Uwithin + Uassay_1 /* Concentration measured in the pool */
154
155                /* Simulation of the concentration measured in each biospeciment (W) */
156                forvalues k = 1/`j' {
157                    qui gen W_`k' = X + Uwithin_`k' + Uassay_`k'
158                }
159
160
161                /* Simulation of the continuous health outcome */
162                qui drawnorm E, mean(0) sds(1650) n($nb_indiv) /* Simulation of the model error,
163                sds(1650) is the SD observed in the French mother child cohort EDEN, for the
164                weight at 3 years */
165                qui gen Y = $b0 + $b1*X + E
166                qui save "$simulation/Correlation 0$c/beta $b1/n
$nb_indiv/simul`i'_`j'samples_n$(nb_indiv}_varUass$(var_Uassay).dta", replace
167
168                drop *
169            }
170        }
171    }
172
173

```

```

174 /*-----*/
175
176 /* **** ANALYSES ****
177 In this section we studied the associations between the outcome (Y) and the real exposure (1) or the
178 error prone variable (2) using 3 different approaches: within-subject pooling (2a), Regression
179 Calibration (2b), Simex (2c). The a posteriori disattenuation is presented line 283.
180 **** */
181
182 foreach corr in $corr_list{
183     global c = `corr' /* ICC */
184     di in green " Analyses for beta:$b1 ICC:$c"
185
186     forvalues i = 1/$nb_simul {
187         di in yellow "Simulation number : " `i' " / " $nb_simul
188
189         local it = 0 /* sample list iterator */
190         foreach j in $sample_list{
191             local it = `it' + 1
192             use "$simulation/Correlation 0$c/beta $b1/n
$nb_indiv/simul`i'_`j'samples_n${nb_indiv}varUass${var_Uassay}.dta", clear
193
194             /* 1- Calculation and storage of the real effect; beta coefficients obtained when we used the
195                real exposure X */
196             qui regress Y X
197             mat table_true = r(table)
198             global beta = table_true[1,1]           // regression coefficient
199             global p = table_true[4,1]            // p-value
200             mata : res_temp_true = (`j', $beta, $p)
201
202             /* 2a- Pooling method */
203             qui regress Y pool
204             mat table_pool = r(table)
205             global beta = table_pool[1,1]          // regression coefficient
206             global se = table_pool[2,1]           // standard error
207             global sd = $sse * sqrt($nb_indiv)   // computation of the standard deviation
208             global p = table_pool[4,1]            // p-value
209             mata : res_temp_pool = (`j', $beta, $sd, $p)
210
211             if (`j'==1){
212                 mata : res_temp_simex = (`j', ., .)
213                 mata : res_temp_rcal = (`j', ., .)
214             }
215             else{
216
217                 /* 2b- SIMEX */
218                 qui simex(Y=) (W : W_1-W`j'), bstrap brep(100) seed(1)
219                 mat tab_simex = e(theta)
220                 global beta = tab_simex[1,2]        //regression coefficient
221                 mat tab_simex = e(V)
222                 global se = sqrt(tab_simex[1,1])    // standard error
223                 global sd = $sse * sqrt($nb_indiv) // computation of the standard deviation
224                 global t = $beta/$se              // student test (hypothesis : beta=0)
225                 global p = 2*ttail(e(df_r),abs($t)) // computation of the p-value
226                 /* note: For large sample size the student distribution could be approximated by a normal distribution,
227                    however for small sample size the normal distribution should be used to compute p-values */
228                 // e(df_r) : degree of freedom
229                 // inverse Student distribution
230                 mata : res_temp_simex = (`j', $beta, $p)
231
232                 /* 2c- Regression Calibration */
233                 qui rcal (Y=) (W : W_1-W`j'), bstrap brep(100) seed(1)
234                 mat tab_rcal = e(b)
235                 global beta = tab_rcal[1,1]        // regression coefficient
236                 mat tab_rcal = e(V)
237                 global se = sqrt(tab_rcal[1,1])    // standard error
238                 global sd = $sse * sqrt($nb_indiv) // computation of the standard deviation
239                 global t = $beta/$se              // student test (hypothesis : beta=0)
240                 global p = 2*ttail(e(df_r),abs($t)) // computation of the p-value
241                 /* note: For large sample size the student distribution could be approximated by a normal distribution,
242                    however for small sample size the normal distribution should be used to compute p-values */
243
244                 mata : res_temp_rcal = (`j', $beta, $p)
245             }
246
247             /* Storage of the results obtained with the different methods in mata matrix */
248             if `i'==1 & `it'==1 {
249                 mata : res_all_true = res_temp_true
250                 mata : res_all_pool = res_temp_pool
251                 mata : res_all_simex = res_temp_simex
252                 mata : res_all_rcal = res_temp_rcal
253             }
254             else {
255                 mata : res_all_true = res_all_true\res_temp_true
256                 mata : res_all_pool = res_all_pool\res_temp_pool
257                 mata : res_all_simex = res_all_simex\res_temp_simex
258                 mata : res_all_rcal = res_all_rcal\res_temp_rcal
259             }
}

```

```

260         }
261     }
262 }
263 clear
264
265 /* -----
266 ****COMPUTATION OF THE RESULTS OBTAINED AFTER RUNNING THE 1000 SIMULATIONS AND STORAGE
267 ****
268 */
269 COMPUTATION OF THE RESULTS OBTAINED AFTER RUNNING THE 1000 SIMULATIONS AND STORAGE
270 ****
271
272 /* Transformation of the matrix into variables */
273
274 // Results obtained with the real exposure
275 getmata (nb_ech all_beta_true all_p_val true) = res_all_true, replace force
276 // Results obtained with the within-subject pooling method
277 getmata (nb_ech all_beta_pool all_sd_pool all_p_val_pool) = res_all_pool, replace force
278 // SIMEX
279 getmata (nb_ech all_beta_simex all_p_val_simex) = res_all_simex, replace force
280 // Regression Calibration
281 getmata (nb_ech all_beta_rcal all_p_val_rcal) = res_all_rcal, replace force
282
283
284 /* 2d- A posteriori disattenuation of the effect estimates obtained with pooling */
285 gen all_beta_corr = all_beta_pool * ( (nb_ech-1)/nb_ech + 1/(nb_ech * $c) )
286 gen all_sd_corr = all_sd_pool * ( (nb_ech-1)/nb_ech + 1/(nb_ech * $c) )
287 gen all_IC_inf_corr = all_beta_corr - 1.96 * all_sd_corr / sqrt($nb_indiv)
288 gen all_IC_sup_corr = all_beta_corr + 1.96 * all_sd_corr / sqrt($nb_indiv)
289
290 /* Results computation for each number of samples 'j' used per participant */
291 local it = 0 /* sample list iterator */
292
293 foreach j in $sample_list{
294     local it = `it' + 1
295     /* Mean and variance */
296     qui sum all_beta_true if nb_ech==`j'
297     global mean_beta_true = r(mean)          // betas true exposure
298     global sd_beta_true = r(sd)
299
300     qui sum all_beta_pool if nb_ech==`j'
301     global mean_beta_pool = r(mean)        // betas pooling
302     global sd_beta_pool = r(sd)
303
304     qui sum all_beta_corr if nb_ech==`j'
305     global mean_beta_corr = r(mean)      /* betas pooling +
306     a posteriori disattenuation */
307     global sd_beta_corr = r(sd)
308
309     if (`j'==1) {
310         global mean_beta_simex = .
311         global mean_beta_rcal = .
312
313         global sd_beta_simex = .
314         global sd_beta_rcal = .
315
316         global puiss_simex = .
317         global puiss_rcal = .
318     }
319     else {
320         qui sum all_beta_simex if nb_ech==`j'
321         global mean_beta_simex = r(mean)    // betas SIMEX
322         global sd_beta_simex = r(sd)
323
324         qui sum all_beta_rcal if nb_ech==`j'
325         global mean_beta_rcal = r(mean)    // betas regression calibration
326         global sd_beta_rcal = r(sd)
327     }
328
329     /* Empirical Confidence Intervals */
330     qui centile (all_beta_pool) if nb_ech==`j', centile (2.5 97.5) // pooling
331     global IC_inf_pool = r(c 1)
332     global IC_sup_pool = r(c 2)
333
334
335     qui centile (all_beta_corr) if nb_ech==`j', centile (2.5 97.5) /* pooling + a
336     posteriori disattenuation */
337     global IC_inf_corr = r(c 1)
338     global IC_sup_corr = r(c 2)
339
340     qui centile (all_beta_simex) if nb_ech==`j', centile (2.5 97.5) // SIMEX
341     global IC_inf_simex = r(c 1)
342     global IC_sup_simex = r(c 2)
343
344     qui centile (all_beta_rcal) if nb_ech==`j', centile (2.5 97.5) // regression calibration
345     global IC_inf_rcal = r(c 1)
346     global IC_sup_rcal = r(c 2)

```

```

347
348     /* Power (percentage of p-value < 0.05) */
349     qui count if all_p_val_true<0.05 & nb_ech=='j'          // true exposure
350     global puiss_true = r(N)/$nb_simul
351
352     qui count if all_p_val_pool<0.05 & nb_ech=='j'          // pooling
353     global puiss_pool = r(N)/$nb_simul
354
355     qui count if all_IC_inf_corr<0 & all_IC_sup_corr>0 & nb_ech=='j' /* pooling +
356     a posteriori disattenuation */
357     global puiss_corr = (${nb_simul}-r(N)) / $nb_simul
358
359     if (`j'>1) {
360         qui count if all_p_val_simex<0.05 & nb_ech=='j' // SIMEX
361         global puiss_simex = r(N)/$nb_simul
362         qui count if all_p_val_rcal<0.05 & nb_ech=='j' // regression calibration
363         global puiss_rcal = r(N)/$nb_simul
364     }
365
366 /* CREATION OF MATRIX RESULT*/
367 mat indiv_beta = `j', $mean_beta_true, $mean_beta_pool, $mean_beta_corr, $mean_beta_simex, $mean_beta_rcal
368 mat indiv_sd = `j', $sd_beta_true, $sd_beta_pool, $sd_beta_corr, $sd_beta_simex, $sd_beta_rcal
369 mat indiv_IC_inf = `j', $IC_inf_pool, $IC_inf_corr, $IC_inf_simex, $IC_inf_rcal
370 mat indiv_IC_sup = `j', $IC_sup_pool, $IC_sup_corr, $IC_sup_simex, $IC_sup_rcal
371 mat indiv_puiss = `j', $puiss_true, $puiss_pool, $puiss_corr, $puiss_simex, $puiss_rcal
372
373 if `it'==1 { // creation of the first line of result tables
374     mat res_beta = indiv_beta // Mean of the estimators
375     mat res_sd = indiv_sd // Standard deviation of the estimators
376     mat res_IC_inf = indiv_IC_inf // Lower bound of the CI of the estimators
377     mat res_IC_sup = indiv_IC_sup // Upper bound of the CI of the estimators
378     mat res_puiss = indiv_puiss // Power of the estimators
379 }
380 else { // add of a line on result tables already existing
381     mat res_beta = res_beta\indiv_beta
382     mat res_sd = res_sd\indiv_sd
383     mat res_IC_inf = res_IC_inf\indiv_IC_inf
384     mat res_IC_sup = res_IC_sup\indiv_IC_sup
385     mat res_puiss = res_puiss\indiv_puiss
386 }
387 }
388
389 /* Rename the name of the matrix column*/
390 matrix colnames res_beta = "p" "True" "Pooling" "Pooling + Disatt" "SIMEX" "Reg-Cal"
391 matrix colnames res_sd = "p" "True" "Pooling" "Pooling + Disatt" "SIMEX" "Reg-Cal"
392 matrix colnames res_IC_inf = "p" "Pooling" "Pooling + Disatt" "SIMEX" "Reg-Cal"
393 matrix colnames res_IC_sup = "p" "Pooling" "Pooling + Disatt" "SIMEX" "Reg-Cal"
394 matrix colnames res_puiss = "p" "True" "Pooling" "Pooling + Disatt" "SIMEX" "Reg-Cal"
395
396 /* -----
397 /* MATRIX TRANSFORMATION IN VARIABLES + RENAME OF THESE VARIABLES*/
398
399 /* Mean of betas*/
400 svmat res_beta
401 rename res_beta1 p
402 rename res_beta2 beta_true
403 rename res_beta3 beta_pool
404 rename res_beta4 beta_corr
405 rename res_beta5 beta_simex
406 rename res_beta6 beta_rcal
407
408 /* Percentages of bias in the effect estimates compared to the real effect */
409 qui gen perc_bias_pool = abs((beta_pool - beta_true) / beta_true) * 100
410 qui gen perc_bias_corr = abs((beta_corr - beta_true) / beta_true) * 100
411 qui gen perc_bias_simex = abs((beta_simex - beta_true) / beta_true) * 100
412 qui gen perc_bias_rcal = abs((beta_rcal - beta_true) / beta_true) * 100
413
414 /* Variance of betas*/
415 svmat res_sd
416 drop res_sd1
417 rename res_sd2 sd_true
418 rename res_sd3 sd_pool
419 rename res_sd4 sd_corr
420 rename res_sd5 sd_simex
421 rename res_sd6 sd_rcal
422
423 /* Confidence interval*/
424 /* Lower bound*/
425 svmat res_IC_inf
426 drop res_IC_inf1
427 rename res_IC_inf2 IC_inf_pool
428 rename res_IC_inf3 IC_inf_corr
429 rename res_IC_inf4 IC_inf_simex
430 rename res_IC_inf5 IC_inf_rcal
431
432 /* Upper bound*/
433 svmat res_IC_sup

```

```

434 drop res_IC_sup1
435 rename res_IC_sup2 IC_sup_pool
436 rename res_IC_sup3 IC_sup_corr
437 rename res_IC_sup4 IC_sup_simex
438 rename res_IC_sup5 IC_sup_rcal
439
440 /* Power*/
441 svmat res_puiss
442 drop res_puiss1
443 rename res_puiss2 puiss_true
444 rename res_puiss3 puiss_pool
445 rename res_puiss4 puiss_corr
446 rename res_puiss5 puiss_simex
447 rename res_puiss6 puiss_rcal
448
449 /* Round of variables */
450 global var_list = "pool corr simex rcal"
451
452 qui replace beta_true = round(beta_true,1)
453 qui replace puiss_true = round(puiss_true,0.01)
454
455 foreach var in $var_list {
456     qui replace beta`var' = round(beta`var',1)
457     qui replace sd`var' = round(sd`var',0.1)
458     qui replace IC_inf`var' = round(IC_inf`var',0.1)
459     qui replace IC_sup`var' = round(IC_sup`var',0.1)
460     qui replace puiss`var' = round(puiss`var',0.01)
461     qui replace perc_bias`var' = round(perc_bias`var',1)
462
463     qui tostring(IC_inf`var'), gen(string_IC_inf`var') force format(%7.0f)
464     qui tostring(IC_sup`var'), gen(string_IC_sup`var') force format(%7.0f)
465     qui gen IC`var' = "[" + string_IC_inf`var' + " ; " + string_IC_sup`var' + "]"
466     if string IC_inf`var' != "."
467     & string_IC_sup`var' != "."
468 }
469
470 /* Back up of data set containing results */
471 save "$results/Correlation 0$c/beta $b1/n $nb_indiv/res_n${nb_indiv}_varUass${var_Uassay}_sim${nb_simul}.dta",
472 replace
473
474 preserve
475     /* Exportation of results on Excel */
476     keep p beta_true puiss_true beta_pool IC_pool puiss_pool perc_bias_pool ///
477         beta_corr IC_corr puiss_corr perc_bias_corr ///
478         beta_simex IC_simex puiss_simex perc_bias_simex ///
479         beta_rcal IC_rcal puiss_rcal perc_bias_rcal
480     export excel using "$results/Correlation 0$c/beta $b1/n
$nb_indiv/tab_corr0${c}_beta${b1}_n${nb_indiv}_varUass${var_Uassay}_sim${nb_simul}.xls", firstrow(variables) replace
481
482 /*
-----*/
483
484     /* GRAPHICS : ARTICLE - FIGURE 1 */
485
486     /* Percentage bias */
487     twoway (line perc_bias_pool p, lcolor(black) lpattern(solid)) ///
488         (line perc_bias_corr p, lcolor(black) lpattern(dot)) ///
489         (line perc_bias_simex p, lcolor(black) lpattern(shorthash)) ///
490         (line perc_bias_rcal p, lcolor(black) lpattern(dash)) if p>=2 & p<=30, ///
491         xtitle("Number of urinary samples per subject", size(medlarge)) xlabel(2 5 10 15 20 25 30) ///
492         ytitle("Bias in the effect estimate (%)", size(medlarge)) legend(label(1 "Pooling method") ///
493         label(2 "Pooling + Disattenuation") label(3 "Simex") label(4 "Regression-Calibration") cols(2) span bmargin(zero) ) ///
494         name(graph_bias, replace) scheme(s2gmanual) graphregion(color(white)) bgcolor(white)
495
496     /* BACK UP OF GRAPHICS */
497     qui graph save graph_bias "$results/Correlation 0$c/beta $b1/n
$nb_indiv/graph_bias_n${nb_indiv}_varUass${var_Uassay}_sim${nb_simul}.gph", replace
498
499
500 }
501
502 timer off 1
503 timer list 1
504
505
506
507
508

```

```

1  ****
2  /* Exposure measurement error in biomarker based studies
3  Codes for the continous outcome, unbalanced desing
4  (Codes for the binary outcome are available upon request) */
5  ****
6
7  -----
8  Creation : 15/04/14 (F.Perrier)
9  Modification : 28/05/15 (F.Perrier)
10 Software used: STATA/SE, version 13 (StataCorp, College Station, TX, USA)
11
12 Package that need to be install before running the program :
13 net from http://www.stata.com/mirror
14 net install mmrro // package needed to use simex and regression calibration fonctions
15
16 *Note: This program does not take into account between-assay error
17 -----
18
19 cap program drop _all
20 clear all
21 *set matsize 6000
22 set more off
23
24 timer clear
25 timer on 1
26
27 /* Definitions of the paths to folders in which we will store the simulated datasets and the
28 results of the simulation */
29 ****
30 cd "D:\Erreurs de mesure"
31 global simulation = "Simulations\nb_ech_diff_lin 22-10-14" /*dataset storage */
32 global results = "Resultats\nb_ech_diff_lin 22-10-14" /*result storage */
33
34 /* Definition of the parameters used in the simulation */
35 ****
36 global b0 = 14900      /* Intercept of the linear regression model */
37 global b1 = -100       /* Effect of the real exposure (X) on the outcome (Y) */
38
39 global nb_indiv = 3000 /* Number of subjects in the simulated population */
40 global nb_simul = 1000 /* Number of repetitions / simulations */
41 global nb_ech_tot = 9000 /* Number total of biospecimens */
42 global corr_list = "0.6 0.2" /* Intraclass correlation coefficient observed between repeated
43 measurements of exposure */
44
45 /*Definition of the different scenarios to simulate */
46 ****
47 /* the following study desing will be simulated
48 Scenario a : 600 subjects with 1 sample / 1500 with 2 samples / 900 with 6 samples
49 Scenario b : 600 subjects with 1 sample / 1800 avec 3 samples / 600 with 5 samples
50 Scenario c : 900 subjects with 1 sample / 1200 with 3 samples / 900 with 5 samples
51 Scenario d : 1200 subjects with 1 sample / 1200 with 2 samples / 600 with 9 samples
52 */
53 global nb_sce = 4      /* Number of different designs to test */
54 mat z_1 = 1\1\1\1      /* Number of biospecimens to simulate for the group 1 for each design */
55 mat z_2 = 2\3\3\2      /* Number of biospecimens to simulate for the group 2 for each design */
56 mat z_3 = 6\5\5\9      /* Number of biospecimens to simulate for the group 3 for each design */
57 mat p_1 = 0.2\0.2\0.3\0.4 /* Percentage of the population belonging to group 1 */
58 mat p_2 = 0.7\0.8\0.7\0.8 /* Cumulative percentage of the population belonging to group 1 or 2*/
59
60 -----
61 /* Folder creation (needed datasets and results storage)
62 * mkdir "...."
63 /*
64 * datasets
65 ****
66 forvalue k = 1/$nb_sce{
67     mkdir "$simulation\Scenario_`k'"
68 }
69
70 foreach c in $corr_list {
71     forvalue k = 1/$nb_sce{
72         mkdir "$simulation\Scenario_`k'\ICC_`c'"
73     }
74 }
75 */
76 -----
77 ****
78 DATA SET SIMULATION /* We assumed a lack of between-assay error */
79 In this section we simulated the datasets
80 Notations:
81 X: Real exposure
82 Uwithin: within-subject error
83 W: Exposure measure with error
84 Y: Continuous health outcome
85 ****
86 set obs $nb_indiv
87

```

```

88  set seed 15042014
89
90 /*-----*/
91 local it = 0 /* simulations iterator */
92
93 forvalue k = 1/$nb_sce {
94
95     global z_1 = z_1[`k',1] /* Number of samples to simulate for the group 1 for the kth scenario */
96     global z_2 = z_2[`k',1] /* Number of samples to simulate for the group 2 for the kth scenario */
97     global z_3 = z_3[`k',1] /* Number of samples to simulate for the group 3 for the kth scenario */
98     global p_1 = p_1[`k',1] /* Percentage of the population belonging to group 1 for the kth
99 scenario */
100    global p_2 = p_2[`k',1] /* Cumulative percentage of the population belonging to groups 1 or
101 2 for the kth scenario */
102
103    di in red " Scenario `k' : - " $p_1*100 "% of the population with " $z_1 " sample(s) "
104    if ($p_1!=1 & $z_2!=0) {
105        di in red " - " ($p_2-$p_1)*100 "% of the population with " $z_2 " samples "
106    }
107    if ($p_2!=1 & $z_3!=0) {
108        di in red " - " (1-$p_2)*100 "% of the population with " $z_3 " samples "
109    }
110
111    foreach corr in $corr_list {
112        global c = `corr'
113        di in green " ICC 0" $c
114
115        forvalue i = 1/$nb_simul {
116            local it = `it' + 1
117            if (mod(`i',$nb_simul/10)==0) {
118                di in yellow " `i' /($nb_simul/10) /10 of data are simulated "
119            }
120
121            qui set obs $nb_indiv
122            gen id = _n
123
124            /* Simulation of X (True exposure) */
125            global fixed mean_X = 0 // mean
126            global fixed var_X = 1 // variance
127            mat m_X = $fixed_mean_X
128            mat sd_X = sqrt($fixed_var_X)
129            qui drawnorm X, mean(m_X) sds(sd_X) n($nb_indiv)
130
131            /* Simulation of Uwithin (within-subject error)
132             / We assumed a lack of between-assay error */
133            qui sum X
134            global var_X_true = r(Var)
135            global var_Uwithin_true = $var_X_true * (1/$c - 1) // fixed variance of error
136            mat m_Uwithin = J(1,$z_1,0)
137            mat sd_Uwithin = J(1,$z_1,sqrt($var_Uwithin_true))
138            global temp = $z_1 + 1
139
140            if $z_1 == 1 { /* Group 1 */
141                qui drawnorm Uwithin_1, mean(m_Uwithin) sds(sd_Uwithin) n($nb_indiv)
142            }
143            else{
144                qui drawnorm Uwithin_1-Uwithin_`z_1 , mean(m_Uwithin) sds(sd_Uwithin) n($nb_indiv)
145            }
146
147
148            if $z_2!=0 { /* Group 2 */
149                forvalues j = $temp/$z_2 {
150                    qui gen Uwithin_`j' = rnormal(0,sqrt($var_Uwithin_true)) if id>$p_1*$nb_indiv
151                }
152            }
153
154            global temp = $z_2 + 1
155            if $z_3!=0 { /* Group 3 */
156                forvalues j = $temp/$z_3 {
157                    qui gen Uwithin_`j' = rnormal(0,sqrt($var_Uwithin_true)) if id>($p_2)*$nb_indiv
158                }
159            }
160
161            /* Simulation of W (exposures measured with error) */
162            global z_max = max($z_1, $z_2, $z_3)
163            forvalues j = 1/$z_max {
164                qui gen W_`j' = X + Uwithin_`j'
165            }
166
167            /* Simulation of the continuous health outcome */
168            qui drawnorm E, mean(0) sds(1650) n($nb_indiv) /* Simulation of the model error,
169             sds(1650) is the SD observed in the French mother child cohort EDEN, for the
170             weight at 3 years */
171
172            qui gen Y = $b0 + $b1*X + E
173
174            if (max($p_1, $p_2) == $p_1 & $p_1 != 1) {

```

```

175          qui sample 50
176          qui replace id = _n
177      }
178
179      qui save "${simulation}\Scenario_`k'\ICC_0${c}\simul`i'_beta${b1}_n${nb_indiv}", replace
180
181
182  /*************************************************************************/
183 ANALYSES
184 In this section we study the associations between the outcome (Y) and the real exposure (1) or the
185 error prone variable using different approaches: Pooling (2), Regression Calibration (3) and Simex (4)
186 /*************************************************************************/
187
188
189 /* 1- Calculation and storage of beta true, the beta coefficient we obtained when we used
190 the real exposure X */
191 qui regress Y X
192 mat table_true = r(table)
193 global beta = table_true[1,1]      //regression coefficient
194 global se = table_true[2,1]      //standard error
195 global sd = $se * sqrt($nb_indiv) //standard deviation
196 global puiss = table_true[4,1]    //p-value
197 mata : res_temp_true = (`k', $c, $beta, $sd, $puiss)
198
199
200 /* 2a- Pooling */
201 qui egen W_pool = rowmean(W_1-W_$z_max) /* computation of the mean of the repeated measures
202 for each individual */
203 qui gen weight = $z_1 // definition of a weight for each subject
204 if (max($z_1, $z_2) != $z_1) {
205     qui replace weight = $z_2 if id > ($p_1)*$nb_indiv & id <= ($p_2)*$nb_indiv
206     qui replace weight = $z_3 if id > ($p_2)*$nb_indiv
207 }
208 qui regress Y W_pool [aw=weight]
209
210 mat table_pool = r(table)
211 global beta = table_pool[1,1]      //regression coefficient
212 global se = table_pool[2,1]      //standard error
213 global sd = $se * sqrt($nb_indiv) //standard deviation
214 global puiss = table_pool[4,1]    //p-value
215 mata : res_temp_pool = (`k', $c, $beta, $sd, $puiss)
216
217
218 /* 3- SIMEX */
219 qui simex(Y=) (W : W_1-W_$z_max), bstrap brep(100) seed(1)
220
221 mat tab_simex = e(theta)
222 global beta = tab_simex[1,2]      //regression coefficient
223 mat tab_simex = e(V)
224 global se = sqrt(tab_simex[1,1]) //standard error
225 global sd = $se * sqrt($nb_indiv) //standard deviation
226 global t = $beta/$se            //student test (hypothesis : beta=0)
227 global puiss = 2*ttail(e(df_r),abs($t)) //computation of the p-value
228 // e(df_r) : degree of freedom
229 // inverse distribution of Student
230 /* note: For large sample size the student distribution could be approximated by a normal
231 distribution, however for small sample size the normal distribution should be used to
232 compute p-values */
233 mata : res_temp_simex = (`k', $c, $beta, $sd, $puiss)
234
235
236 /* 4- Regression-Calibration */
237 qui rcal (Y=) (W : W_1-W_$z_max), bstrap brep(100) seed(1)
238
239 mat tab_rcal = e(b)
240 global beta = tab_rcal[1,1]      //regression coefficient
241 mat tab_rcal = e(V)
242 global se = sqrt(tab_rcal[1,1]) //standard error
243 global sd = $se * sqrt($nb_indiv) //standard error
244 global t = $beta/$se            //student test (hypothesis : beta=0)
245 global puiss = 2*ttail(e(df_r),abs($t)) //computation of the p-value
246 /* note: For large sample size the student distribution could be approximated by a normal
247 distribution, however for small sample size the normal distribution should be used to
248 compute p-values */
249 mata : res_temp_rcal = (`k', $c, $beta, $sd, $puiss )
250
251
252 /* Storage of the results obtained with the different methods in mata matrix */
253 if `it'==1 {
254     mata : res_all_true = res_temp_true
255     mata : res_all_pool = res_temp_pool
256     mata : res_all_simex = res_temp_simex
257     mata : res_all_rcal = res_temp_rcal
258 }
259 else {
260     mata : res_all_true = res_all_true\res_temp_true
261     mata : res_all_pool = res_all_pool\res_temp_pool
262     mata : res_all_simex = res_all_simex\res_temp_simex
263     mata : res_all_rcal = res_all_rcal\res_temp_rcal
264 }
```

```

262         drop *
263     }
264   }
265 }
266
267 /*-----*/
268
269 /* **** COMPUTATION OF THE RESULTS OBTAINED AFTER RUNNING THE 1000 SIMULATIONS AND STORAGE ****
270 COMPUTATION OF THE RESULTS OBTAINED AFTER RUNNING THE 1000 SIMULATIONS AND STORAGE
271 **** */
272
273 /* Transformation of the matrix into variables */
274
275 //Beta true
276 getmata (all_num_sce all_ICC all_beta_true all_sd_true all_p_val_true) = res_all_true, replace force
277 //Pooling method
278 getmata (all_num_sce all_ICC all_beta_pool all_sd_pool all_p_val_pool) = res_all_pool, replace force
279 //SIMEX
280 getmata (all_num_sce all_ICC all_beta_simex all_sd_simex all_p_val_simex) = res_all_simex, replace force
281 //Regression Calibration
282 getmata (all_num_sce all_ICC all_beta_rcal all_sd_rcal all_p_val_rcal) = res_all_rcal, replace force
283
284 /*-----*/
285 /* Results computation for each number of samples 'j' used per participant (n = 1000 simulations) */
286 local it = 0
287
288 forvalue k = 1/$nb_sce {
289   global z_1 = z_1[`k',1] /* Number of samples simulated for the group 1 for the kth scenario */
290   global z_2 = z_2[`k',1] /* Number of samples simulated for the group 2 for the kth scenario */
291   global z_3 = z_3[`k',1] /* Number of samples simulated for the group 3 for the kth scenario */
292   global p_1 = p_1[`k',1] /* Percentage of the population belonging to group 1 for the kth
293 scenario */
294   global p_2 = p_2[`k',1] /* Cumulative percentage of the population belonging to groups 1 or 2
295 for the kth scenario */
296
297   foreach corr in $corr_list {
298     global c = `corr'
299     local it = `it' + 1
300
301     /* Mean and variance */
302     qui sum all_beta_true if all_num_sce==`k' & all_ICC==$c
303     global mean_beta_true = r(mean)          //betas true
304     global sd_beta_true = r(sd)
305
306     qui sum all_beta_pool if all_num_sce==`k' & all_ICC==$c
307     global mean_beta_pool = r(mean)          //pooling method
308     global sd_beta_pool = r(sd)
309
310     qui sum all_beta_simex if all_num_sce==`k' & all_ICC==$c
311     global mean_beta_simex = r(mean)          //Simex
312     global sd_beta_simex = r(sd)
313
314     qui sum all_beta_rcal if all_num_sce==`k' & all_ICC==$c
315     global mean_beta_rcal = r(mean)          //Regression calibration
316     global sd_beta_rcal = r(sd)
317
318     /* Confidence Interval of estimators*/
319     qui centile (all_beta_pool) if all_num_sce==`k' & all_ICC==$c, centile (2.5 97.5)
320     global IC_inf_pool = r(c_1)    //pooling method
321     global IC_sup_pool = r(c_2)
322
323     qui centile (all_beta_simex) if all_num_sce==`k' & all_ICC==$c, centile (2.5 97.5)
324     global IC_inf_simex = r(c_1)  //Simex
325     global IC_sup_simex = r(c_2)
326
327     qui centile (all_beta_rcal) if all_num_sce==`k' & all_ICC==$c, centile (2.5 97.5)
328     global IC_inf_rcal = r(c_1)  //Regression calibration
329
330     global IC_sup_rcal = r(c_2)
331
332     /* Power (percentage of p-value upper than 0.05 => acceptance percentage of H0 (beta=0) )*/
333     qui count if all_p_val_true<0.05 & all_num_sce==`k' & all_ICC==$c      //true exposure
334     global puiss_true = r(N)/$nb_simul
335
336     qui count if all_p_val_pool<0.05 & all_num_sce==`k' & all_ICC==$c      //pooling method
337     global puiss_pool = r(N)/$nb_simul
338
339     qui count if all_p_val_simex<0.05 & all_num_sce==`k' & all_ICC==$c      //Simex
340     global puiss_simex = r(N)/$nb_simul
341
342     qui count if all_p_val_rcal<0.05 & all_num_sce==`k' & all_ICC==$c      //Regression calibration
343     global puiss_rcal = r(N)/$nb_simul
344
345     /* CREATION OF MATRIX RESULT*/
346     mat indiv_beta = `k', $c, $mean_beta_true, $mean_beta_pool, $mean_beta_simex, $mean_beta_rcal
347     mat indiv_sd = `k', $c, $sd_beta_true, $sd_beta_pool, $sd_beta_simex, $sd_beta_rcal
348     mat indiv_IC_inf = `k', $c, $IC_inf_pool, $IC_inf_simex, $IC_inf_rcal

```

```

349     mat indiv_IC_sup = `k', $c, $IC_sup_pool, $IC_sup_simex, $IC_sup_rcal
350     mat indiv_puiss = `k', $c, $puiss_true, $puiss_pool, $puiss_simex, $puiss_rcal
351
352     if `it'==1 { //creation of the first line of result tables
353         mat res_beta = indiv_beta //Mean of the estimators
354         mat res_sd = indiv_sd //Standard deviation of the estimators
355         mat res_IC_inf = indiv_IC_inf //Lower bound of the confidence interval of the estimators
356         mat res_IC_sup = indiv_IC_sup //Upper bound of the confidence interval of the estimators
357         mat res_puiss = indiv_puiss //Estimators power
358     }
359     else { //add of a line on result tables already existing
360         mat res_beta = res_beta\indiv_beta
361         mat res_sd = res_sd\indiv_sd
362         mat res_IC_inf = res_IC_inf\indiv_IC_inf
363         mat res_IC_sup = res_IC_sup\indiv_IC_sup
364         mat res_puiss = res_puiss\indiv_puiss
365     }
366 }
367 }
368
369 /* Rename the name of the matrix column */
370 matrix colnames res_beta = "Scenario" "ICC" "True" "Pooling" "SIMEX" "Reg-Cal"
371 matrix colnames res_sd = "Scenario" "ICC" "True" "Pooling" "SIMEX" "Reg-Cal"
372 matrix colnames res_IC_inf = "Scenario" "ICC" "Pooling" "SIMEX" "Reg-Cal"
373 matrix colnames res_IC_sup = "Scenario" "ICC" "Pooling" "SIMEX" "Reg-Cal"
374 matrix colnames res_puiss = "Scenario" "ICC" "True" "Pooling" "SIMEX" "Reg-Cal"
375
376 /*-----*/
377 /* MATRIX TRANSFORMATION IN VARIABLES + RENAME OF THESE VARIABLES */
378
379 /* Mean of beta */
380 svmat res_beta
381 rename res_beta1 num_sce
382 rename res_beta2 ICC
383 rename res_beta3 beta_true
384 rename res_beta4 beta_pool
385 rename res_beta5 beta_simex
386 rename res_beta6 beta_rcal
387
388 /* Difference between beta mean and the true value of beta */
389 qui gen perc_bias_pool = abs((beta_pool - beta_true) / beta_true) * 100
390 qui gen perc_bias_simex = abs((beta_simex - beta_true) / beta_true) * 100
391 qui gen perc_bias_rcal = abs((beta_rcal - beta_true) / beta_true) * 100
392
393 /* Variance of beta */
394 svmat res_sd
395 drop res_sd1
396 drop res_sd2
397 rename res_sd3 sd_true
398 rename res_sd4 sd_pool
399 rename res_sd5 sd_simex
400 rename res_sd6 sd_rcal
401
402 /* Confidence interval */
403 /* Lower bound */
404 svmat res_IC_inf
405 drop res_IC_Inf1
406 drop res_IC_inf2
407 rename res_IC_inf3 IC_inf_pool
408 rename res_IC_inf4 IC_inf_simex
409 rename res_IC_inf5 IC_inf_rcal
410 /* Upper bound */
411 svmat res_IC_sup
412 drop res_IC_sup1
413 drop res_IC_sup2
414 rename res_IC_sup3 IC_sup_pool
415 rename res_IC_sup4 IC_sup_simex
416 rename res_IC_sup5 IC_sup_rcal
417
418 /* Power */
419 svmat res_puiss
420 drop res_puiss1
421 drop res_puiss2
422 rename res_puiss3 puiss_true
423 rename res_puiss4 puiss_pool
424 rename res_puiss5 puiss_simex
425 rename res_puiss6 puiss_rcal
426
427 /* Round of variables */
428 global var_list = "pool simex rcal"
429
430 qui replace beta_true = round(beta_true,1)
431
432 foreach var in $var_list {
433     qui replace beta_`var' = round(beta_`var',1)
434     qui replace sd_`var' = round(sd_`var',0.1)
435     qui replace IC_inf_`var' = round(IC_inf_`var',0.1)

```

```

436     qui replace IC_sup `var' = round(IC_sup `var',0.1)
437     qui replace puiss `var' = round(puiss `var',0.01)
438     qui replace perc_bias_`var' = round(perc_bias_`var',1)
439
440     qui tostring(IC_inf `var'), gen(string IC_inf `var') force format(%7.0f)
441     qui tostring(IC_sup `var'), gen(string IC_sup `var') force format(%7.0f)
442     qui gen IC_`var' = "[" + string_IC_inf `var' + " ; " + string_IC_sup `var' + "]"
443         if string_IC_inf `var' != "." & string_IC_sup `var' != "."
444
445
446 /* Back up of data set containing results */
447 save "${results}\res_beta${b1}_n${nb_indiv}", replace
448
449 /* Exportation of results on Excel */
450 export excel num_sce ICC beta_true beta_pool IC_pool puiss_pool perc_bias_pool ///
451     beta_simex IC_simex puiss_simex perc_bias_simex ///
452     beta_rcal IC_rcal puiss_rcal perc_bias_rcal ///
453     using "${results}\tab_res_beta${b1}_n${nb_indiv}.xls", firstrow(variables) replace
454
455
456 /*-----*/
457 /* GRAPHICS */
458
459 /* Percent bias */
460 graph bar perc_bias_pool perc_bias_simex perc_bias_rcal, over(ICC, relabel(1 "0.2" 2 "0.6")) ///
461     over(num_sce, relabel(1 "Scenario 1" 2 "Scenario 2" 3 "Scenario 3" 4 "Scenario 4")) bargap(-30) ///
462     bar(1, color(blue)) bar(2, color(red)) bar(3, color(green)) ///
463     ytitle("Bias in the effect estimate (%)", size(medlarge)) ///
464     legend(label(1 "Pooling method") label(2 "Simex") label(3 "Regression-Calibration") cols(3) span ///
465         bmargin(zero) ) name(graph_bias, replace)
466
467 /* Power */
468 graph bar (mean) puiss_pool puiss_simex puiss_rcal, over(ICC, relabel(1 "0.2" 2 "0.6")) ///
469     over(num_sce, relabel(1 "Scenario 1" 2 "Scenario 2" 3 "Scenario 3" 4 "Scenario 4")) bargap(-30) ///
470     bar(1, color(blue)) bar(2, color(red)) bar(3, color(green)) ///
471     exclude0 ylabel(0.4(0.1)1) ///
472     ytitle("Power", size(medlarge)) ///
473     legend(label(1 "Pooling method") label(2 "Simex") label(3 "Regression-Calibration") cols(3) span ///
474         bmargin(zero) ) name(graph_puiss, replace)
475
476 timer off 1
477 timer list
478
479 /*-----*/
480
481
482

```