**SDC Table 1**. PCR assays sensitivity

|  |  |  |  |
| --- | --- | --- | --- |
| **Cell line** | **TRG-2T-2F**  | **TRG-1T-2F** | **IVS-1T-1F** |
| **HSB-2** | 1% | 5% | 5% |
| **Jurkat** | 1% | 5% | 5% |
| **RPMI 8402** | 1% | 1% | 5% |
| **CCRF-CEM** | 10% | 10% | 5% |
| **HPB-ALL** | 1% | 1% | 1% |
| **MOLT-3** | 5% | 1% | 1% |
| **SUDHL-1** | 1% | 1% | 1% |

**SDC Table 2.** Outlier PCR assay for cases with a discrepancy regarding conclusion

|  |  |
| --- | --- |
|  |  |
| Case | **Disease category** | **Discrepancy** | **Outlier assay** |
| FR-211 | PTCL | clonal *vs* oligoclonal | TRG-2T-2F (oligoclonal) |
| ES-195 | PTCL | clonal *vs* oligoclonal | TRG-2T-2F (oligoclonal) |
| DE-202 | LGL | clonal *vs* oligoclonal | TRG-2T-2F (oligoclonal) |
| FR-204 | ALCL | minor clonal *vs* oligoclonal | TRG-2T-2F (oligoclonal) |
| DE-195 | LGL | minor clonal *vs* polyclonal | TRG-1T-2F (minor clone) |
| FR-245 | AITL | minor clonal *vs* polyclonal | TRG-1T-2F (polyclonal) |
| DE-084 | AITL | minor clonal *vs* polyclonal | TRG-1T-2F (polyclonal) |
| DE-086 | AITL | minor clonal *vs* polyclonal | TRG-1T-2F (polyclonal) |
| FR-207 | AITL | clonal *vs* oligoclonal | TRG-1T-2F (clonal) |
| DE-258 | PTCL | clonal *vs* oligoclonal | TRG-1T-2F (clonal) |
| ES-208 | PTCL | clonal *vs* oligoclonal | TRG-1T-2F (clonal) |
| FR-179 | PTCL | clonal *vs* oligoclonal | TRG-1T-2F (oligoclonal) |
| DE-217 | PTCL | clonal *vs* oligoclonal | TRG-1T-2F (oligoclonal) |
| DE-002 | PTCL | clonal *vs* polyclonal | IVS-1T-1F (polyclonal) |
| DE-193 | LGL | minor clonal *vs* polyclonal | IVS-1T-1F (polyclonal) |
| DE-196 | LGL | minor clonal *vs* polyclonal | IVS-1T-1F (polyclonal) |
| 4F | Reactive | minor clonal *vs* polyclonal | IVS-1T-1F (polyclonal) |
| DE-188 | LGL | clonal *vs* oligoclonal | IVS-1T-1F (clonal) |
| NL-254 | T-PLL | clonal *vs* oligoclonal | IVS-1T-1F (clonal) |
| DE-256 | PTCL | clonal *vs* oligoclonal | IVS-1T-1F (oligoclonal) |
| FR-041 | AITL | minor clonal *vs* oligoclonal | IVS-1T-1F (minor clone) |
| DE-189 | LGL | minor clonal *vs* oligoclonal | IVS-1T-1F (minor clone) |
| DE-203 | LGL | minor clonal *vs* oligoclonal | IVS-1T-1F (minor clone) |

**SDC Table 3.** Characteristics of the 19 cases with TRGJP rearrangement

|  |  |  |  |
| --- | --- | --- | --- |
| **Case** | **Tissue type** | **TRGJP rearrangement** | **Clinical context**  |
| TRGJP-1 | Blood | Canonical | *T cell lymphoma in a 50yo woman* |
| TRGJP-2 | Blood | Canonical | *Chronic pleural effusion in a 76yo man containing a suspicious T cell population population* |
| TRGJP-3 | Blood | Canonical | *LGL population in a 57yo man* |
| TRGJP-4 | Blood | Canonical | *Suspicion of T cell lymphoma in a 41yo man with macrophageactivation syndrome* |
| TRGJP-5 | Blood | Canonical | *Atypical gamma-delta population in a 75yo woman*  |
| TRGJP-6 | Blood | Canonical | *Suspicion of cutaneous lymphoma in a 58yo man* |
| TRGJP-7 | Blood | Canonical | *Lymphocytosis with phenotypically abnormal T cells in a 70yo woman* |
| TRGJP-8 | Bone marrow | Canonical | *BCP-ALL in a 1yo child* |
| TRGJP-9 | Blood | Canonical | *Suspicion of T-LGLin a 60yo woman* |
| TRGJP-10 | Blood | Canonical | *Eosinophilia in a 76yo man* |
| TRGJP-11 | Blood | Canonical | *Chronic neutropenia in a 38yo man* |
| TRGJP-12 | Blood | Canonical | *Hepatosplenic lymphoma in a 22yo man, post renal transplant* |
| TRGJP-13 | Blood | Pathological | *Follow up of circulating gamma-delta T cells in a 15 yo girl* |
| TRGJP-14 | Blood | Pathological | *Gamma-delta T cell population in a 46yo HIV-positive man* |
| TRGJP-15 | Blood | Pathological | *B cell lymphoma in a 9yo girl, IgH and IgK clonal* |
| TRGJP-16 | Bone marrow | Pathological | *BCP-ALL in a 3yo boy* |
| TRGJP-17 | Bone marrow | Pathological | *BCP-ALL in a 60yo man* |
| TRGJP-18 | Bone marrow | Pathological | *T-Lymphoblastic lymphoma in a 5yo boy* |
| TRGJP-19 | Blood | Pathological | *Sezary syndrome in an 83yo woman* |

BCP-ALL : B-cell Precursor Acute Lymphoblastic Leukemia

**Correction to SDC Table 4**

Correction to HemaSphere3(3):e255, June 2019. doi: [10.1097/HS9.0000000000000255](http://dx.doi.org/10.1097/HS9.0000000000000255); published online: 1 June 2019.

Following the publication of this article the authors noted that one of the TRG primers (TRGJ1/J2) depicted in Supplemental Digital Content Table 4, appears in reverse orientation (3’ to 5’). The Table has now been corrected and the TRGJ1/J2 primer is therefore shown in the proper 5’ to 3’ orientation.

The authors wish to apologize for any inconvenience it may have caused.

**SDC Table 4.** Nucleotide sequence of the EuroClonality primers and their location on the TRGV and TRGJ genes compared to the BIOMED-2 primers. See reference 18 for BIOMED-2 primer sequences.

|  |  |  |
| --- | --- | --- |
| Primers | Sequence | Gene Location compared to BIOMED-2 primers |
| **TRGV1f** | GGTTGTGTTGGAATCAGGAGTCA | 46 nucleotides downstream |
| **TRGV9** | CGGCACTGTCAGAAAGGAATC | unchanged |
| **TRGV10** | AGCATGGGTAAGACAAGCAA | unchanged |
| **TRGV11** | TTGCTCAGGTGGGAAGACTA | 56 nucleotides upstream |
| **TRGJ1/J2** | GTGTTGTTCCACTGCCAAAGAG | unchanged |
| **TRGJP1/JP2** | AGTTACTATGAGCYTAGTCCCTT | 10 nucleotides downstream |
| *- Optional -*  |
| **TRGJP** | GAAAACTTACCTGTAATGATAAGCTTT | Not included |

|  |  |  |
| --- | --- | --- |
| **Phenomenon** | **Pitfall(s)** | **Potential solution** |
| 1R in expected biallelic sample or lack of R in an expected clonal sample | - R using a TRGJP gene | - Use an assay with a TRGJP primer |
| - dNTP source | - Change dNTP source (see Material & Methods)  |
| - In case of lack of R in an expected clonal sample, confirm clonality with TRB locus analysis |
| 1R in expected biallelic sample | Superposition of two peaks | Does not change clonality status, but consider using an assay with wider distribution of rearrangements, such as the TRG-1T, as described here. |
| 2R with different intensities, difficult to discriminate biallelic from biclonal unless more than 2 clonal peaks. | Unbalanced analyzer settings  | Use the same fluorochrome to label the primers. Probably bi-allelic if both clonal peak intensities are comparable. Probably biclonal if more than 2 clear clonal peaks or two peaks with different intensities.  |
| R in an expected polyclonal sample  | Amplification of canonical TRGV9-JP rearrangement | Use an assay without a TRGJP primer or label it distinctly and possibly place these products outside the normal size distribution, as in Derrieux C *et al*. , J. Mol. Diagn. 2019;21(1):111-122. |

**SDC Table 5**. Troubleshooting of TRG analysis.

*R : rearrangement*

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