

2 **Respiratory microbiota predicts clinical disease course of acute otorrhea in**  
3 **children with tympanostomy tubes**

4 Wing Ho Man, Thijs M.A. van Dongen, Roderick P. Venekamp, Vincent G. Pluimakers, Mei Ling  
5 J.N. Chu, Marlies A. van Houten, Elisabeth A.M. Sanders, Anne G.M. Schilder, Debby Bogaert

6 *Supplemental methods*..... 2

7     16S rRNA Gene Amplification and Sequencing ..... 2

8     Bioinformatics Analysis ..... 2

9 *eTable 1. Characteristics of the study population at baseline*..... 4

10 *eTable 2. Qualitative agreement between matched pairs of NP and TTO samples on OTU level*. .... 5

11 *eFigure 1. Culture results confirm the taxonomic annotation of the corresponding OTU's*. .... 6

12 *eFigure 2. Flow chart participants and samples*..... 7

13 *eFigure 3. Rarefaction curves on raw count data*. .... 8

14 *eFigure 4.  $\alpha$ -Diversity* ..... 9

15 *eFigure 5. Similarity of paired NP and TTO samples does not vary with clinical variables*. .... 10

16 *References* ..... 12

## 18 **Supplemental methods**

### 19 *16S rRNA Gene Amplification and Sequencing*

20 Bacterial DNA was isolated from samples and quantified as previously described.<sup>1,2</sup> In short, an  
21 aliquot of 200µl of each sample was added to 650µl lysis buffer with 0.1 mm zirconium beads and  
22 550µl phenol. All samples were mechanically lysed with a bead beater procedure. Amplification of the  
23 V4 hypervariable region of the 16S rRNA gene was performed using barcoded universal primer pair  
24 533F/806R. Amplicons were quantified by PicoGreen (ThermoFisher) and pooled in equimolar  
25 amounts. Amplicon pools of samples and controls were sequenced using the Illumina MiSeq platform  
26 (San Diego, CA, USA).

### 27 *Bioinformatics Analysis*

28 Raw sequences were trimmed using an adaptive, window-based trimming algorithm (Sickle, Q>20,  
29 length threshold of 150 nucleotides).<sup>3</sup> We aimed to further reduce the number of sequence errors in the  
30 reads by applying an error correction algorithm (BayesHammer, SPAdes genome assembler toolkit).<sup>4</sup>  
31 Forward and reverse reads were then assembled into contigs using PANDAseq.<sup>5</sup> Merged reads were  
32 demultiplexed using QIIME v1.9.<sup>6</sup> After removal of singleton sequences, we removed chimeras using  
33 both *de novo* and reference (against Gold database) chimera identification (UCHIME algorithm in  
34 VSEARCH).<sup>7,8</sup> VSEARCH abundance-based greedy clustering was used to pick OTUs at a 97%  
35 identity threshold.<sup>9</sup> Taxonomic annotation was executed using the RDP-II naïve Bayesian classifier on  
36 SILVA v119 training set.<sup>10</sup> After aligning the node representative sequences to the Silva v119 core  
37 alignment database using the PyNAST method,<sup>11</sup> a rooted phylogenetic tree was calculated using  
38 FastTree.<sup>12</sup> We generated an abundance-filtered dataset by including only those OTUs that were  
39 present at or above a confident level of detection (0.1% relative abundance) in at least 2 samples,  
40 retaining 138 OTUs in total.<sup>13</sup> To avoid OTUs with identical annotations, we refer to OTUs using their  
41 taxonomical annotations combined with a rank number based on the abundance of each given OTU.  
42 The raw OTU-counts table was used for calculations of  $\alpha$ -diversity and analyses using the  
43 *metagenomeSeq* package.<sup>14</sup> The OTU-proportions table was used for all other downstream analyses,  
44 including hierarchical clustering and random forest modelling. Moreover, the Bray-Curtis

45 (dis)similarity metric was consistently used to express ecological distance ( $\beta$ -diversity) in all analyses  
46 because it includes proportional abundance information and excludes joint-absence information, and  
47 thereby yields useful insights into the specific structure of our data.<sup>15</sup>

**eTable 1. Characteristics of the study population at baseline.**

	Overall (n=94)
Boys, n (%)	57 (60.6)
Mean age, yrs (SD)	3.38 (1.41)
Indication for tube insertion, n (%)	
Otitis media with effusion	43 (45.7)
Acute otitis media	32 (34)
Both	19 (20.2)
Mean duration of otorrhea in days before enrolment (SD)	2.70 (1.78)
Vaccinated, n (%)	
Received PCV7	73 (78.5)
Antibiotics in previous 14 days, n (%)	
Eardrops	0 (0)
Oral	0 (0)
Mean number of tympanostomy tube insertions (SD)	1.24 (0.54)
Mean number of siblings (SD)	1.27 (0.59)
Day care or school, n (%)	
Yes, day care	54 (57.4)
Yes, school	31 (33)
No	9 (9.6)
Breastfed, n (%)	68 (72.3)
Household smoking, n (%)	12 (12.9)

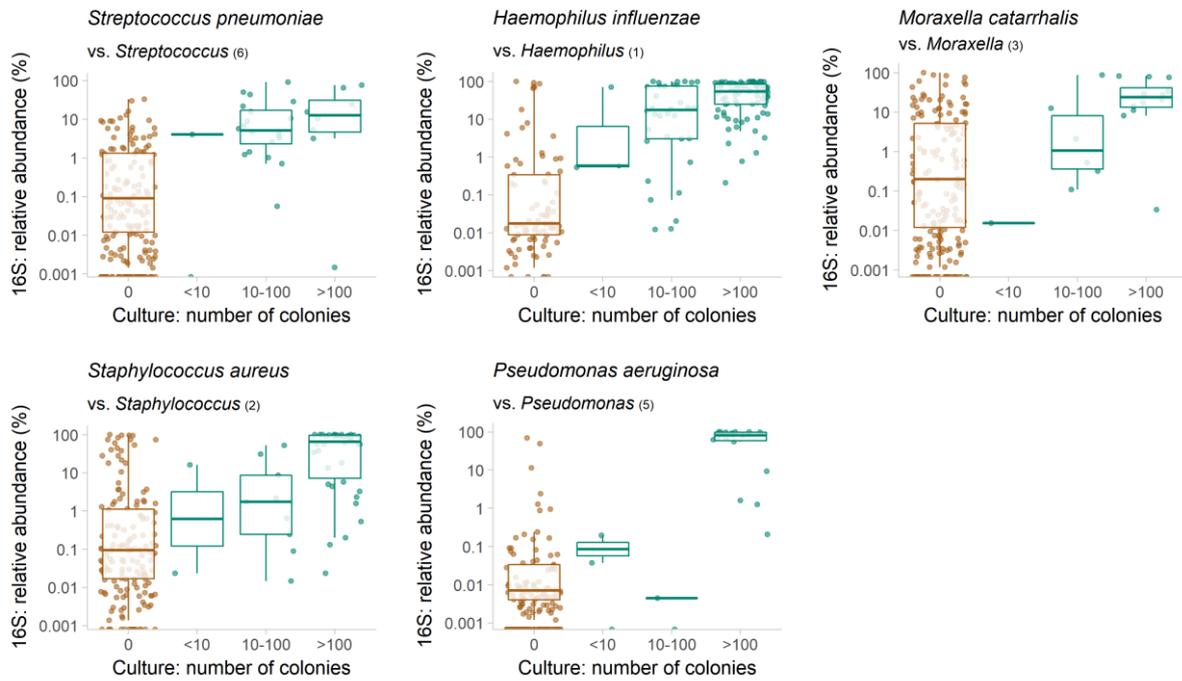
50 **eTable 2. Qualitative agreement between matched pairs of NP and TTO samples on OTU level.**

51 We calculated the overall positive predictive value, negative predictive value, sensitivity and  
52 specificity using the TTO sample as the reference. Also, we calculated the prevalence of OTU's in  
53 both niches and the concordance expressed as the proportion of overall agreement.

	Point Estimate	95% CI
Targets	15042	-
Sensitivity	0.59	0.57 - 0.61
Specificity	0.83	0.82 - 0.84
Positive predictive value	0.40	0.39 - 0.42
Negative predictive value	0.91	0.91 - 0.92
Prevalence NP	0.24	0.23 - 0.25
Prevalence TTO	0.16	0.16 - 0.17
Agreement	0.79	0.78 - 0.8

54

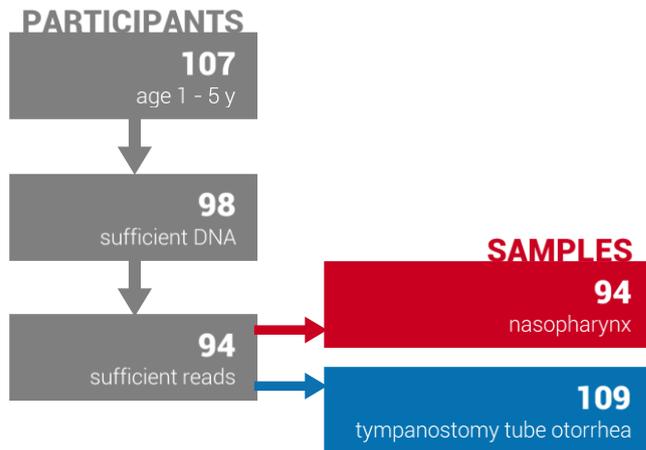
55 **eFigure 1. Culture results confirm the taxonomic annotation of the corresponding OTU's.**  
56 Boxplots visualizing the relation between the culture results for *Streptococcus pneumoniae*,  
57 *Haemophilus influenzae*, *Moraxella catarrhalis*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*  
58 and the relative abundance of the corresponding OTU as determined by 16S rRNA sequencing.



59

60 **eFigure 2. Flow chart participants and samples.**

61 Flow chart describing the number of participants and samples analyzed in this study. Only participants  
62 that had both a high-quality nasopharynx sample and a high-quality TTO samples were used for  
63 downstream analysis.

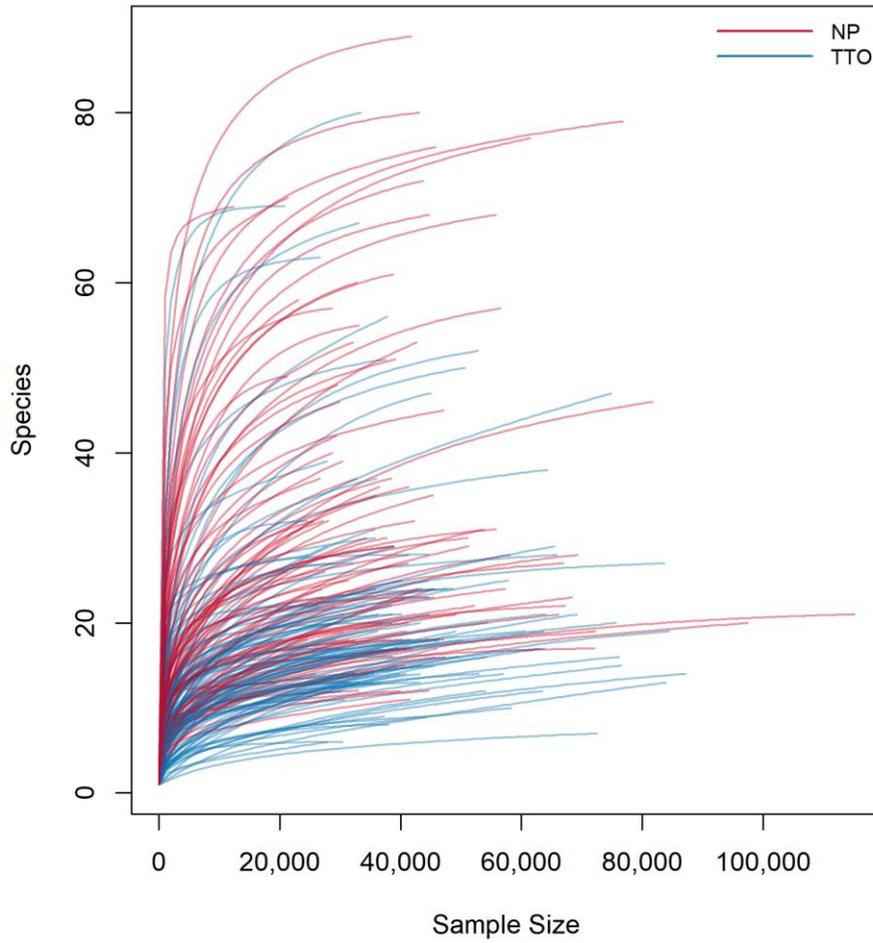


64

65

66 **eFigure 3. Rarefaction curves on raw count data.**

67 Rarefaction curves on raw count data approached plateau for both NP samples (red)  
68 (blue).

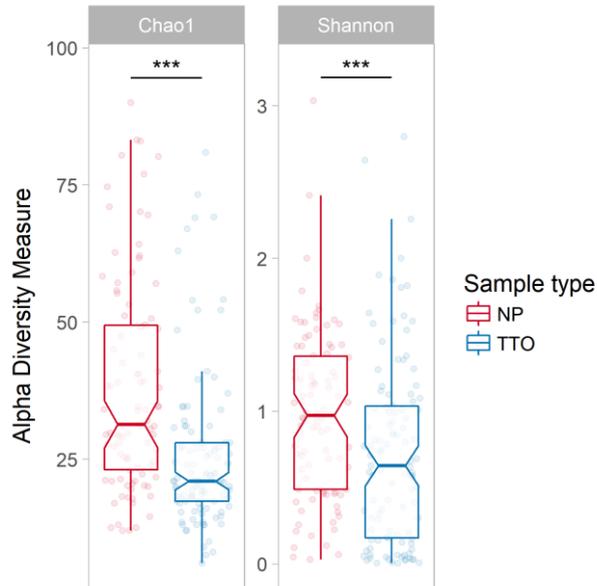


69

70 **eFigure 4.  $\alpha$ -Diversity.**

71 The ecological diversity was significantly higher in NP samples (red) compared to the TTO samples  
72 (blue), according to the Chao 1 estimate and Shannon's diversity index.

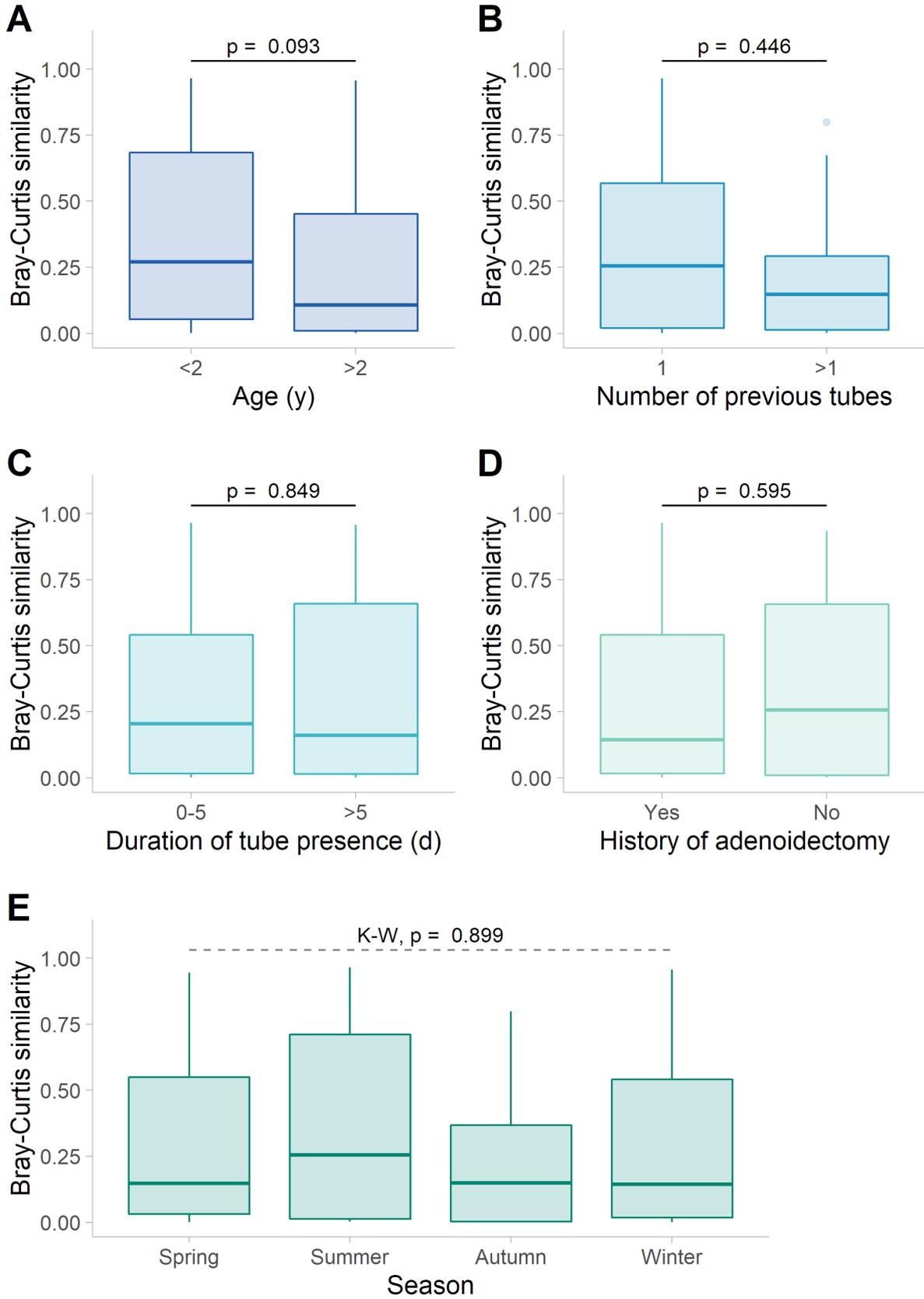
73 Significance symbols: \*\*\* =  $p < 0.001$ ; \*\* =  $p < 0.01$ ; \* =  $p < 0.05$ .



74

75

76 **eFigure 5. Similarity of paired NP and TTO samples does not vary with clinical variables.**  
77 Bray-Curtis similarity (1 – Bray-Curtis dissimilarity) of the paired NP and TTO samples of the same  
78 participant stratified by age (<2 years, n=32; >2 years n=47; A), number of previous tympanostomy  
79 tubes (including the insertion of the current tympanostomy tube; 1 tube, n=63; >1 tube, n=16; B),  
80 duration of tube presence (0-5 days, n=40; >5 days, n=39; C), history of prior adenoidectomy (yes,  
81 n=47; no, n=32; D), and season of sampling (Spring, March-May, n=18; Summer, June-August, n=17;  
82 Autumn, September-November, n=20; Winter, December-February, n=21). The Bray-Curtis similarity  
83 is bounded between 0 and 1, where 0 means that two samples are completely dissimilar, and 1 means  
84 the two sites are completely similar. P-values are based on Wilcoxon rank-sum tests (A-D) and a  
85 Kruskal-Wallis test (E).



**References**

- 88 1. Prevaes SMPJ, de Winter-de Groot KM, Janssens HM, et al. Development of the  
89 Nasopharyngeal Microbiota in Infants with Cystic Fibrosis. *Am J Respir Crit Care Med.*  
90 2016;193(5):504-515.
- 91 2. Wyllie AL, Chu MLJN, Schellens MHB, et al. Streptococcus pneumoniae in saliva of Dutch  
92 primary school children. *PLoS One.* 2014;9(7):e102045.
- 93 3. Joshi N, Fass J. Sickle: A sliding-window, adaptive, quality-based trimming tool for FastQ files  
94 (Version 1.33) [Software].
- 95 4. Nikolenko SI, Korobeynikov AI, Alekseyev MA. BayesHammer: Bayesian clustering for error  
96 correction in single-cell sequencing. *BMC Genomics.* 2013;14(Suppl 1):S7.
- 97 5. Masella AP, Bartram AK, Truszkowski JM, Brown DG, Neufeld JD. PANDAseq: paired-end  
98 assembler for illumina sequences. *BMC Bioinformatics.* 2012;13(1):31.
- 99 6. Caporaso JG, Kuczynski J, Stombaugh J, et al. QIIME allows analysis of high-throughput  
100 community sequencing data. *Nat Methods.* 2010;7(5):335-336.
- 101 7. Edgar RC, Haas BJ, Clemente JC, Quince C, Knight R. UCHIME improves sensitivity and  
102 speed of chimera detection. *Bioinformatics.* 2011;27(16):2194-2200.
- 103 8. Rognes T, Mahé F, Flouri T, Quince C, Nichols B. VSEARCH.
- 104 9. Westcott SL, Schloss PD. De novo clustering methods outperform reference-based methods for  
105 assigning 16S rRNA gene sequences to operational taxonomic units. *PeerJ.* 2015;3:e1487.
- 106 10. Quast C, Pruesse E, Yilmaz P, et al. The SILVA ribosomal RNA gene database project:  
107 improved data processing and web-based tools. *Nucleic Acids Res.* 2012;41(D1):D590-D596.
- 108 11. Caporaso JG, Bittinger K, Bushman FD, DeSantis TZ, Andersen GL, Knight R. PyNAST: a  
109 flexible tool for aligning sequences to a template alignment. *Bioinformatics.* 2010;26(2):266-  
110 267.
- 111 12. Price MN, Dehal PS, Arkin AP. FastTree 2--approximately maximum-likelihood trees for large  
112 alignments. *PLoS One.* 2010;5(3):e9490.
- 113 13. Subramanian S, Huq S, Yatsunenko T, et al. Persistent gut microbiota immaturity in  
114 malnourished Bangladeshi children. *Nature.* 2014;510(7505):417.
- 115 14. Paulson JN, Stine OC, Bravo HC, Pop M. Differential abundance analysis for microbial  
116 marker-gene surveys. *Nat Methods.* 2013;10(12):1200-1202.
- 117 15. Anderson MJ, Crist TO, Chase JM, et al. Navigating the multiple meanings of  $\beta$  diversity: a  
118 roadmap for the practicing ecologist. *Ecol Lett.* 2011;14(1):19-28.
- 119