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Gene targeted	Sense	Antisense
β-actin	5'-TGTTACCAACTGGGACGACA-3'	5'-CTGGGTCATCTTTTCACGGT-3'
GAPDH	5'-ACCCAGAAGACTGTGGATGG-3'	5'-GGATGCAGGGATGATGTTCT-3'
Wnt1	5'-CAGCACCACTAGAGGAAACGA-3'	5'-CTGGGCACATATCTTACAGCATT-3'
Wnt2	5'-CCTCCGAAGTAGTCGGGAATC-3'	5'-GCAGGACTTTAATTCTCCTTGGC-3'
Vnt2b	5'-ACATGCAAGGCCCCTAAG-3'	5'-AGGATTGAGGGTAGAGGAAGG-3'
Wnt3a	5'-GTGAGGACATTGAATTTGGAGG-3'	5'-ACTTGAGGTGCATGTGACTG-3'
Wnt4	5'-TGCGAGGTAAAGACGTGCTG-3'	5'-CTTGAACTGTGCATTCCGAGG-3
Wnt5a	5'-CAACTGGCAGGACTTTCTCAA-3'	5'-CATCTCCGATGCCGGAACT-3'
Wnt5b	5'-GACTGACGCCAACTCCTG-3'	5'-TGCTCCTGATACAACTGACAC-3'
Wnt6	5'-TCAAGACTCTTTATGGATGCGC-3'	5'-ATGGCACTTACACTCGGTG-3'
Wnt7a	5'-ACGAGTGTCAGTTTCAGTTCC-3'	5'-AATCGCATAGGTGAAGGCAG-3'
Wnt7b	5'-AATGAGGCGGGCAGAAAG-3'	5'-TGCGTTGTACTTCTCCTTGAG-3'
Wnt8a	5'-TCATGTACGCAGTCACCAAG-3'	5'-TTTTCCCCGAACTCCACG-3'
Wnt8b	5'-GTACACCCTGACTAGAAACTGC-3'	5'-AAACTGCTTGGAAATTGCCTC-3'
Wnt9a	5'-GGTTCCTGTAGCCCACTAATG-3'	5'-GAAAGAGATGAAGCTCCCGTC-3'
Wnt9b	5'-CCAAGAGAGGAAGCAAGGAC-3'	5'-AACAGGTACGAACAGCACAG-3'
Wnt10a	5'-CGCTTCTCTAAGGACTTTCTGG-3'	5'-GTGGCATTTGCACTTACGC-3'
Wnt10b	5'-ATCGCCGTTCACGAGTGTC-3'	5'-GGAAACCGCGCTTGAGGAT-3'
Wnt11	5'-CCAAGCCAATAAACTGATGCG-3'	5'-GGCATTTACACTTCGTTTCCAG-3'
Wnt16	5'-AGTGCAGGCAACATGACCG-3'	5'-CCACATGCCGTACTGGACATC-3'
Lef1	5'-ACACCCTGATGAAGGAAAGC-3'	5'-GACCCATTTGACATGTACGG-3'
Tcf4	5'-ACGTTTGAGCTATCCATCCC-3'	5'-GTGCAGGAAGAGGTGCTGTA-3'
Axin2	5'-ACACATGCAGAAATGGGTCA-3'	5'-GGACGTCTGTGACAAGCAGA-3'
ALP	5'-ACCTTGACTGTGGTTACTGC-3'	5'-CATATAGGATGGCCGTGAAGG-3'
Oc	5'-TGTGACGAGCTATCAAACCAG-3'	5'-GAGGATCAAGTTCTGGAGAGC-3'
Osx	5'-GGAGACCTTGCTCGTAGATTTC-3'	5'-GGGATCTTAGTGACTGCCTAAC-3'
FABP4	5'-AAGAAGTGGGAGTGGGCTTT-3'	5'-AATCCCCATTTACGCTGATG-3'
PPAR-γ	5'-ATAGGTGTGATCTTAACTGCCG-3'	5'-CCAACAGCTTCTCCTTCTCG-3'

A Primers for quantitative Real-time polymerase chain reaction

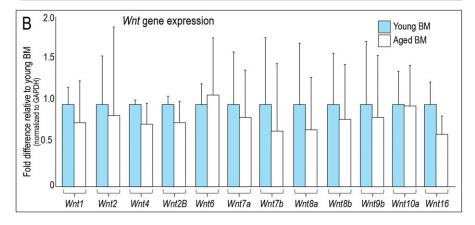


Fig. E-1

Fig. E-1A Primers used for quantitative real-time polymerase chain reaction. **Fig. E-1B** Quantitative real-time polymerase chain reaction to evaluate relative expression levels of Wnt ligands in bone marrow harvested from young (blue bars; n = 3) and aged (white bars; n = 3) bone marrow (BM) donors. Gene expression levels normalized to glyceraldehyde 3-phosphate dehydrogenase (GAPDH).

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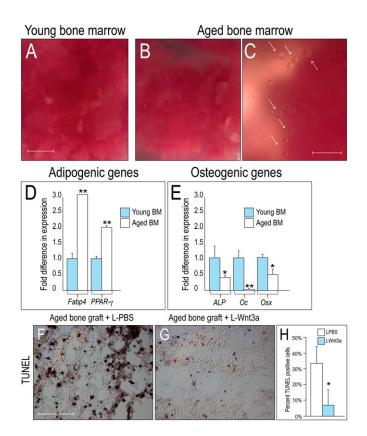


Fig. E-2

Aged bone marrow undergoes fatty degeneration. Compared with the gross appearance of bone marrow harvested from young donors (**Fig. E-2A**), bone marrow from aged donors is fatty (**Fig. E-2B**); arrows indicate fat droplets (**Fig. E-2C**). **Fig. E-2D** Quantitative reverse transcription-polymerase chain reaction(qRT-PCR) analyses for the adipogenic genes fatty acid-binding protein 4 (Fabp4) and peroxisome proliferator-activated receptor gamma (PPAR- γ) in bone marrow (BM) from young animals (blue bars, n = 3) compared with bone marrow from aged animals (white bars; n = 3). **Fig. E-2E** qRT-PCR analyses for the osteogenic genes alkaline phosphatase (ALP), osteocalcin (Oc), and osterix (Osx) in bone marrow (BM) from young animals (blue bars; n = 3). Gene expression levels normalized to beta-actin. **Figs. E-2F and E-2G** Terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) staining in aged bone marrow following a twelve-hour incubation in liposomal phosphate-buffered saline solution (L-PBS) (**Fig. E-2F**) (n = 3) or L-Wnt3a (**Fig. E-2G**) (n = 3; effective concentration = 150 ng/mL), quantified in **Fig. E-2H**. Single asterisk denotes p < 0.01. Scale bars: 1 mm (Fig. E-2A [scale bar in Fig. E-2A also applies to Fig. E-2B]); 400 µm (Fig. E-2C); and 50 µm (Fig. E-2F [scale bar in Fig. E-2F also applies to Fig. E-2G)].

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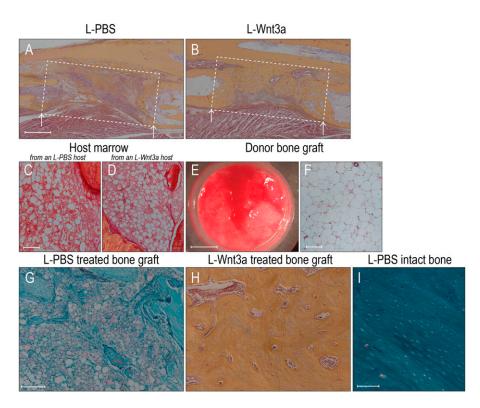


Fig. E-3

Movat pentachrome staining of injury site (boxed area) treated with aged bone marrow incubated in liposomal phosphate-buffered saline solution (L-PBS) (Fig. E-3A) or L-Wnt3a (Fig. E-3B). Fig. E-3C and E-3D Under polarized light, picrosirius red staining confirms that aged host rabbits treated with L-PBS (Fig. E-3C) or liposomal Wnt3a protein (L-Wnt3a) (Fig. E-3D) both showed evidence of fatty degeneration in their bone marrow. Fig. E-3E Morphology of fatty bone marrow harvested from an aged donor rabbit, prior to transplantation. Fig. E-3F Gömöri trichrome staining confirms that the graft consists of fatty bone marrow. Fig. E-3G Safranin O/fast green staining identifies cartilage, fibrocartilage, immature osteoid, and adipose in bone grafts treated with L-PBS. Fig. E-3H Pentachrome staining identifies new osteoid matrix with large blood vessel spaces formed in bone grafts treated with L-Wnt3a. Compare this new bone matrix with Gömöri trichrome (Fig. E-3I) staining of the host's intact bone cortex, which has a lamellar organization, typical of mature bone. Arrows mark the edge of intact bone. Scale bars: 500 µm (Fig. E-3A [scale bar in Fig. E-3A also applies to Fig. E-3B]); 200 µm (Fig. E-3C [scale bar in Fig. E-3C also applies to Fig. E-3D]); 250 µm (Fig. E-3E); 100 µm (Fig. E-3F); 200 mm (Fig. E-3G [scale bar in Fig. E-3G also applies to Fig. E-3D]); and 100 µm (Fig. E-3F); 200 mm (Fig. E-3G)]; 250 µm (Fig. E-3H]); and 100 µm (Fig. E-3F); 200 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]); and 100 µm (Fig. E-3F)]; 200 µm (Fig. E-3H]);