

TABLE E-1 Characteristics of Oligonucleotide Primers and Probes Used in the PCR-RLB

Oligonucleotide Name	Sequence	Target	Pmol of Oligonucleotide per Lane on Membrane	Nucleotide Position in the 16S rRNA Gene
Primers				
16S8-27F	AGAGTTTGATCMTGGCTCAG	Eubacteria		8-27
B-16S509-525R*	GCTGCTGGCACGDAGTT	Eubacteria		509-525
Probes**				
A-16S A339	TCCTACGGGAGGC(AT)GCA	Eubacteria	25 & 50	339-355
A-16S CAMPY113	GTGAGTAAGGTATAGTTAATC	Campylobacter genus	200	113-133
A-16S CORYN451	CGAAGCTTTTGTGACGG	Corynebacterium genus	20	451-467
A-16S Eaero474	GGCGATTGACGTTACTC	<i>Enterobacter aerogenes</i>	100	474-491
A-16S Eagg1471	CCTGTCGATTGACGTTACC	<i>Enterobacter agglomerans</i>	200	471-489
A-16S ECEAE136	CTGATGGAGGGGGATAA	Enterobacteriaceae group	1500	136-153
A-16S Ecoli451	AAGGGAGTAAAGTTAATACCT	<i>Escherichia coli</i>	50	451-471
A-16S Efaec220	TTTCGGGTGTCGCTGAT	<i>Enterococcus faecalis/faecium</i>	100	220-236
A-16S ENTER448	AGGAAGGTGTTGTGGTTAA	Enterobacter genus	50	448-467
A-16S Fmagn210	CAAAGATTTATCGGTCATAGA	<i>Finexgoldia magna</i> (<i>Peptostreptococcus magnus</i>)	1500	210-230
A-16S Hpylo82	CTTGCTAGAGTGCTGATTA	<i>Helicobacter pylori</i>	1500	82-100
A-16S Kpneu73	CACAGAGAGCTTGCTCT	<i>Klebsiella pneumoniae</i>	1500	73-90
A-16S Mpneu75	TAGTAATACTTTAGAGGCGAA	<i>Mycoplasma pneumoniae</i>	1000	75-95
A-16S Mtube211	CTTTAGCGGTGTGGGAT	<i>Mycobacterium tuberculosis</i>	800	211-227
A-16S MYCOB72	AAAGGTCTCTTCGGAGATA	Mycobacterium genus	800	72-90
A-16S NEISS120	AACATATCGGAACGTACC	Neisseria genus	400	120-137
A-16S Ngono129	GAACGTACCGGGTAGC	<i>Neisseria gonorrhoeae</i>	200	129-144
A-16S Nmeni185	TCTTGAGAGAGAAAAGCAGG	<i>Neisseria meningitidis</i>	50	185-201
A-16S Pacne69	GAAAGGCCCTGCTTTTG	<i>Propionibacterium acnes</i>	1000	69-85
A-16S Pmira461	GGTTAATACCCTTATCAATTGA	<i>Proteus mirabilis</i>	200	461-482
A-16S PSEUD141	GGATAACGTCCGGAAC	Pseudomonas genus	800	141-157
A-16S Pvulg463	TTAATACTCTTAGCAATTGACG	<i>Proteus vulgaris</i>	2	463-484
A-16S SALMO452	GGTGTGTGGTTAATAACC	Salmonella genus	400	452-470
A-16S Saure195	CATGGTTCAAAAAGTGAAAGAC	<i>Staphylococcus aureus</i>	400	195-215
A-16S Spyog85	TAACGCATGTTAGTAATTTAAAAG	<i>Streptococcus pyogenes</i>	50	85-108
A-16S STAPH129	AACCTACCTATAAGACTGG	Staphylococcus genus	400	129-147
A-16S STREP142	GCGGGGGATAACTATTG	Streptococcus genus	750	142-158
A-16S TpD	CCGCTTCTATGTGACAG	Spike DNA	25 & 100	
Hybridization control				
B-A339/Salg/Ngon101	GCTACCCGGTACGTTCCAATAGTTAT CCCCCGCTGC			
Spike oligonucleotide				
TpD spike	AGAGTTTGATCATGGCTCAGCCGCTT CTATGTGACAGAACTTCGTGCCAGCA GC			

*Reversed primer is labeled with a 5'-biotin label.

**All oligonucleotide probes are labeled with a 5'-C6-aminolinker.

TABLE E-2 Species Identification by Culture, PCR-RLB, and DNA Sequencing of PCR Products in 76 Patients

Culture	No. of Patients (%)	RLB	No. of Patients (%)	Sequencing of RLB Samples	No. of Patients (%)
<i>S. aureus</i>	8 (11)	<i>S. aureus</i>	10 (13)	<i>S. aureus</i>	8 (11)
				Not determinable	2 (3)
Coagulase-negative staphylococci	5 (7)	Staphylococcus genus	4 (5)	<i>S. epidermidis</i>	2 (3)
<i>S. lugdunensis</i>	1 (1)			<i>S. lugdunensis</i>	1 (1)
No specification	4 (5)			Not determinable	1 (1)
Streptococcus spp.	5 (7)	Streptococcus genus	8 (11)	<i>S. dysgalactiae</i>	3 (4)
α -hemolytic streptococci	2 (3)			<i>S. mitis</i>	2 (3)
β -hemolytic streptococci	2 (3)			Not determinable	3 (4)
<i>S. oralis</i>	1 (1)				
<i>E. faecalis</i>	1 (1)	<i>E. faecalis/faecium</i>	1 (1)	<i>Haemophilus influenzae</i>	1 (1)
<i>E. cloacae</i>	1 (1)	<i>E. agglomerans</i>	2 (3)	<i>E. cloacae</i>	1 (1)
				Not determinable	1 (1)
<i>P. canis</i>	1 (1)	Eubacteria	2 (3)	<i>P. canis</i>	1 (1)
<i>H. influenzae</i>	1 (1)			<i>H. influenzae</i>	1 (1)
		Enterobacteriaceae	1 (1)	Not determinable	1 (1)
		Corynebacterium genus	1 (1)	<i>C. xerosis</i>	1 (1)
		<i>P. acnes</i>	1 (1)	<i>P. acnes</i>	1 (1)
Mixed culture	4 (5)	Mixed culture	5 (7)		
<i>M. morganii</i> & Enterococcus spp. & <i>P. aeruginosa</i>	1 (1)	<i>E. agglomerans</i> & <i>E. faecalis</i> & <i>P. aeruginosa</i>	1 (1)	<i>M. morganii</i>	1 (1)
<i>E. coli</i> & Enterococcus spp.	1 (1)	<i>E. coli</i> & <i>E. faecalis</i>	1 (1)	<i>E. coli</i>	1 (1)
<i>M. tuberculosis</i> & <i>S. aureus</i>	1 (1)	<i>M. tuberculosis</i> & <i>S. aureus</i>	1 (1)	Not determinable	1 (1)
α -hemolytic streptococci & <i>S. aureus</i>	1 (1)	Streptococcus genus & Staphylococcus genus	1 (1)	<i>S. dysgalactiae</i>	1 (1)
		<i>E. faecalis/faecium</i> & Staphylococcus genus	1 (1)	<i>S. epidermidis</i>	1 (1)
<i>C. albicans</i>	1 (1)				
Negative	49 (64)	Negative	41 (54)		

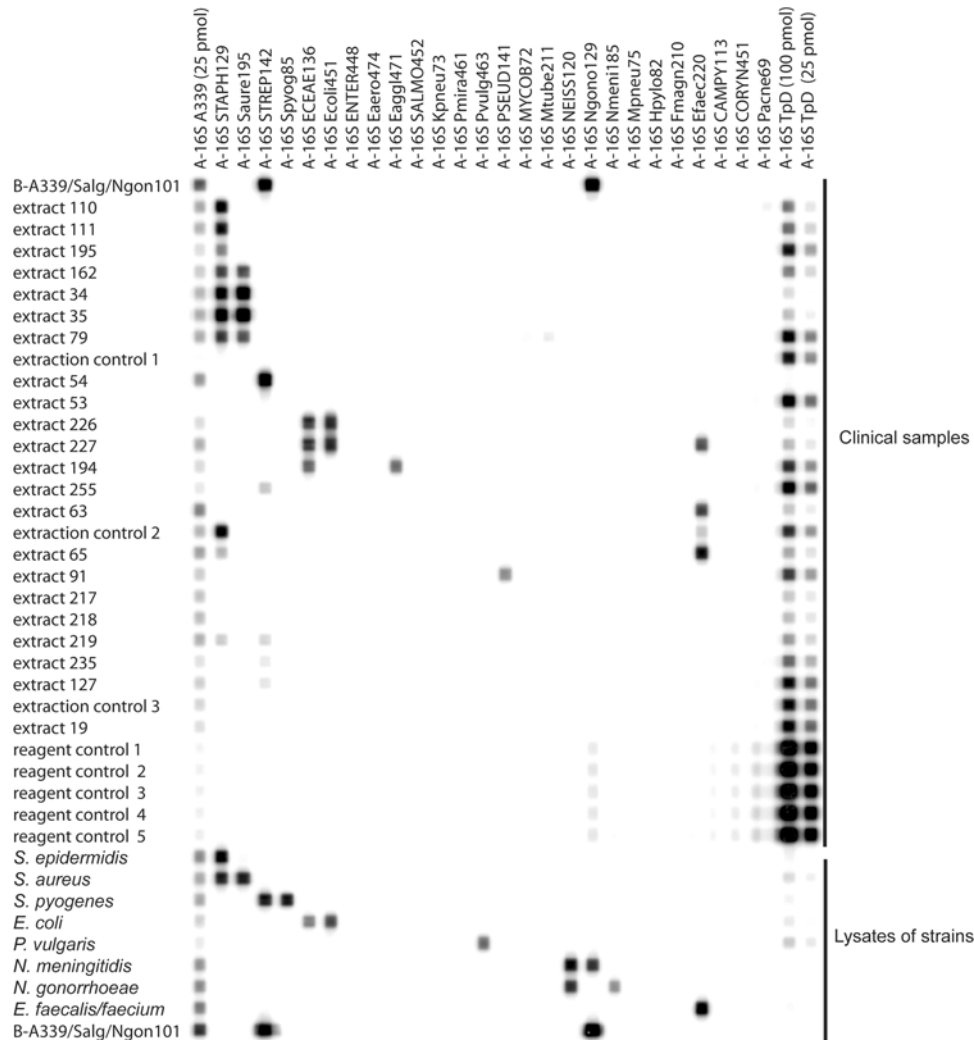


Fig. E-1

Reverse line blot analysis of 16S gene amplicons from patients in whom an orthopaedic infection was suspected on the basis of patient data. Polymerase-chain-reaction products obtained from patients and controls were applied in the rows and oligonucleotide probes, in the columns. The specificities of the oligonucleotide probes can be found in Table E-1. In the first and last row, a biotin-labeled oligonucleotide reacting with three different oligonucleotide probes was included as a positive hybridization control. The eight lanes at the bottom of the blot display results obtained with lysates of a number of bacterial species. Extraction control 2 exemplifies a false-positive extraction control. Extraction control 3 shows weak reactivity with the eubacterial probe, illustrating the normal background reactivity with this probe in the assay. Extract = polymerase chain reaction on extracts from clinical samples, extraction control = polymerase chain reaction on negative controls obtained during DNA extraction from clinical samples, and reagent control = negative control obtained by polymerase chain reaction on reagents only.