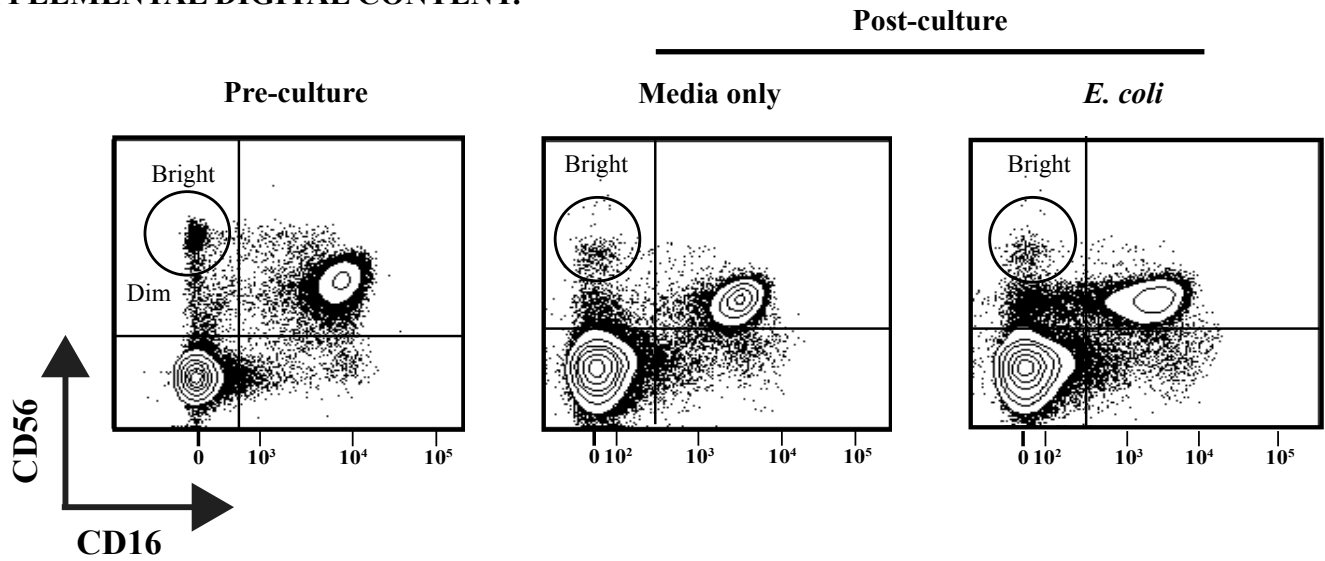
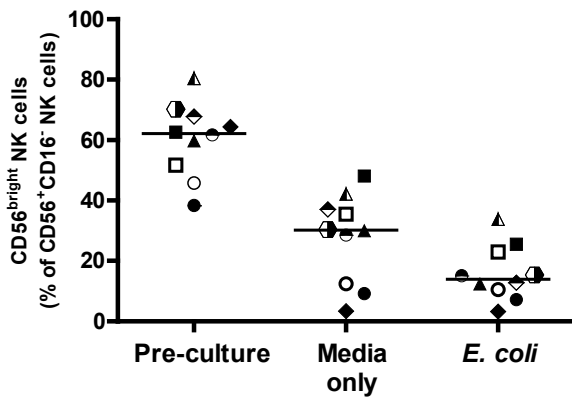


SUPPLEMENTAL DIGITAL CONTENT.

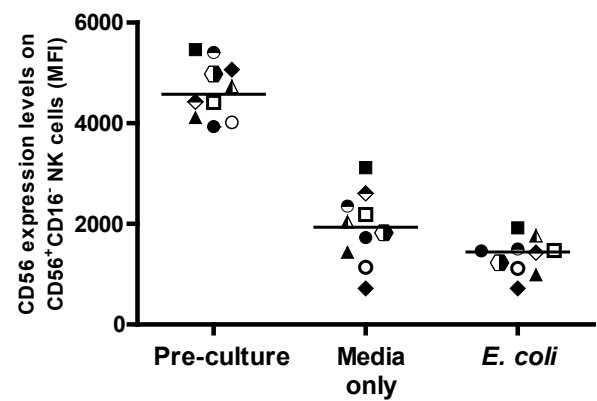
A)



B)



C)



D)

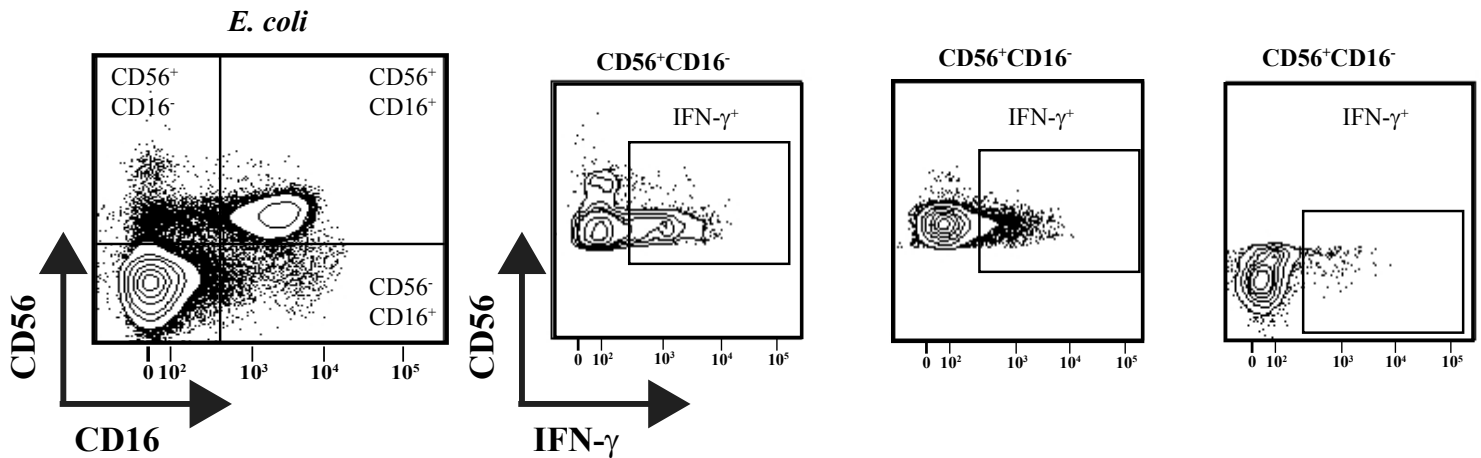


Figure S1. Gating strategy to identify IFN- γ -producing NK cells. PBMC were cultured for 16-22hrs with commensal *E. coli* (6 *E. coli*:1 PBMC) with Brefeldin A added for the final 12-18hrs. Multi-color flow cytometry techniques were used to identify intracellular IFN- γ within the NK cell subsets. An initial lymphocyte gate was established that excluded dead cells and cellular debris and NK cells were defined using CD56 and CD16 within CD3⁻ lymphocytes (gating not shown). (A) Flow plots showing down regulation of CD56 expression (bright/dim) within the CD56⁺CD16⁻ cells following *in vitro* culture and commensal *E. coli* stimulation of PBMC from an uninfected subject. Enumeration of the (B) percent of CD56^{bright} NK cells within CD56⁺CD16⁻ cells and (C) expression levels of CD56 within CD56⁺CD16⁻ NK cells are shown from 10 uninfected subjects pre and post *in vitro* culture. Symbols represent individual donors and bars indicate the median values. (D) To accommodate the down regulation of CD56, IFN- γ expression was determined within each NK cell subset defined as CD56⁺CD16⁻, CD56⁺CD16⁺ and CD56⁻CD16⁺ (same donor as shown in (A)).

SUPPLEMENTAL DIGITAL CONTENT.

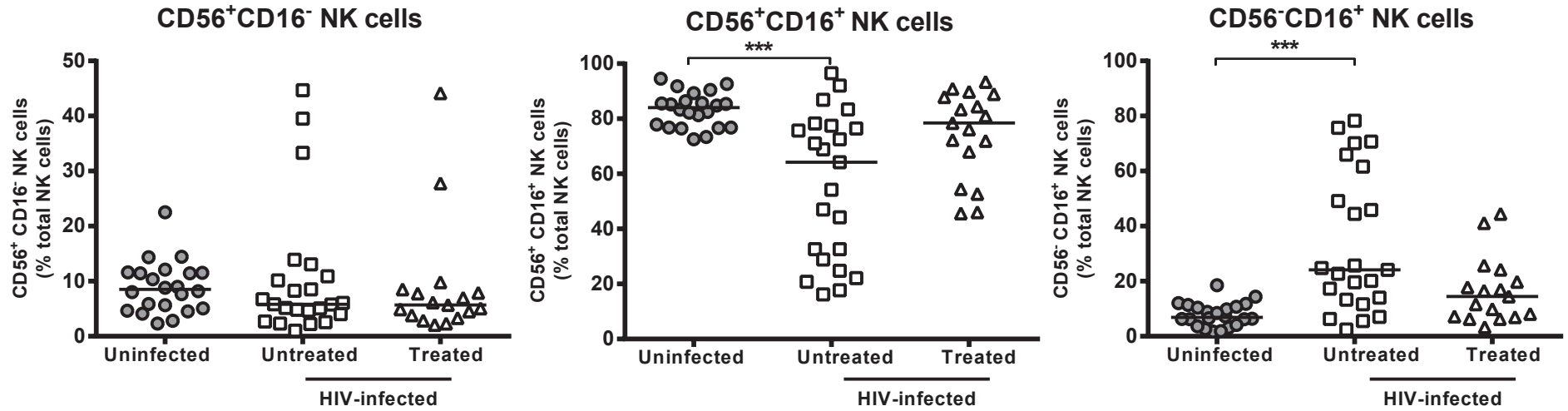


Figure S2. Altered frequencies of blood NK cells in untreated, HIV-1 infected individuals. Percentages of blood NK cells prior to *in vitro* stimulation (baseline) were evaluated using multi-color flow cytometry. Baseline percentages of each NK cell subset, as a fraction of total NK cells, within PBMC from uninfected (n=22), untreated (n=23) and treated (n=17) HIV-infected subjects were evaluated. Lines represent median values. Comparisons between multiple groups were performed using the Kruskal-Wallis test and comparisons conducted between the cohorts when $P < 0.05$ using the Dunn's Multiple Comparison test. *** $P < 0.001$.