Figure S1. Panel A: 2 x 10^5 Jurkat E6-1 cells were pre-treated with 10 uM of DMSO (control), PP1, PP2 or SU6656 for 1 hr, washed in PBS, and then grown for two days in RPMI. Cells were then counted using trypan blue stain exclusion. **Panel B:** T cell lines were transduced with adenovectors expressing c-Src mutants. Adenovirus vectors (EV = empty vector, WT c-Src = Wildtype c-Src, DN c-Src = Dominant Negative c-Src) were administered to Jurkat E6-1, Hut 78 and Kit 225 cells at an MOI of 750 for 48 hr. The percent of live cells (7-AAD -ve) was then measured using FACS.

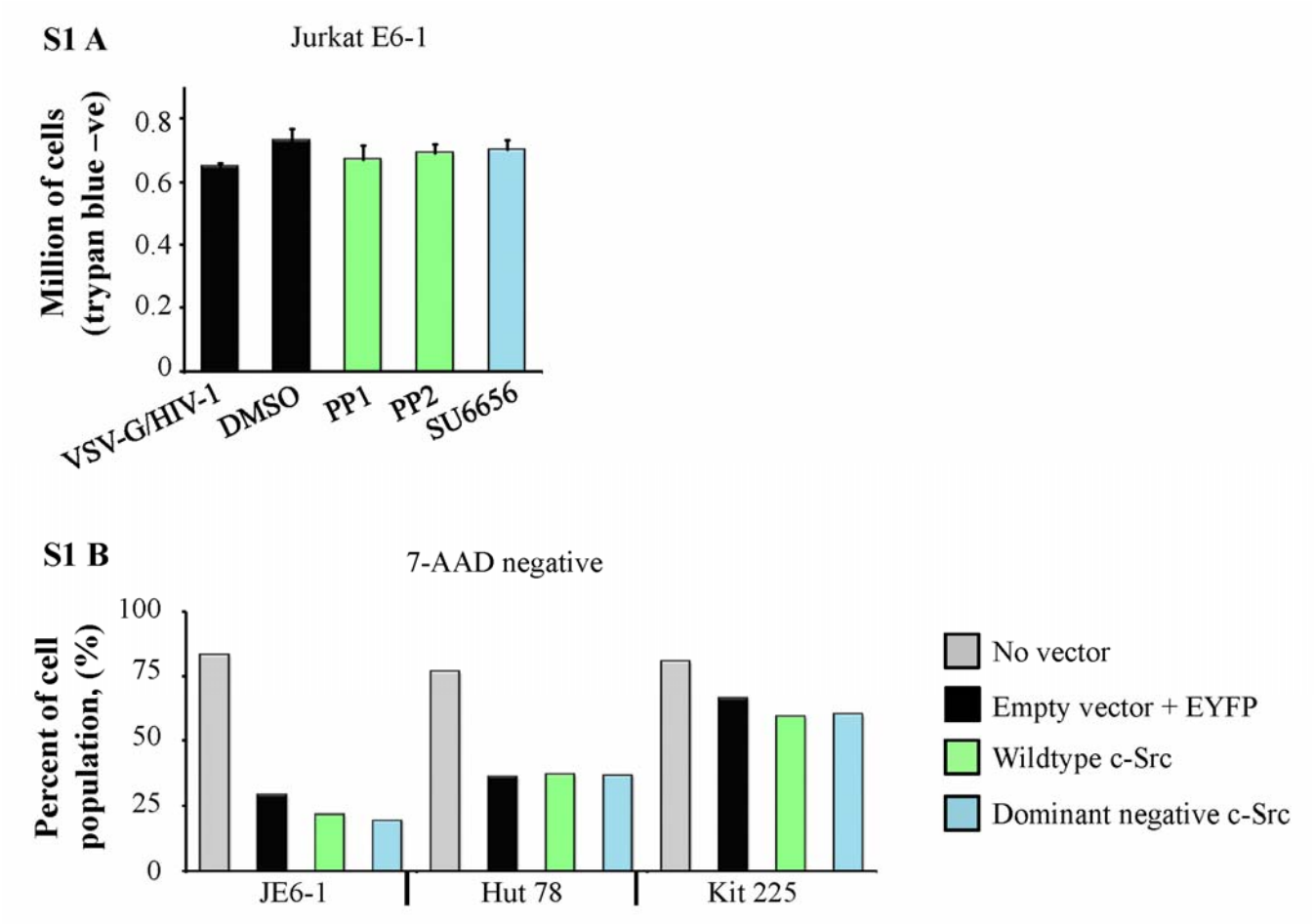

Figure S2. siRNA titration and time-course optimization in Jurkat E6-1 cells. **Panel A**: Jurkat E6-1 cell survival two days post-transfection of non-targeting siRNA, c-Src siRNA or Pyk2 siRNA, as measured by trypan blue staining. **Panels B-E**: Two days post-lipofection, 5 x 10^5 cells were lysed with RIPA to perform an 8% SDS-PAGE followed by a Western blot. Blots were stripped and re-probed with anti-c-Src (B and C), anti-Pyk2 (D and E) and anti-a-tubulin. Quantification of Pyk2 and c-Src protein was relative to α-tubulin control. **Panels F and G**: Time-course for optimal c-Src knockdown post-lipofection.

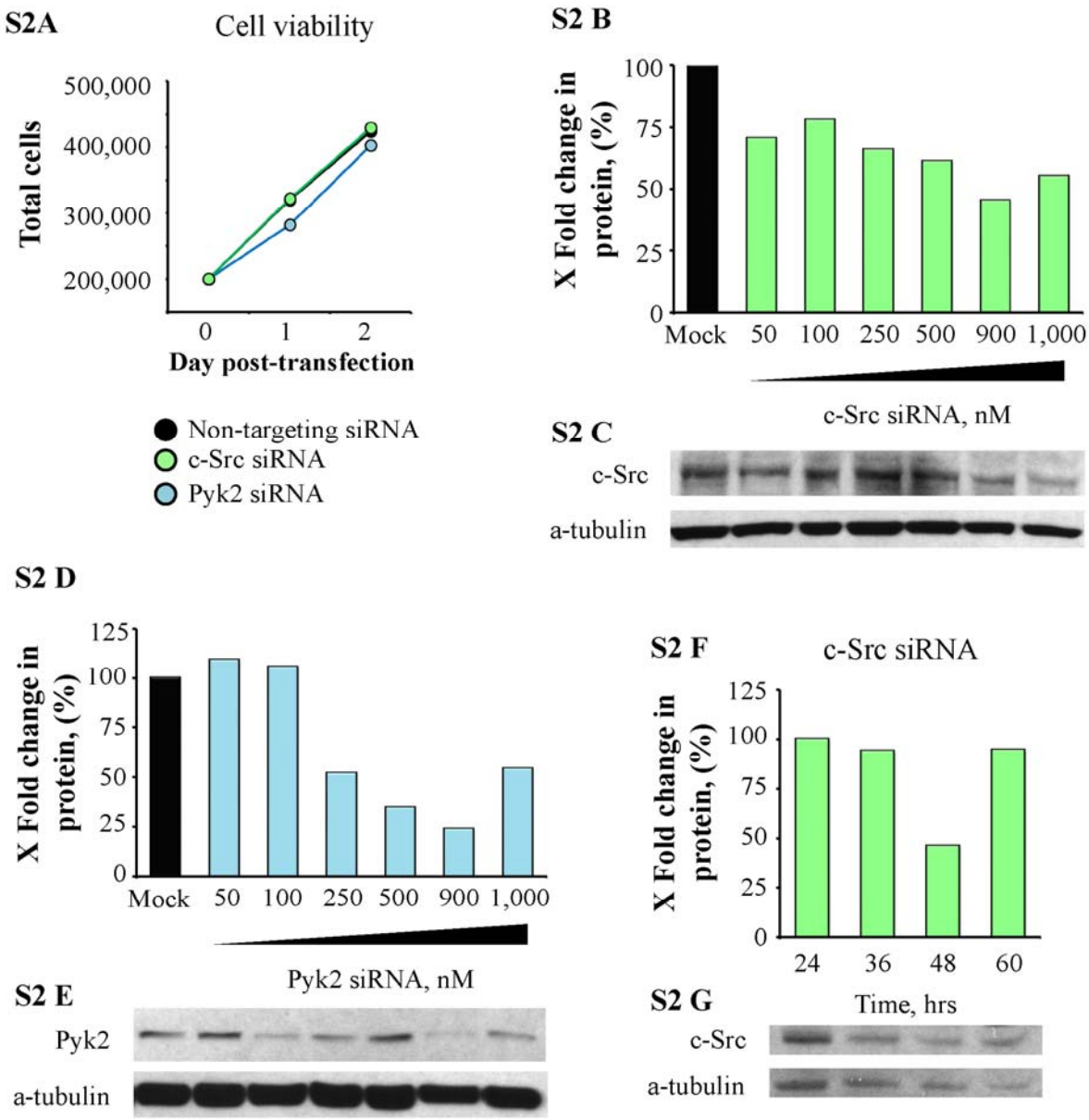

Figure S3. To measure reverse transcriptase activity, a standard curve was constructed by plotting the qPCR cycle threshold of amplified target cDNA verses the activity of recombinant HIV-1 reverse transcriptase (**Panel B**).