SDC, Figure 1. Gating strategy to evaluate CD154+T-cytotoxic memory cells (CD154+TcM) by flow cytometry in each reaction condition. SDC, 1a. Scatterplot shows the location of lymphocytes on the basis of forward and side scatter. SDC, 1b. T-cells (CD3+) are gated from total lymphocytes using side scatter by CD3 in the FITC channel. SDC, 1c. Live T-cells are identified after exclusion of dead cells which take up the dye 7-AAD (CD3+7AAD+). SDC, Panel 1d. By gating further on live CD3 cells, responder T-cytotoxic cells (Tc or CD8) which have been pre-labeled with anti-CD8-APCH7 are separated from stimulator Tc which have been prelabeled with anti-CD8-PE-Cy7, and from T-cells which do not express the CD8 marker (Q3). SDC, 1e. By gating further on responder Tc, those with the memory marker CD45RO (TcM) and are labeled with APC-labeled anti-CD45RO are separated from those that do not express this marker (naïve). SDC, 1f-1h. By gating further on responder TcM, those that express CD154 and (CD154+TcM) and are labeled with PE-labeled anti-CD154 are separated from those that do not express this marker. SDC, 1f. The area occupied by CD154+TcM is identified by a gate which excludes all cells in the negative control reaction. In this reaction, recipient cells are incubated alone without fluorescent antibody to CD154. This scatterplot configuration is fixed and used to analyze all other reaction conditions. SDC, 1g. The background reaction in which recipient cells are incubated with anti-CD154-PE without stimulator shows "background" counts of CD154+TcM. SDC, 1h. The addition of HLA-non-identical stimulator cells in this stimulated reaction results in stimulation-induced increase in CD154+TcM counts.

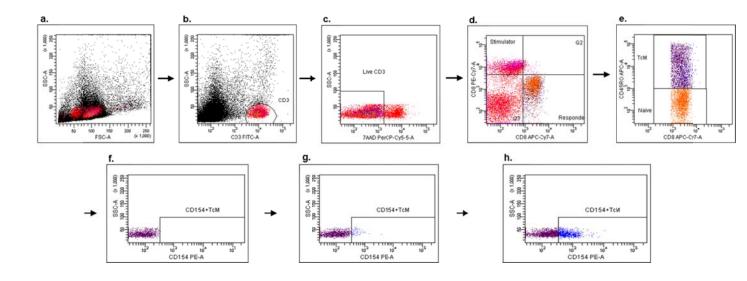
FSC: Forward scatter, SSC: Side scatter, A: Area, FITC: Fluorescein isothiocyanate, PerCP-Cy5.5: Peridinin chlorophyll protein and cyanine 5.5 tandem dye, APCH7: allophycocyanin and cyanine

Supplemental Digital Content (SDC)

Figure

7 tandem dye, PECy7: phytoerythrin and cyanine 7 tandem dye, APC: allophycocyanin, PE: Phycoerythrin.

SDC, Figure 1



Supplemental Digital Content (SDC)

Figure

SDC Figure 2. General approach to training-set/validation-set testing maximizes utilization of samples from this rare test population. A subject e.g. subjects X or Y, can provide up to one pretransplant and one post-transplant sample in either training set or the validation set. Arrows indicate that a sample used for model building in the training set can only be used to predict outcomes of independent validation set samples for the corresponding time period. Therefore, if a subject, e.g. Subject Z, provided samples for model building in the training set, subsequent samples from a different time period could still be included in the validation set. Strikethroughs imply unacceptable samples.

Figure S2

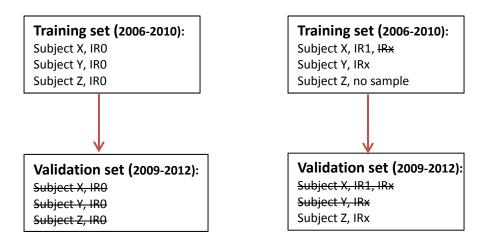


Table S1: Mean immunoreactivity index (IR) for allospecific CD154+T-cytotoxic memory cells (CD154+TcM) in training and validation set samples R=Rejector, NR=Non-rejector, IR0=pretransplant samples, IR1+IRx=post-transplant samples obtained within the first 60 days after transplantation (IR1), and days 200-onward after transplantation (IRx), CI=confidence interval, low=lower, up=upper, SD=standard deviation, Min=minimum, Max=maximum.

Data Type	Sample type	Outcome	N	Mean	CI-low	CI-up	SD	Median	Min	Max
Training	IR0	R + NR	49	3.1	1.1	5.2	7.1	1.4	0.06	46.4
		NR	24	1.1	0.5	1.7	1.4	0.7	0.06	7.0
		R	25	5.0	1.1	9.0	9.6	1.9	0.5	46.4
	IR1 +	R + NR	98	1.6	1.0	2.3	3.2	0.7	0.03	26.4
	IRx	NR	74	0.8	0.5	1.1	1.2	0.6	0.03	6.7
		R	24	4.1	1.8	6.4	5.5	2.5	0.05	26.4
Data Type	IR Level	Outcome	N	Mean	CI-low	CI-up	SD	Median	Min	Max
Validation	IR0	R + NR	33	1.2	0.7	1.6	1.2	0.9	0.01	7.1
		NR	19	0.7	0.6	0.9	0.3	0.8	0.01	1.5
		R	14	1.7	0.8	2.7	1.6	1.4	0.4	7.1
	IR1 +	R + NR	64	1.2	1.0	1.4	0.8	0.9	0.3	3.9
	IRx	NR	45	1.0	0.8	1.3	0.8	0.8	0.3	3.9
		R	19	1.6	1.2	2.0	0.8	1.4	0.5	3.7

The following additional studies were performed to determine test variability.

- 1. To document variability if three-different operators and three different flow cytometers tested the same sample on the same day, assays between twenty one unique pairs of HLA-mismatched PBL from normal human subjects were performed by three different operators and instruments. Variability in CD154+TcM generated in the stimulated reaction was determined. Results are summarized in SDC, Tables 2a and 2b.
- To determine variability in test results between two technologists, five samples were
 assayed simultaneously by two different technologists on the same day. Variability in
 CD154+TcM generated in the stimulated reaction was determined. Results are
 summarized in SDC, Tables 3a and 3b.

Table S2a: Mean %CD154TcM on same-day testing by three different technologists/instruments (1a, 1b, 1c)

Run	Reaction	N	Mean	CI-low	CI-up	SD	Median	Min	Max
1a	Background	21	4.0	2.3	5.7	3.8	2.9	0.5	13.2
	Stimulated	21	24.5	19.5	29.6	11.0	24.7	9.0	55.0
1b	Background	21	5.0	2.8	7.2	4.9	3.5	0.9	19.2
	Stimulated	21	22.8	18.0	27.6	10.5	22.4	9.8	52.2
1c	Background	21	5.8	3.2	8.3	5.7	3.4	1.2	18.4
	Stimulated	21	22.9	18.0	27.8	10.8	22.4	9.8	51.7

Table S2b. Mean %CV for %CD154+TcM in samples assayed by three technologists on three instruments

Run	Reaction	N	Mean	CI-	CI-	SD	Median	Min	Max
				low	up				
1a, 1b &	Background	21	34.3	26.1	42.2	17.9	32.2	7.7	76.3
1c	Stimulated	21	8.2	6.0	10.4	4.8	6.5	2.4	20.7
1a & 1d	Background	20	45.3	33.8	56.8	24.5	42.6	5.0	87.2
14 & 10	Stimulated	20	8.9	5.6	12.2	7.0	6.5	0.8	24.9

Table S3a: Mean %CD154+TcM for all (n=5) samples for each of two technicians

Tech	Reaction	N	Mean	CI- low	CI- up	SD	Median	Min	Max
Tech 1	Background	5	1.2	0.6	1.9	0.5	1.3	0.4	1.7
	Stimulated	5	5.6	3.1	8.0	2.0	4.8	3.1	7.8
Tech 2	Background	5	0.7	0	1.5	0.6	0.6	0.1	1.7
	Stimulated	5	5.8	3.2	8.4	2.1	5.3	3.0	8.2

Table S3b: Mean %CV for %CD154+TcM for all (n=5) samples for each of two technicians

Tech	Reaction	N	Mean	CI- low	CI- up	SD	Median	Min	Max
Tech 1	Background	5	55.1	19.4	90.9	28.8	64.5	18.7	84.0
and 2	Stimulated	5	4.8	1.1	8.5	3.0	4.2	1.8	8.4

Table S4: Performance of optimal (multiple variable) post-transplant (7a upper table) and pre-

transplant (7b lower table) models based on IR of CD154+TcM in training and validation sets.

Cohort	AUC	Cut value	Sensitivity	95%	Specificity	95%	PPV	95%	NPV	95% CI
				CI		CI		CI		
Training set (n=98)	0.937	-0.85	95%	74%,	89%	78%,	71%	51%,	98% (62/63)	90%,
			(20/21)	100%	(62/70)	95%	(20/28)	86%		100%
Validation set	0.711	-0.85	53% (10/19)	29%,	83% (40/48)	69%,	56%	31%,	82% (40/49)	67%,
(n=67)				75%		92%	(10/18)	78%		91%

Cohort	AUC	Cut value	Sensitivity	95%	Specificity	95%	PPV	95%	NPV	95% CI
				CI		CI		CI		
Training set (n=49)	0.913	0.121	88% (22/25)	68%,	88% (21/24)	67%,	88%	68%,	88% (21/24)	67%,
				97%		97%	(22/25)	97%		97%
Validation set	0.761	0.121	57%	30%,	79% (15/19)	54%,	67% (8/12)	35%,	71% (15/21)	48%,
(n=33)			(8/14)	81%	, ,	93%	,	89%		88%

Table S5: Coefficients of variation for the multivariate (upper table) and single variable (lower table) models for post-transplant training set samples.

IR1+IRx Training full model	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-1.573	0.852	-1.848	0.065
Race	1.561	0.812	1.923	0.054
IR	3.413	0.930	3.672	2.41E-04
Organ.1	-1.397	0.491	-2.846	0.004
Time between Transplant and sample days	-0.001	0.000	-1.861	0.063
FK level-day of sample	0.223	0.078	2.872	0.004

IR1+IRx Training full model	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-1.191	0.304	-3.913	9.11E-05
IR	3.306	0.766	4.317	1.58E-05

Effect of confounders: Comparable test performance within the range seen in training and validation set samples was also seen in samples sub-grouped by the time of sampling with respect to transplantation, type of organ transplanted, type of induction, whether actual or surrogate donor stimulators were used, and whether rejection or non-rejection were diagnosed by "for-cause" or surveillance biopsy or clinically (SDC, Tables 6-10 in Supplemental Digital Content: Tables). Performance estimates are less likely to be meaningful for those subgroups with small numbers of samples.

Table S6: General test performance stratified by time after transplantation for training and validation set. R=rejector, NR=Non-rejector, AUC=Area under the receiver-operating characteristic curve, Sens=sensitivity, Spec=specificity, LCI=lower confidence interval, UCI=upper confidence interval, PPV=positive predictive value, NPV=negative predictive value.

Dataset	Total (n)	R (n)	NR (n)	AUC	Sens n	95% LCI	95% UCI	Spec n	95% LCI	95% UCI	PPV n	95% LCI	95% UCI	NPV n	95% LCI	95% UCI
IR0 Validation	33	14	19	0.842	57.1 8/14	29.6	81.2	89.5 17/19	65.5	98.2	80 8/10	44.2	96.5	73.9 17/23	51.3	88.9
IR0 Training	49	25	24	0.82	80 20/25	58.7	92.4	70.8 17/24	48.8	86.6	74.1 20/27	53.4	88.1	77.3 17/22	54.2	91.3
IR1&IRx Training	98	24	74	0.878	91.7 22/24	71.5	98.5	83.86 2/74	73	91	64.7 22/34	46.5	79.7	96.9 62/64	88.2	99.5
IR1&IRx Validation	64	19	45	0.791	84.2 16/19	59.5	95.8	8036/ 45	64.9	89.9	6416/ 25	42.6	81.3	92.3 36/39	78	98
IR1 Training	48	17	31	0.841	88.2 15/17	62.3	97.9	80.6 25/31	61.9	91.9	71.4 15/21	47.7	87.8	92.6 25/27	74.2	98.7
IR1 Validation	30	15	15	0.796	80 12/15	51.4	94.7	86.7 13/15	58.4	97.7	85.7 12/14	56.2	97.5	81.2 13/16	53.7	95
IRx Training	50	7	43	0.95	100 7/7	56.1	100	86 37/43	71.4	94.2	53.87/ 13	26.1	79.6	100 37/37	88.3	100
IRx Validation	34	4	30	0.858	100 4/4	39.6	100	76.7 23/30	57.3	89.4	36.44/ 11	12.4	68.4	100 23/23	82.2	100

Table S7: Test performance stratified by type of donor stimulator, whether actual donor or surrogate donor. AUC=Area under the receiver-operating characteristic curve, Sens=sensitivity, Spec=specificity PPV=positive predictive value, NPV=negative predictive value.

Dataset	Donor cell type	Total n	Rejector n	Non- rejector n	AUC	Sens	Spec	PPV	NPV
IR0 Training	Surrogate Donor	40	20	20	0.87	90	70	75	87.5
IR0 Validation	Surrogate Donor	31	14	17	0.868	57.1	88.2	80	71.4
IR1&IRx Training	Surrogate Donor	80	19	61	0.847	89.5	80.3	58.6	96.1
IR1&IRx Validation	Surrogate Donor	59	17	42	0.817	88.2	81	65.2	94.4
IR0 Training	Actual Donor	9	5	4	0.75	40	75	66.7	50
IR0 Validation	Actual Donor	2	0	2	NA	NA	100	NA	100
IR1&IRx Training	Actual Donor	18	5	13	1	100	100	100	100
IR1&IRx Validation	Actual Donor	5	2	3	0.5	50	66.7	50	66.7

Table S8: Test performance stratified by organ type. L=Liver, SB=small bowel or intestine, LSB=combined liver-intestine, LK=combined liver-kidney, LL=liver-lung. AUC=area under the receiver-operating –characteristic curve, sens=sensitivity, spec=specificity, PPV=positive predictive value, NPV=positive predictive value, NA=could not be calculated, n=number.

Dataset	Organ	Total n	Rejector n	Non-rejector n	AUC	Sens	Spec	PPV	NPV
IR0 Training	L	25	14	11	0.89	85.7	90.9	92.3	83.3
IR0 Validation	L	26	11	15	0.897	63.6	86.7	77.8	76.5
IR1&IRx Training	L	68	18	50	0.905	94.4	88	73.9	97.8
IR1&IRx Validation	L	54	17	37	0.755	82.4	75.7	60.9	90.3
IR0 Training	LSB	11	7	4	0.75	71.4	75	83.3	60
IR0 Validation	LSB	1	1	0	NA	0	NA	NA	0
IR1&IRx Training	LSB	15	3	12	0.833	66.7	75	40	90
IR1&IRx Validation	LSB	2	1	1	1	100	100	100	100
IR0 Training	LL	0	NA	NA	NA	NA	NA	NA	NA
IR0 Validation	LL	0	NA	NA	NA	NA	NA	NA	NA
IR1&IRx Training	LL	0	NA	NA	NA	NA	NA	NA	NA
IR1&IRx Validation	LL	1	0	1	NA	NA	100	NA	100
IR0 Training	SB	10	2	8	0.875	100	50	33.3	100
IR0 Validation	SB	6	2	4	1	50	100	100	80
IR1&IRx Training	SB	13	2	11	0.955	100	72.7	40	100
IR1&IRx Validation	SB	7	1	6	1	100	100	100	100
IR0 Training	LK	3	2	1	0	50	0	50	0
IR0 Validation	LK	0	NA	NA	NA	NA	NA	NA	NA
IR1&IRx Training	LK	2	1	1	1	100	100	100	100
IR1&IRx Validation	LK	0	NA	NA	NA	NA	NA	NA	NA

Table S9: Test performance stratified by type of induction. None=no induction, thymo=thymoglobulin or rabbit anti-human thymocyte globulin, campath=alemtuzumab.

Sens=sensitivity, Spec=specificity PPV=positive predictive value, NPV=negative predictive value

Dataset	Induction Type	Total n	Rejector n	Non- rejector n	AUC	Sens	Spec	PPV	NPV
IR0 Training	None	10	6	4	0.875	83.3	100	100	80
IR0 Validation	None	13	6	7	0.952	50	100	100	70
IR1&IRx Training	None	33	4	29	0.935	100	89.7	57.1	100
IR1&IRx Validation	None	29	10	19	0.787	80	84.2	72.7	88.9
IR0 Training	Thymo	29	15	14	0.786	73.3	64.3	68.8	69.2
IR0 Validation	Thymo	19	8	11	0.807	62.5	81.8	71.4	75
IR1&IRx Training	Thymo	54	17	37	0.859	88.2	81.1	68.2	93.8
IR1&IRx Validation	Thymo	32	9	23	0.78	88.9	73.9	57.1	94.4
IR0 Training	Campath	10	4	6	0.958	100	66.7	66.7	100
IR0 Validation	Campath	1	0	1	NA	NA	100	NA	100
IR1&IRx Training	Campath	11	3	8	0.917	100	75	60	100
IR1&IRx Validation	Campath	3	0	3	NA	NA	100	NA	100

Table S10: Test performance stratified by whether outcome was determined without biopsy (no Bx), with for cause Bx or with surveillance Bx. Sens=sensitivity, Spec=specificity PPV=positive predictive value, NPV=negative predictive value

Dataset	Bx type	Total n	Rejector n	Non- rejector n	AUC	Sens	Spec	PPV	NPV
IR0 Training	no Bx	11	2	9	0.889	100	77.8	50	100
IR0 Validation	no Bx	12	1	11	1	100	81.8	33.3	100
IR1&IRx Training	no Bx	57	2	55	0.932	100	89.1	25	100
IR1&IRx Validation	no Bx	39	1	38	0.934	100	78.9	11.1	100
IR0 Training	for cause Bx	17	13	4	0.904	84.6	100	100	66.7
IR0 Validation	for cause Bx	11	10	1	0.7	60	100	100	20
IR1&IRx Training	for cause Bx	20	15	5	0.88	93.3	80	93.3	80
IR1&IRx Validation	for cause Bx	13	13	0	NA	76.9	NA	100	NA
IR0 Training	Surveillance Bx	21	10	11	0.768	70	54.5	58.3	66.7
IR0 Validation	Surveillance Bx	10	3	7	0.714	33.3	100	100	77.8
IR1&IRx Training	Surveillance Bx	21	7	14	0.791	85.7	64.3	54.5	90
IR1&IRx Validation	Surveillance Bx	12	5	7	0.971	100	85.7	83.3	100