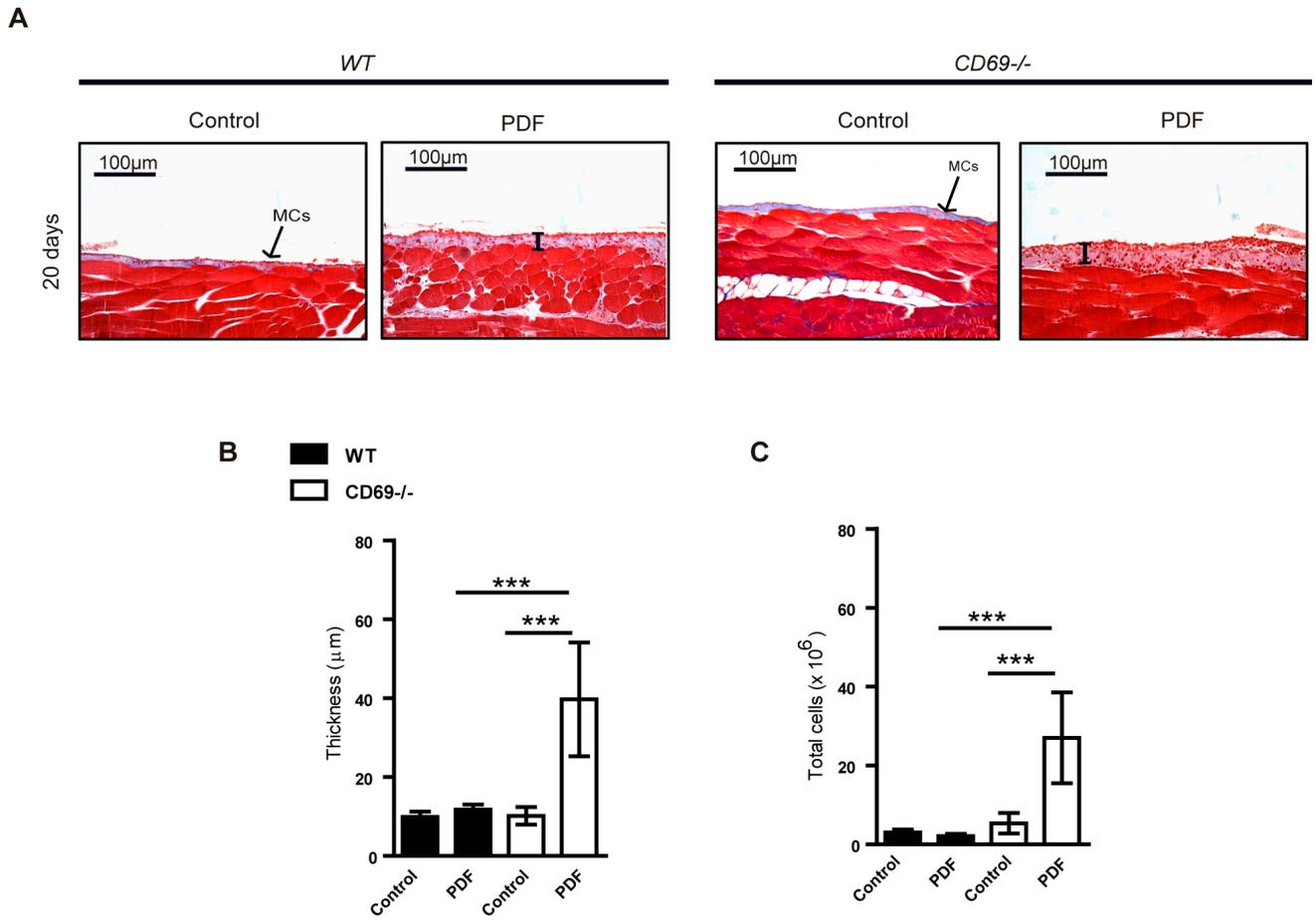


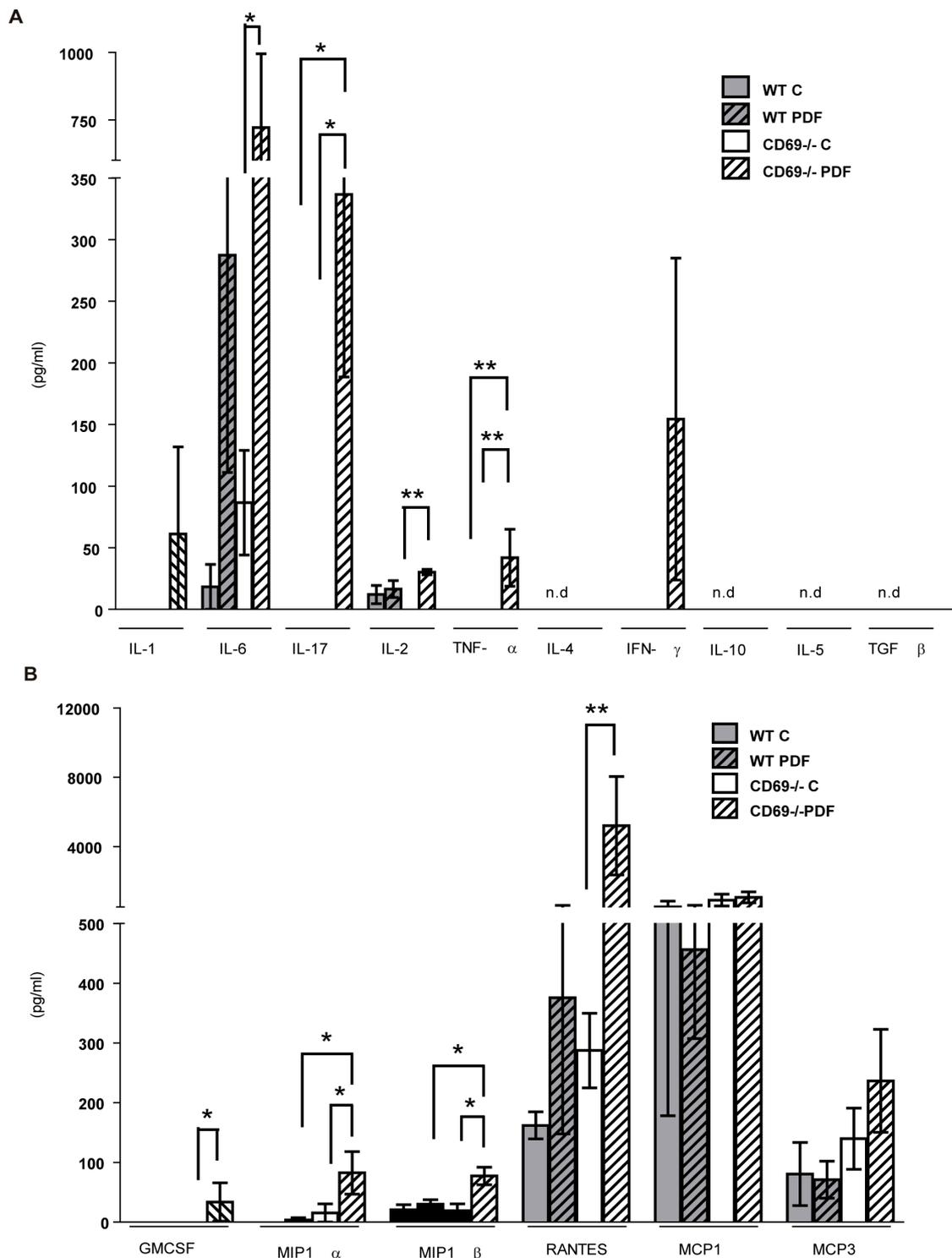
**Supplementary Table S1:** Flow cytometry analysis of the percentage of different cell populations in the peritoneal membrane tissue of WT and *cd69*<sup>-/-</sup> mice after 40 days of treatment with PDF or saline as control. Data are means ± SEM.

(%)	WT		CD69 <sup>-/-</sup>	
	Control	PDF	Control	PDF
CD11b <sup>+</sup> Gr-1 <sup>+</sup> (monocytes)	0.05 ± 0.02	0.18 ± 0.09	0.05 ± 0.01 @	0.21 ± 0.05 @
CD11b <sup>+</sup> Gr-1 <sup>high</sup> (neutrophils)	0.04 ± 0.01	0.20 ± 0.11	0.03 ± 0.01 £	0.34 ± 0.07 £
B220 <sup>+</sup> (B cells)	8.12 ± 1.87 £	20.0 ± 2.21 £, \$	9.26 ± 1.19 @	4.99 ± 0.98 @, \$
cDCs (c dendritic cells)	0.04 ± 0.01 @	0.12 ± 0.03 @	0.03 ± 0.01	0.10 ± 0.02
pDCs (p dendritic cells)	0.57 ± 0.19	1.68 ± 0.60	0.32 ± 0.08 &	1.97 ± 0.40 &
CD4 <sup>+</sup> (T cells)	0.04 ± 0.02 @	0.26 ± 0.08 @	0.04 ± 0.01 &	0.21 ± 0.04 &
CD8 <sup>+</sup> (T cells)	0.04 ± 0.02 &, *	0.18 ± 0.04 &, *	0.05 ± 0.01 &, ©	0.14 ± 0.02 &, ©

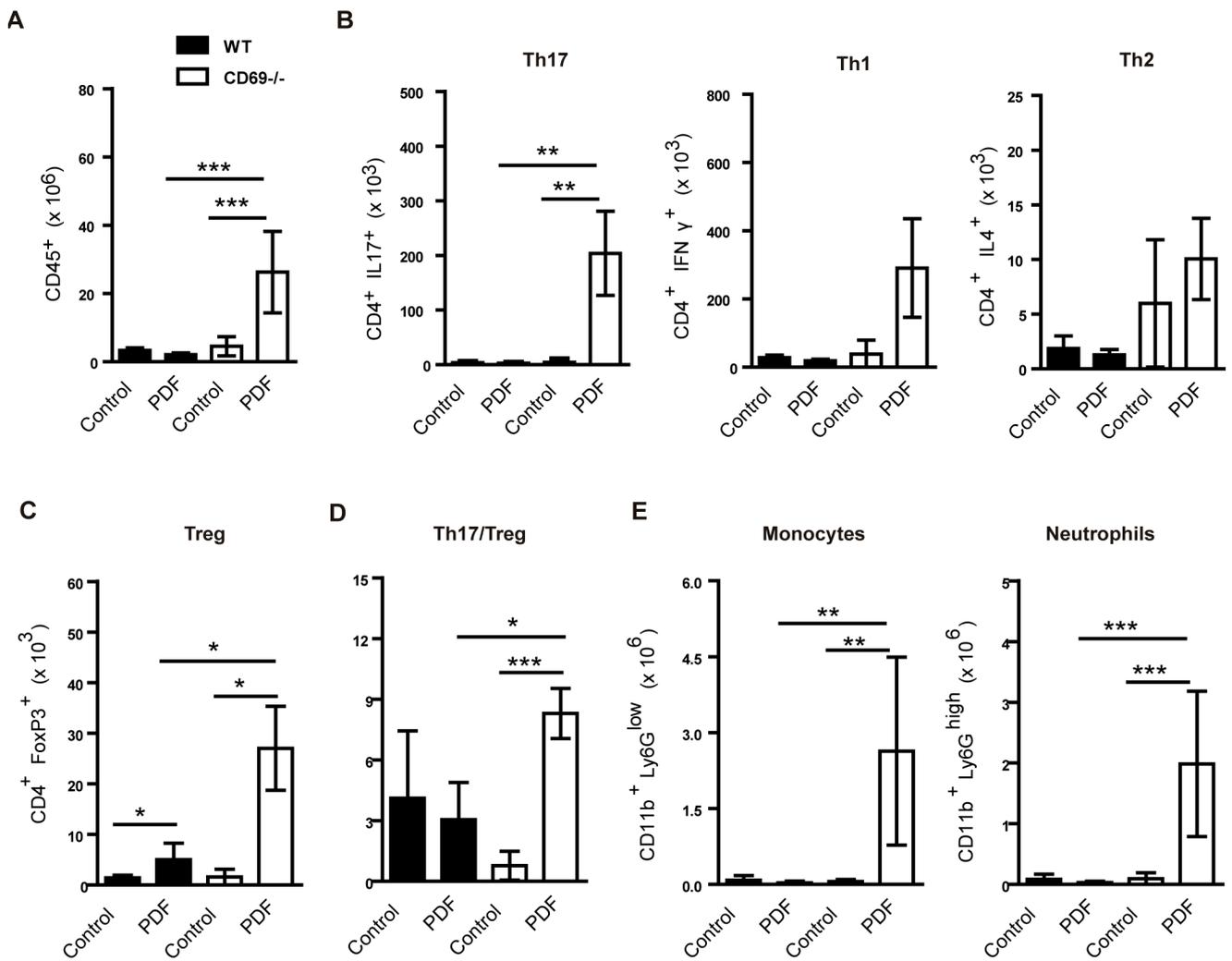
@: p < 0.05; &: p < 0.01; £: p < 0.001; \$: p < 0.001: WT PDF vs. *cd69*<sup>-/-</sup> PDF; \*: p < 0.01: WT Control vs. WT PDF, ©: p < 0.01: CD69<sup>-/-</sup> Control vs. *cd69*<sup>-/-</sup> PDF



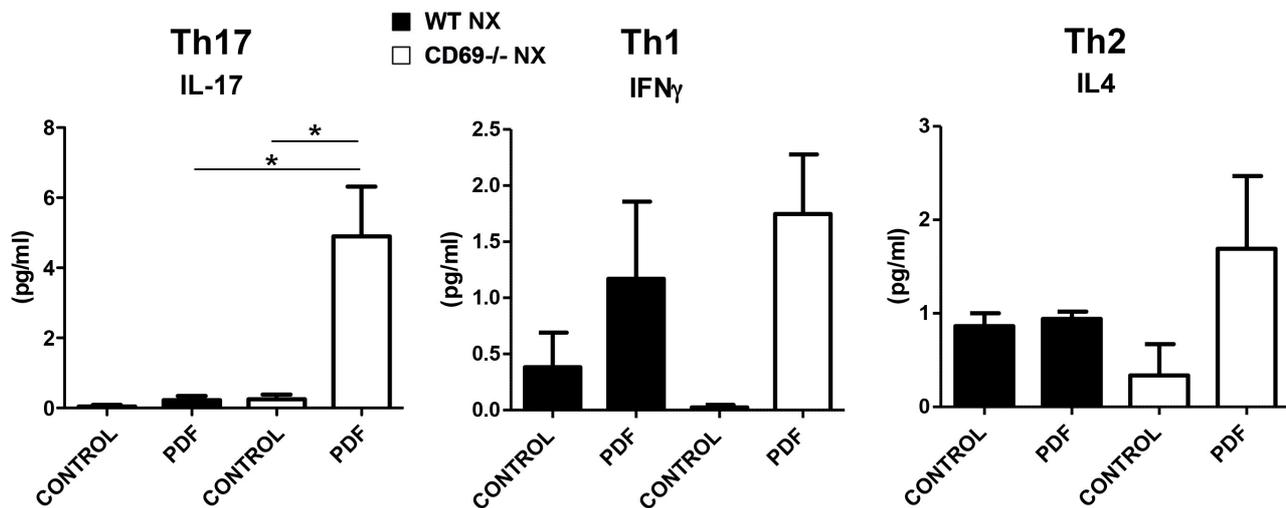
**Supplementary Figure S1:** PDF treatment of WT and *cd69*<sup>-/-</sup> mice (20 days). **(A)** Trichrome Masson's staining of peritoneal tissue in *cd69*<sup>-/-</sup> and WT mice treated or not with PDF. **(B)** Peritoneal membrane thickness. **(C)** Total cell counts (x 10<sup>6</sup>) in the peritoneal cavity effluents. Data are means ± SD (n≥5). P values p < 0.05 are considered statistically significant one-way Anova test. \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001.



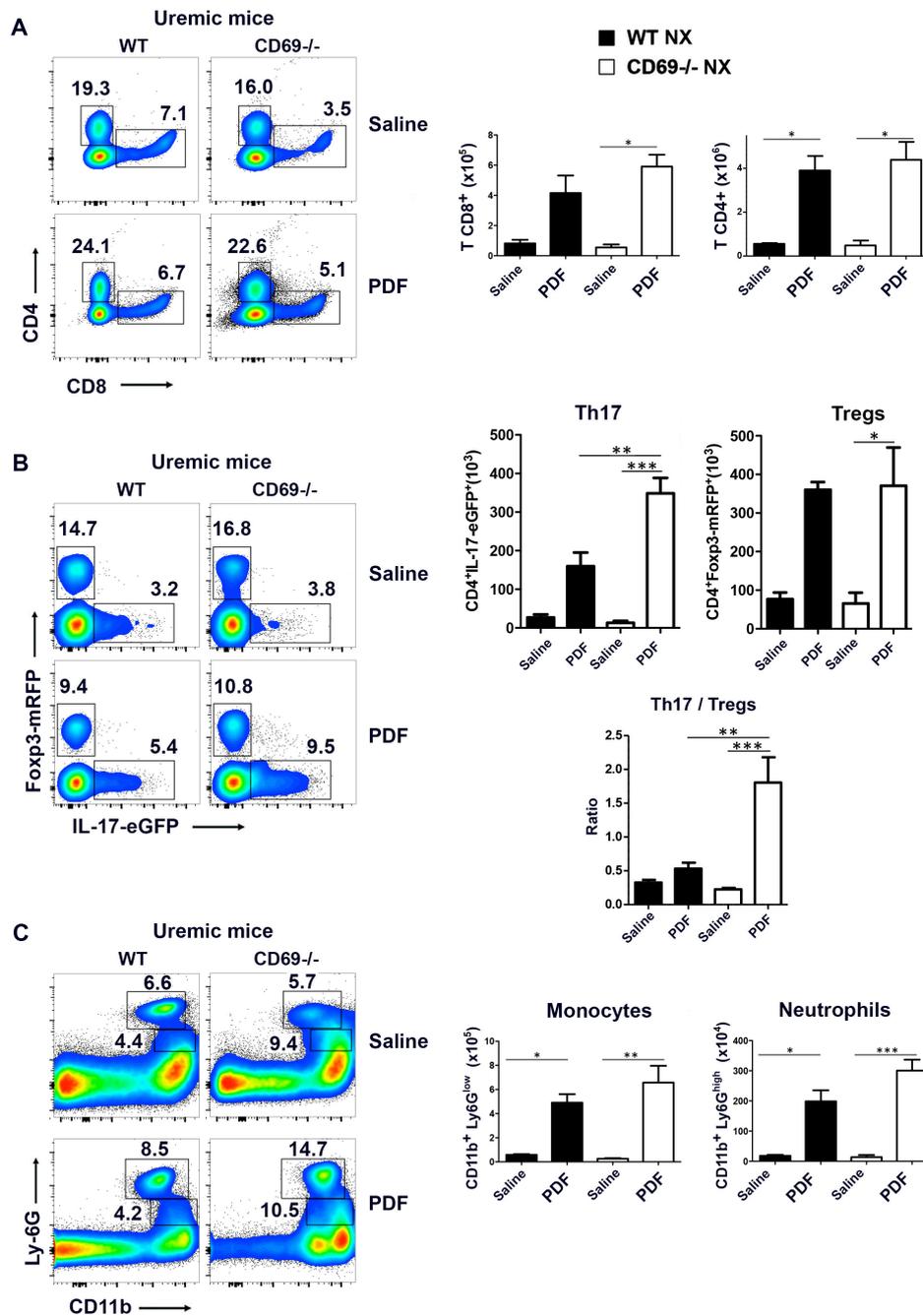
**Supplementary Figure S2:** Analysis of (A) cytokine and (B) chemokine profiles released in the peritoneal cavity after 20 days of PDF treatment. Production of pro-inflammatory cytokines and chemokines was assessed by Flow Cytomix cytokine array and analyzed by FACS. Bars are means  $\pm$  SD ( $n \geq 5$ ). P values  $p < 0.05$  are considered statistically significant using one-way Anova test. \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ .



**Supplementary Figure S3:** (A-C) Quantification of flow-cytometry analysis of the expression of total number of (A) CD45<sup>+</sup>, (B) Th17, Th1 and Th2 and (C) Treg cells after 20 days treatment with saline or PDF. (D). Ratio between the total Th17 and regulatory T cells in all groups. (E) CD11b<sup>+</sup>Ly6G<sup>low</sup> (monocytes) and CD11b<sup>+</sup>Ly6G<sup>high</sup> (neutrophils) in the peritoneal effluents from indicated groups. P values p <0.05 are considered statistically significant using one-way Anova test. \*p <0.05, \*\*p <0.01, \*\*\*p <0.001.

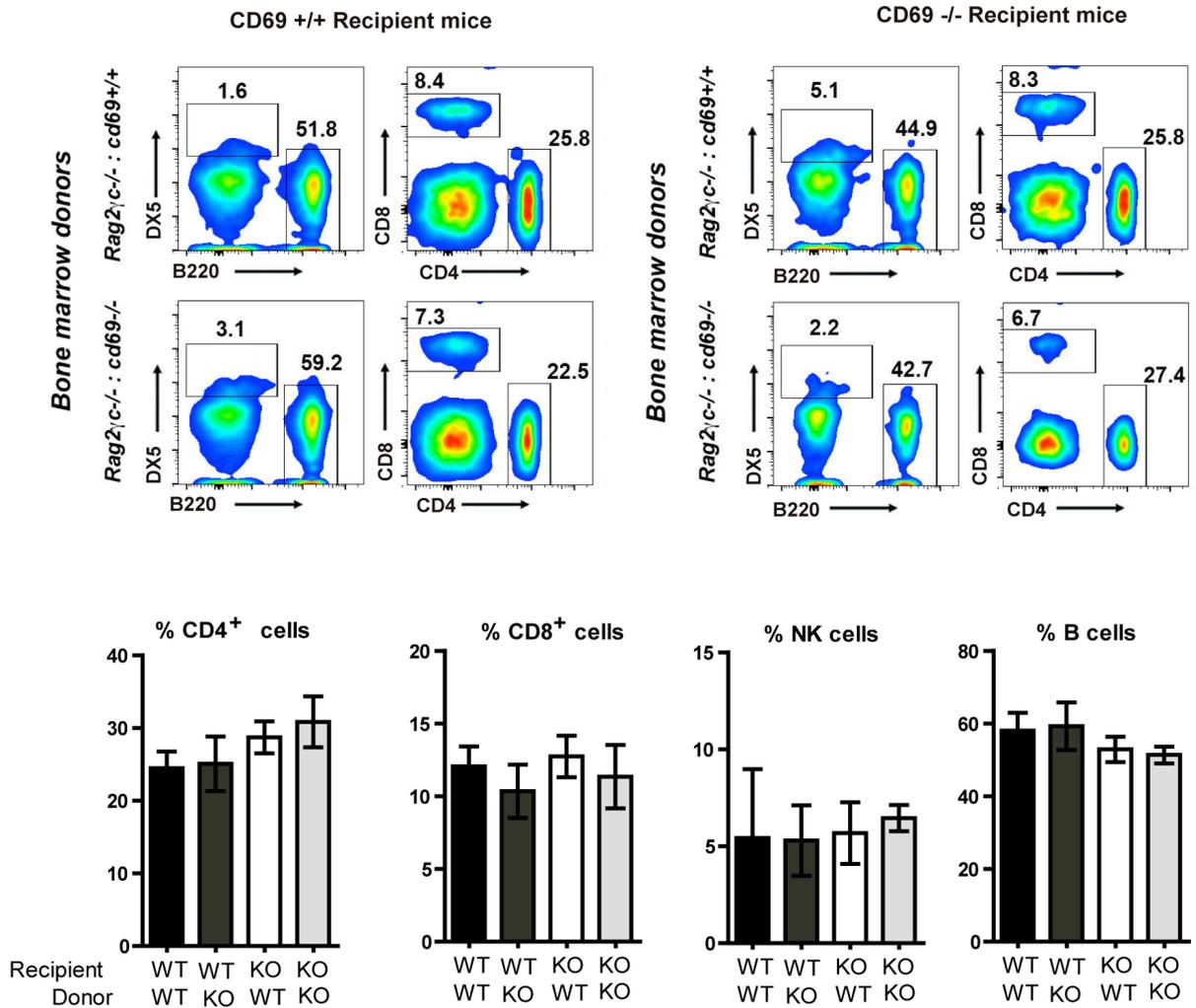


**Supplementary Figure S4.** Analysis of cytokine expression in the peritoneal cavity of uremic WT and *cd69*<sup>-/-</sup> mice. Mice undergo 5/6 nephrectomy before treatment with saline or PDF for a period of 40 days. Production of T helper-associated cytokines was assessed by multiplexed flow cytometric bead array. Bars are means  $\pm$  SD ( $n \geq 6$ ). P values  $p < 0.05$  were considered statistically significant using one-way Anova test, and Bonferroni post-test were used to compare selected pairs of means and all pairs of means, respectively (\* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ ).



**Supplementary Figure S5.** Density plots of flow-cytometry analysis in peritoneal effluents from uremic *cd69*<sup>-/-</sup> double reporter (*cd69*<sup>-/-</sup>-dRep) mice (Foxp3-mRFP in *foxp3* locus and IL-17A-eGFP in *Il17a* locus) or wt littermates (*cd69*<sup>+/+</sup>-dRep). (A) Total numbers of CD8<sup>+</sup> and CD4<sup>+</sup> T cells were assessed in mice treated during 40 days with saline or PDF. (B) Analysis of CD4<sup>+</sup>FoxP3-RFP (Tregs) and IL-17eGFP from indicated groups. The Th17/Treg ratio is shown. (C) CD11b<sup>+</sup>Ly6G<sup>low</sup> (monocytes) and CD11b<sup>+</sup>Ly6G<sup>high</sup> (neutrophils) quantification in peritoneal effluents. P values  $p < 0.05$  are considered statistically significant using one-way Anova test. \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ .

PBL from chimeric mice



**Supplementary Figure S6:** WT and *cd69*<sup>-/-</sup> recipient mice were transplanted with a mixture of BM cells from Rag2<sup>-/-</sup>;  $\gamma$ c<sup>-/-</sup> and *cd69*<sup>-/-</sup> or Rag2<sup>-/-</sup>;  $\gamma$ c<sup>-/-</sup> and *cd69*<sup>+/+</sup> (WT) cells as described in the Material and Methods section. In the following graphs recipient mice are WT or KO for CD69 alone and donor mice are WT or KO mixed with Rag2<sup>-/-</sup>;  $\gamma$ c<sup>-/-</sup> cells. Flow-cytometry analysis of CD4<sup>+</sup>, CD8<sup>+</sup>, NK (DX5<sup>+</sup>) and B (B220<sup>+</sup>) cells in peripheral blood (PBL) samples in reconstituted mice before the initiation of PDF treatment. (n $\geq$ 8). No statistically significant differences were found using one-way Anova test.