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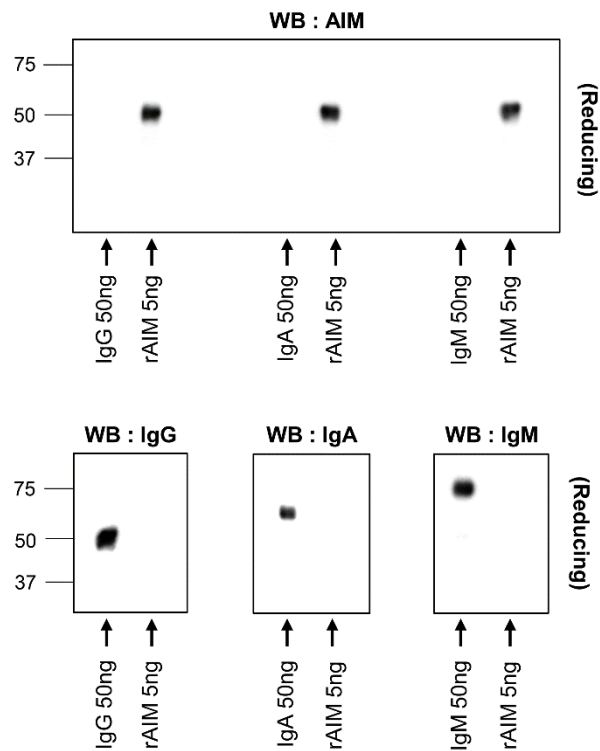


Figure S1. No crossreactivity of anti-AIM with immunoglobulins

Commercial murine IgG, IgA, and IgM were loaded for SDS-PAGE and immunoblotting with anti-AIM, anti-IgG, anti-IgA, and anti-IgM under reducing conditions was performed. No immunoglobulin was detected by anti-AIM.

IgG from mouse serum (15381-1MG; SIGMA), IgA, Kappa from murine myeloma (M1421; SIGMA), and rAIM (purified as previously described²²) were used. Monoclonal IgM protein was purified using the HiTrap IgM purification column (17511001; GE Healthcare Life Sciences) from the culture supernatant of B-cell hybridoma (clone TC-31, CRL-2494; American Type Culture Collection, Manassas, VA, USA) according to the manufacturer's protocol.

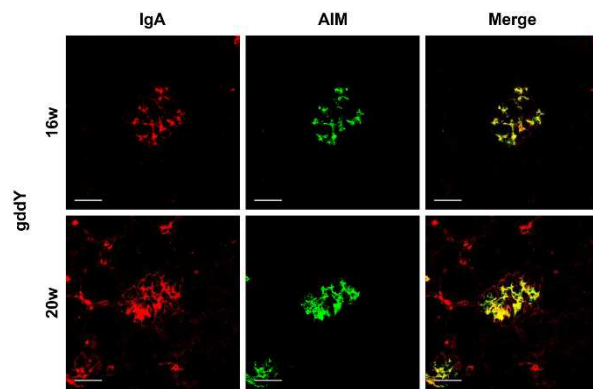


Figure S2. AIM expression in gddY mice

Glomerular AIM co-deposition with IgA and expression of AIM in mesangial cells were examined at different ages. Representative immunostaining images of IgA (red) and AIM (green) in the renal tissue of 16-and 20-week-old gddY and *AIM*^{-/-}gddY (n = 5). All samples were stained and observed with the samples shown in Figure 1A. Scale bars: 50 μ m.

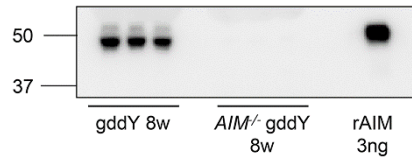


Figure S3. *AIM*^{-/-}gddY mouse model establishment using the CRISPR/Cas9 system

To confirm the knockout of AIM in *AIM*^{-/-}gddY mice, we performed immunoblotting.

Representative image of immunoblotting for AIM, using reducing conditions, in serum from

8-week-old gddY and *AIM*^{-/-}gddY mice.

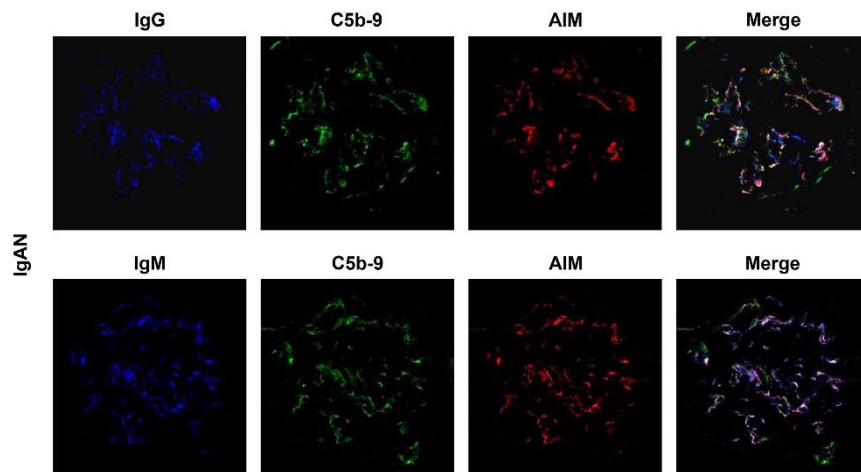


Figure S4. C5b-9 co-deposits with IgG/IgM and AIM in IgAN

Representative immunostaining images of C5b-9 (green) co-deposition with IgG (blue)/AIM (red) and IgM (blue)/AIM (red) (n = 4). Scale bars: 50 μ m.

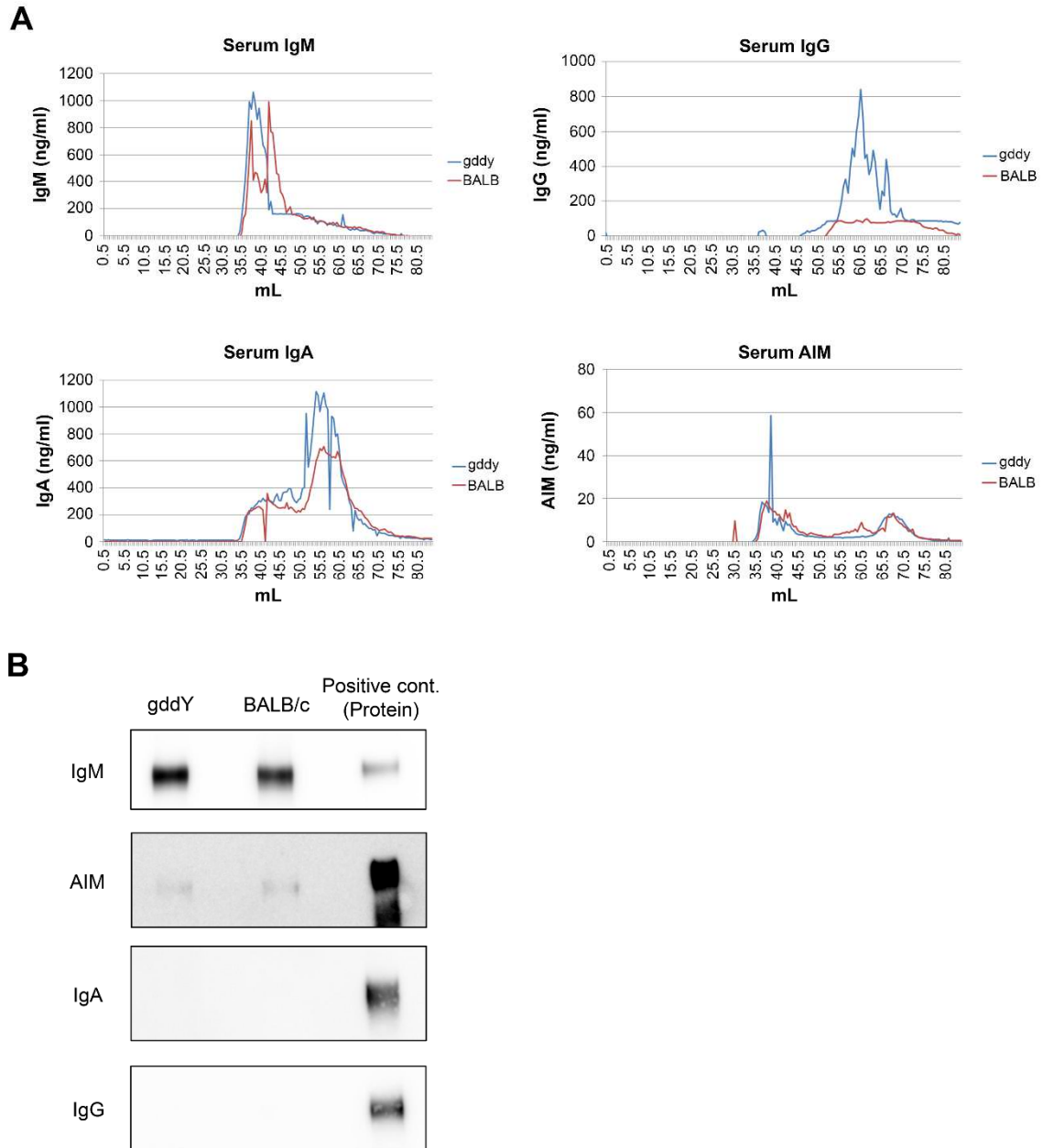


Figure S5. No associations of IgA and IgM containing AIM were observed in serum fractionation samples from gddy or BALB/c mice.

(A) Size fractionation on serum from gddy and BALB/c mice was performed, and each fraction was examined for the presence of IgM, IgA, IgG, and AIM using ELISA. Sera were subjected to

size-exclusion chromatography using the HiPrep 16/60 Sephacryl S-300 HR column (17116701, GE Healthcare Life Sciences) with Dulbecco's PBS. AIM was measured by ELISA using two different rat anti-mouse AIM monoclonal antibodies (rat IgG, clones #36 and #35; generated in our laboratory). Regarding IgM, IgA and IgG, we used an ELISA Quantification Set (IgM: E90-101, IgA: E90-103, IgG: E90-131; Bethyl Laboratories), and we performed assays according to the manufacturer's protocol.

(B) The IgM peak fractions at 39–39.5 ml (10 μ l each) were immunoblotted for IgM, IgA, IgG, and AIM under reducing conditions. We used antibodies for immunoblotting as described in the Methods section.

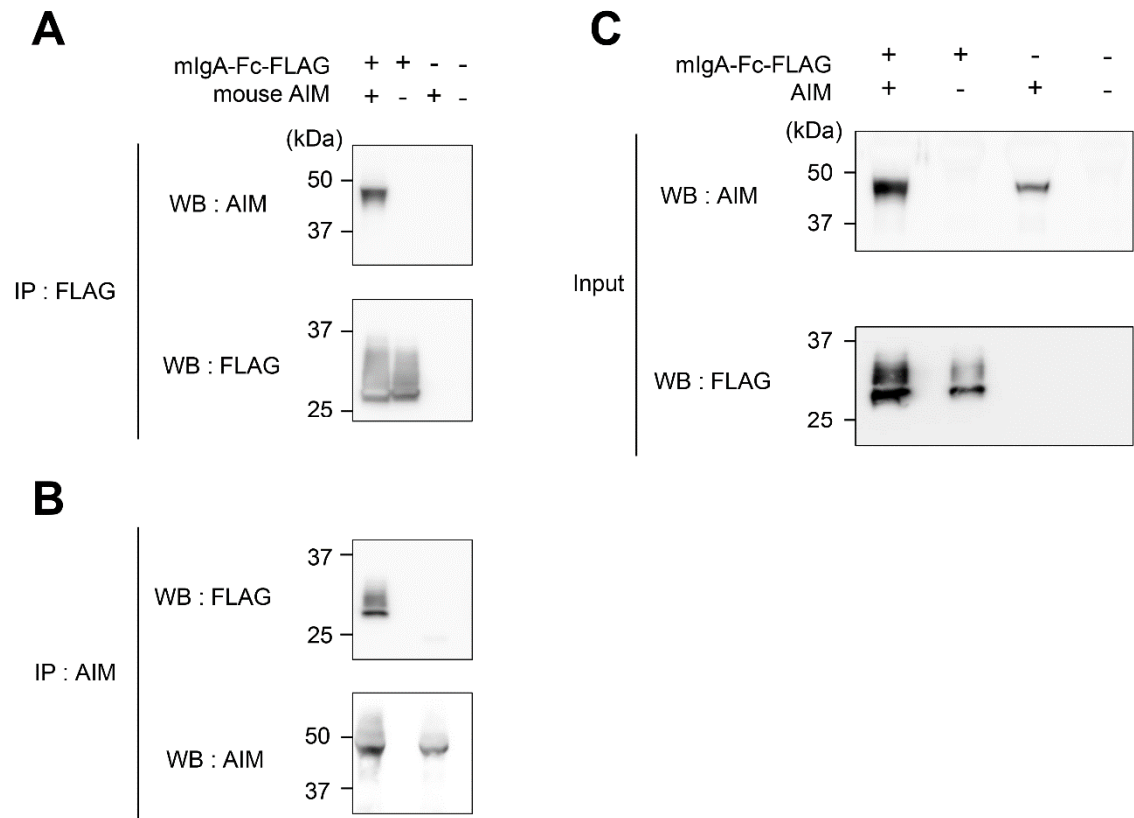


Figure S6. AIM associates with IgA-Fc in mice and humans.

HEK293T cells overexpressing mouse AIM and those overexpressing FLAG-tagged mouse

IgA-Fc were co-cultured for 48 h, and supernatants were prepared from co-cultures.

Immunoblotting of the supernatant for anti-AIM and IgA were performed (C). The supernatants

were incubated with an anti-AIM antibody conjugated with protein G Sepharose

(Sigma-Aldrich, P3296) or anti-FLAG M2 affinity gel (Sigma-Aldrich, A2220) at 4°C

overnight. The precipitates were washed five times with a wash buffer (TBS plus 0.1%

Tween-20) and eluted by heating at 95°C for 5 min in SDS sample buffer containing

2-mercaptoethanol. Association was assessed by immunoblotting for IgA-Fc (A) and AIM (B).

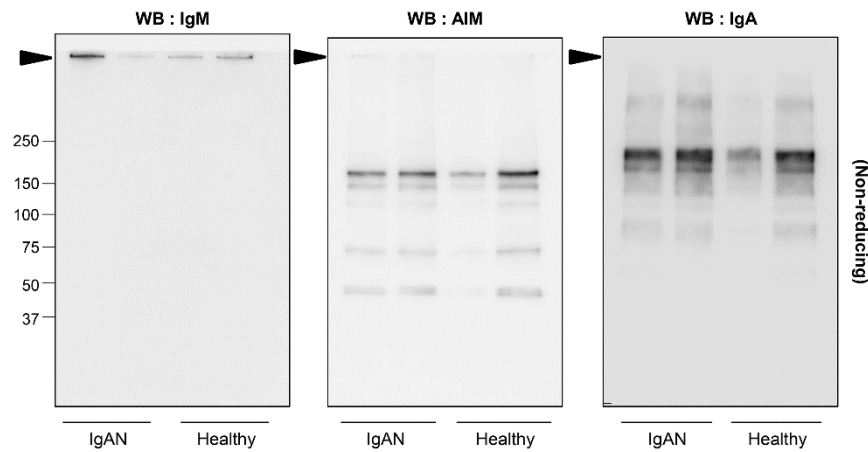


Figure S7. No IgA/AIM/IgM immune complex was observed in serum samples from patients with IgAN.

Immunoprecipitation was performed on sera from human IgAN and healthy control using anti-IgA and the immune complex was analyzed by immunoblotting with anti-IgM (left), AIM (middle), and IgA (right) under nonreducing conditions ($n = 2$ for each group). Methods and materials are described in the Methods section.