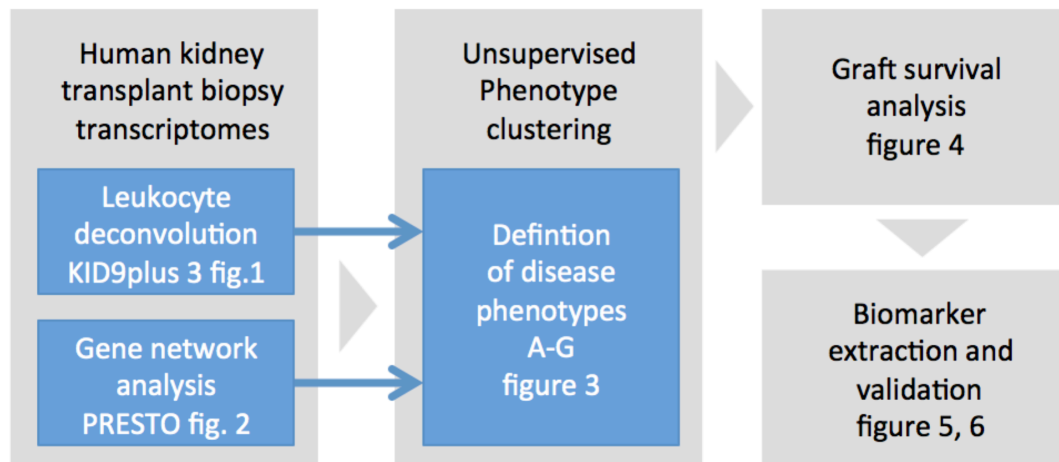
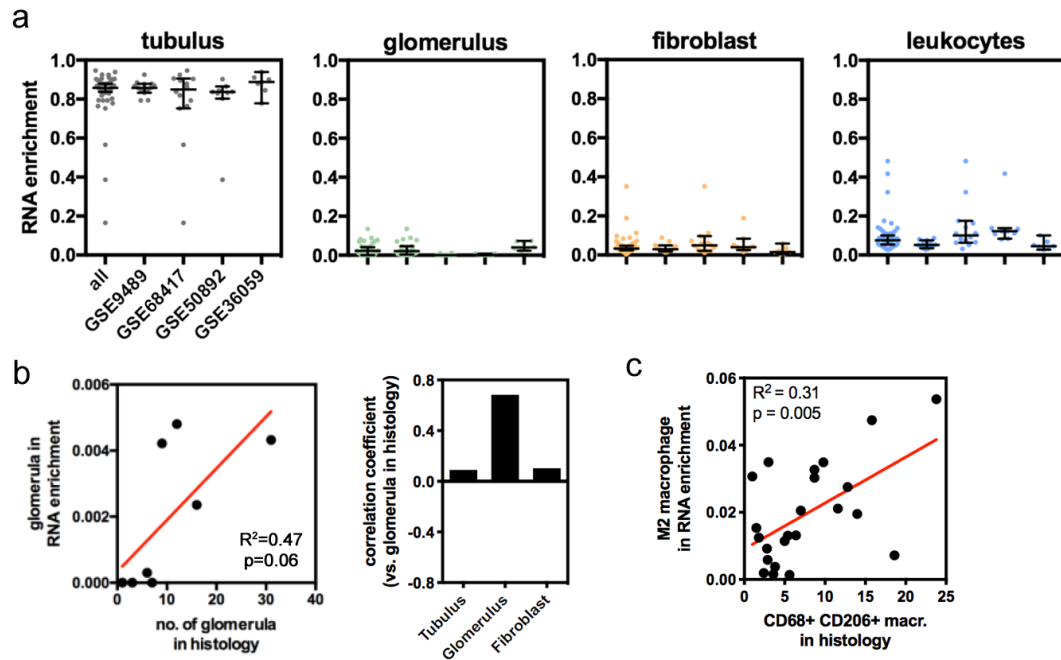


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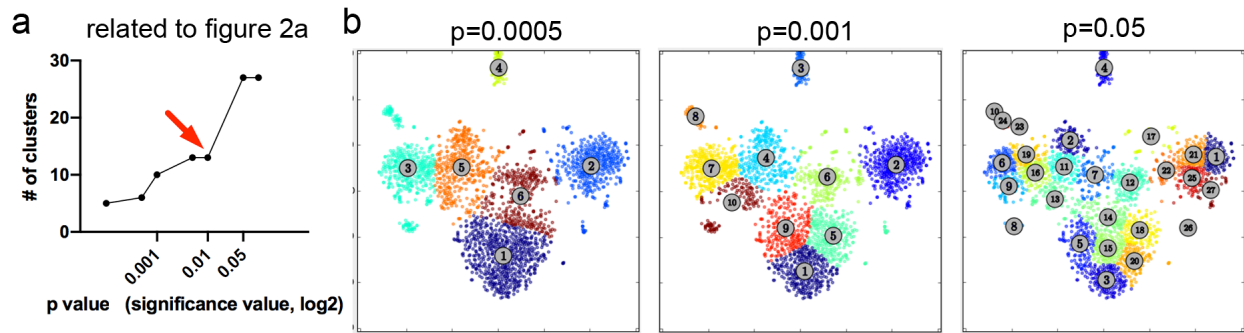
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**Supplemental figure 1: Workflow overview.**

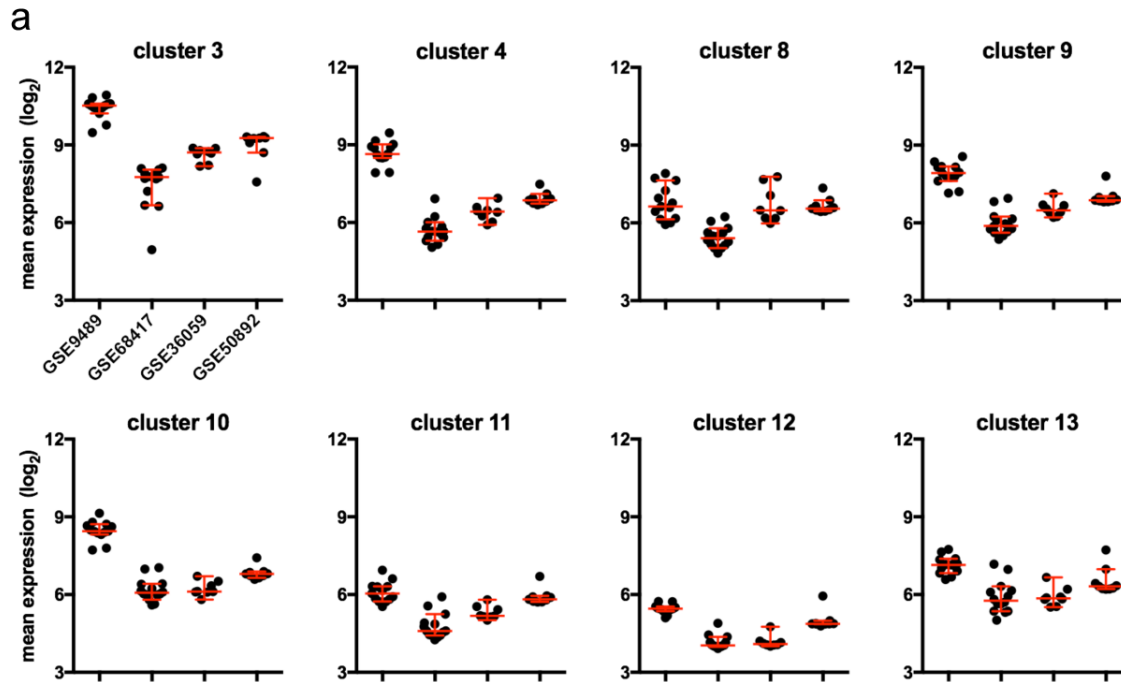


**Supplemental figure 2: Validation of leukocyte deconvolution in independent data sets of healthy kidney transplant biopsies.** a) The KID9plus3 signature was used to deconvolute GSE36059 (only nephrectomy samples, n=7) and three independent kidney biopsy data sets (GSE9489 n=13, GSE68417 only nephrectomy samples n=14, GSE50892 only normal kidney biopsies n= 9). The enrichment score for tubules, glomeruli, fibroblasts and leukocytes are shown for all data sets. b) The number of glomeruli as counted in histological sections was compared to the glomerulus/tubule/fibroblast enrichment score of KID9plus3 of paired transcriptome samples (GSE50892). c) The KID9plus3 M2 macrophage enrichment score was compared to the number of CD68+ CD206+ macrophages of paired transcriptome samples (GSE65326).

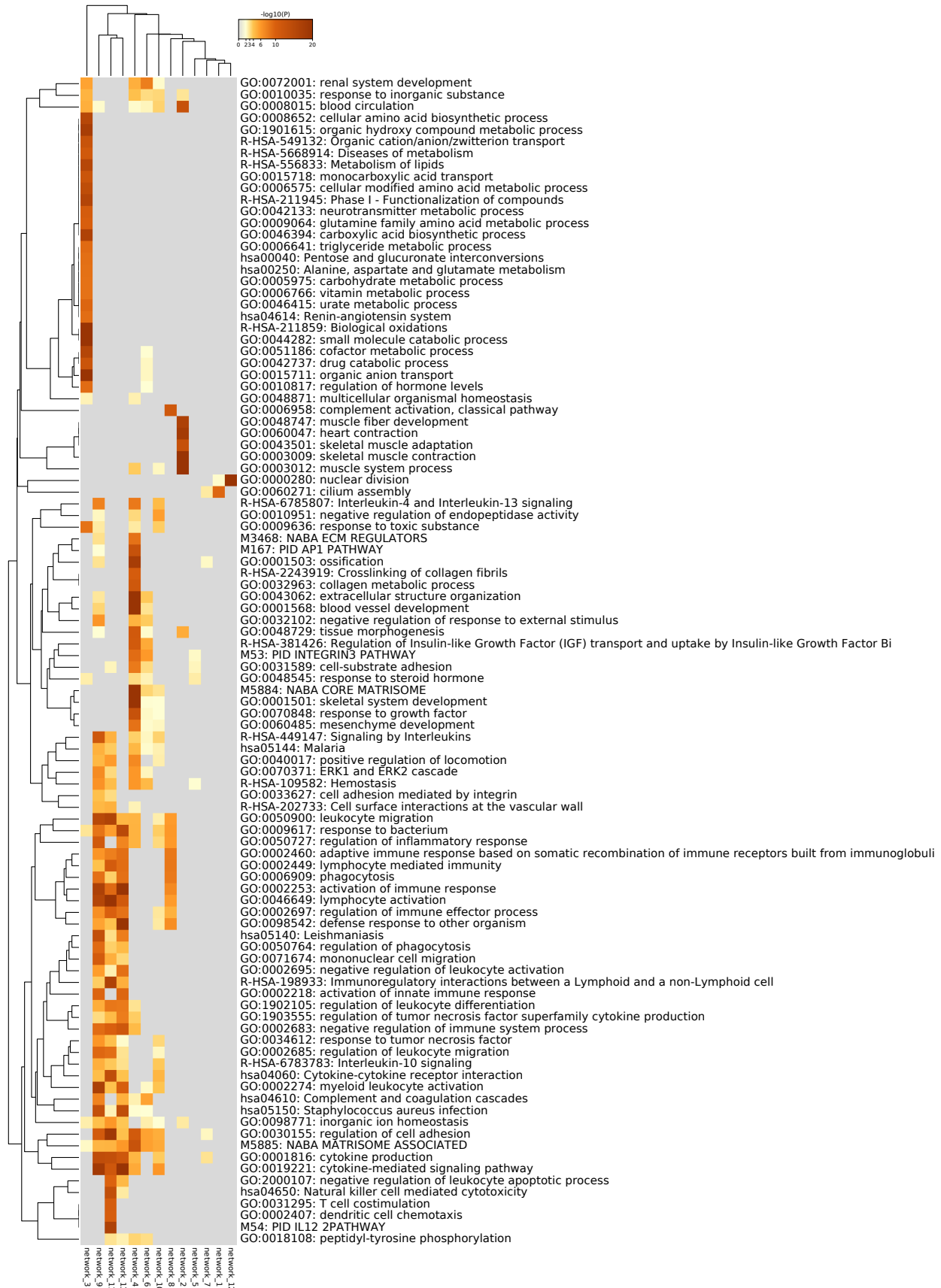


**Supplemental figure 3: Cluster sizes depending on user input.** a,b) Number of identified PRESTO clusters depending on the significance level in a modified k-means clustering algorithm (see materials and methods). The value of  $k=0.01$  (red arrow) was chosen in manuscript figure 2a.

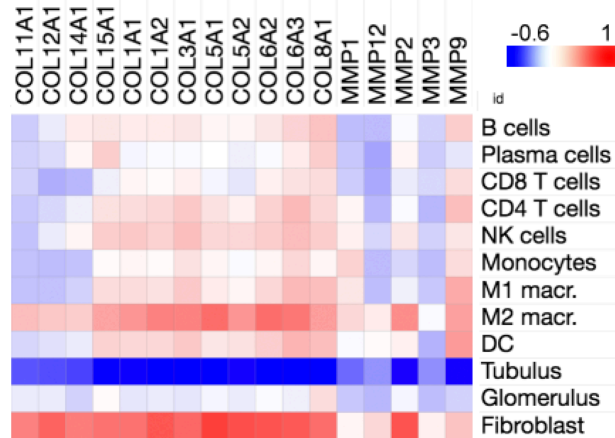




**Supplemental figure 4: Gene network analysis in independent data sets of healthy kidney transplant biopsies.** Network activity (mean of gene expression per network) for each PRESTO gene network (see main figure 2) was compared across independent data sets. Each dot represents one sample/biopsy. Mean and standard deviation are indicated. (GSE9489 n=13, GSE68417 only nephrectomy samples n=14, GSE50892 only normal kidney biopsies n= 9; see materials and methods).



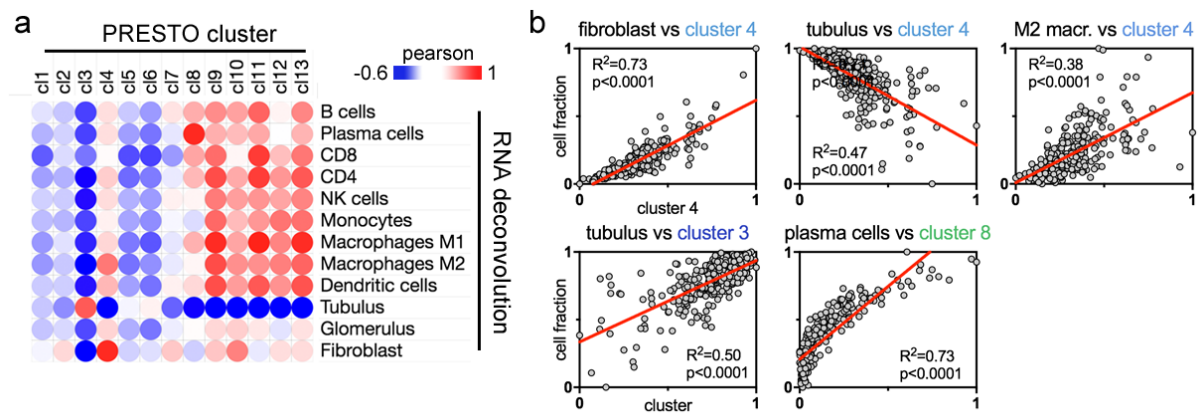
**Supplemental figure 5: Functional annotation of gene networks.** Metascape was used to compare significant functional annotations in PRESTO gene networks 1-13 (shown in figure 2). The color codes for the p-value ( $-\log_{10}(p)$ ).



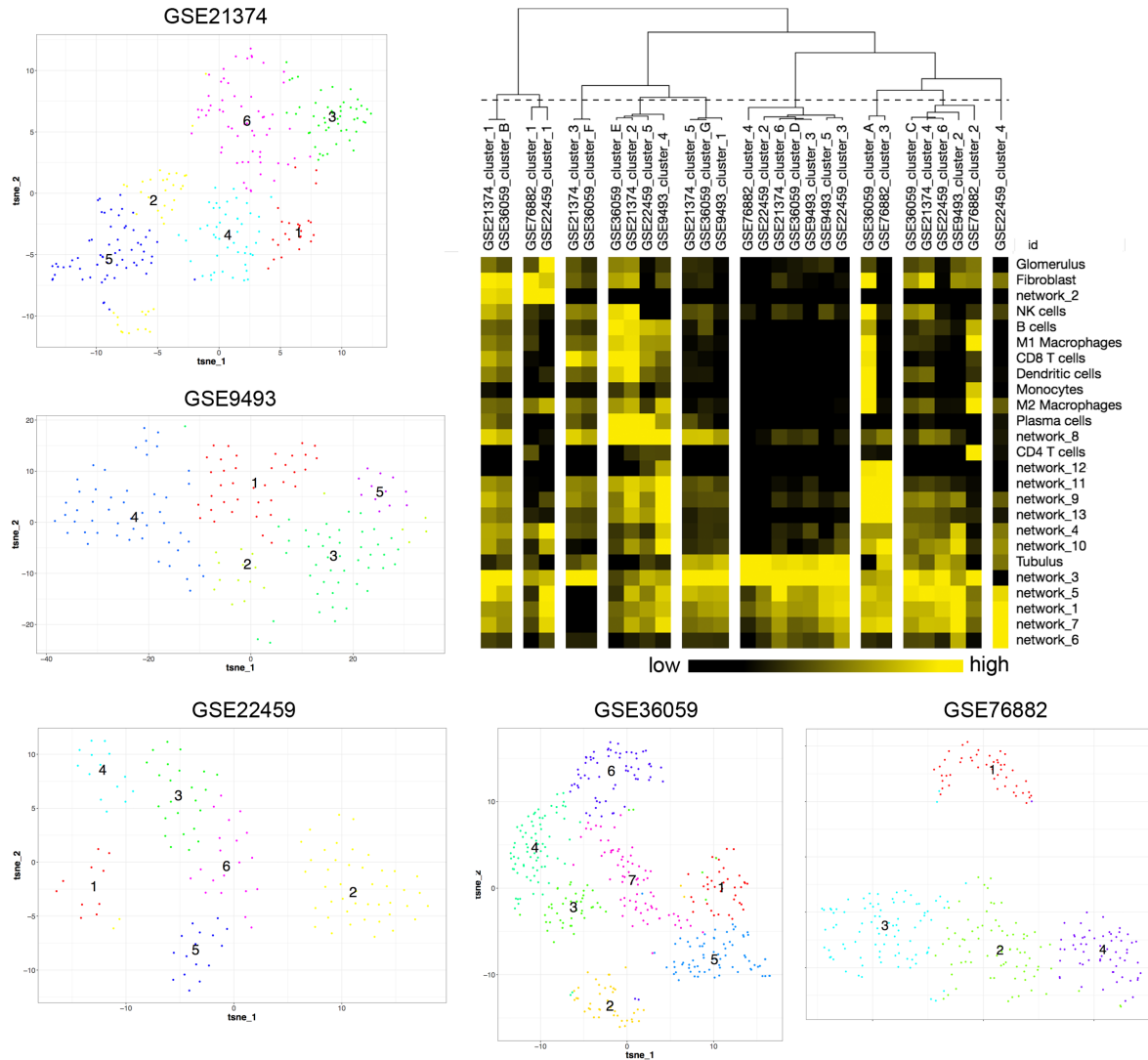
**Supplemental figure 6: Collagen and MMP gene correlation with cell types.**

Individual genes were Pearson correlated with the KID9plus3 cell type abundance.

GSE36059, n=411.

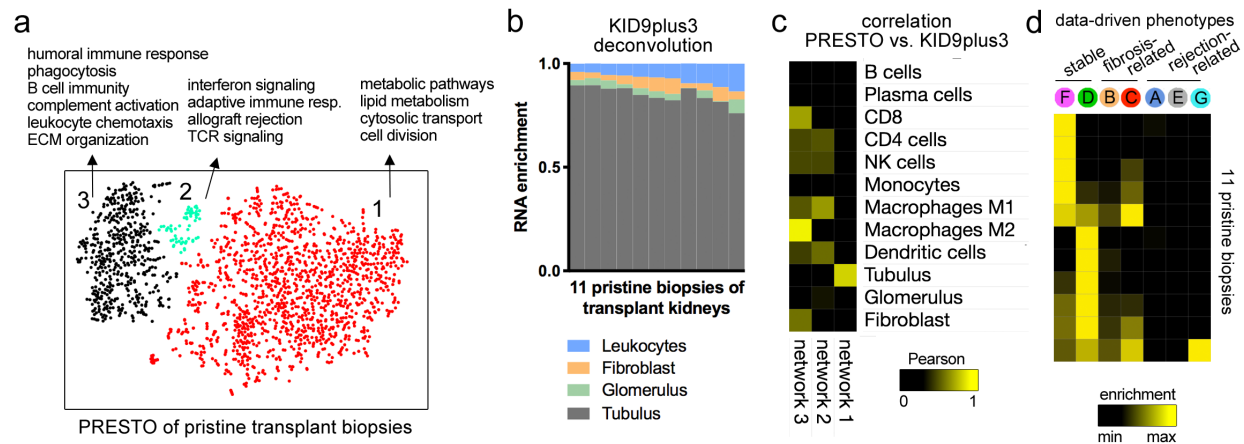


**Supplemental figure 7: Cell type specific gene network activity in an independent data set.** a) The PRESTO gene network correlation with leukocyte subsets based on RNA deconvolution was performed in an independent data set (GSE21374, n = 282 biopsies) similar to main figure 2d. b) Specific correlations are shown as XY histogram with linear regression analysis.

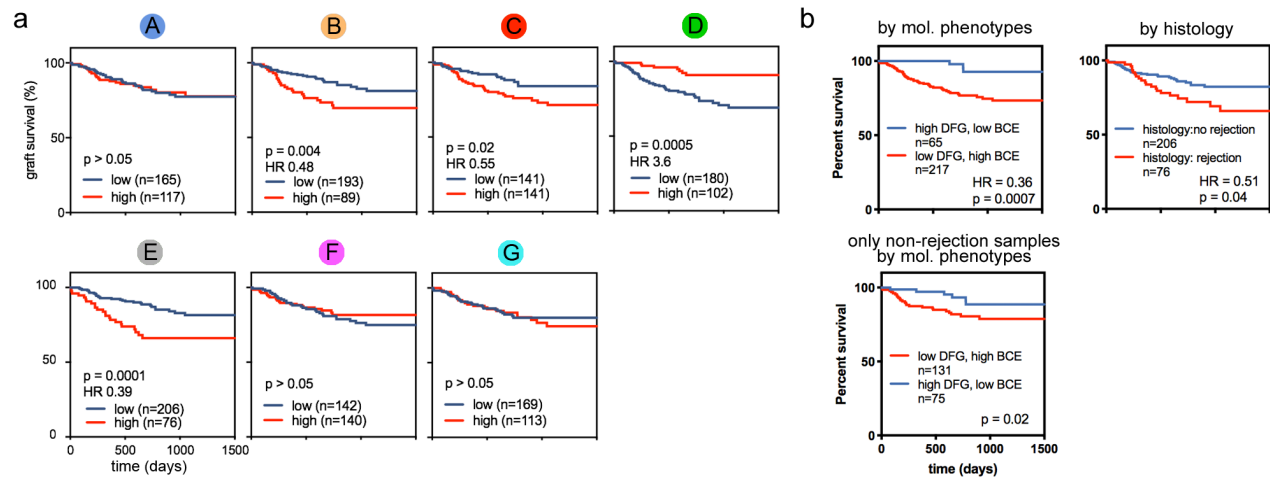


### Supplemental figure 8: Immune phenotype clustering in independent data sets.

Data driven high dimensional clustering and visualization was performed similar to main figure 3 using independent data sets GSE21374 (n=282), GSE9493 (n=82), GSE22459 (n=65), GSE76882 (n=274) (see materials and methods for more information and the data sets). GSE36059 is also shown in main figure 3. The heatmap shows differential expression patterns of input parameters (PRESTO gene network activity and KID9plus3 leukocyte enumeration). The dendrogram shows hierarchical clustering using Euclidean distance metrics (similar to main figure 3d).

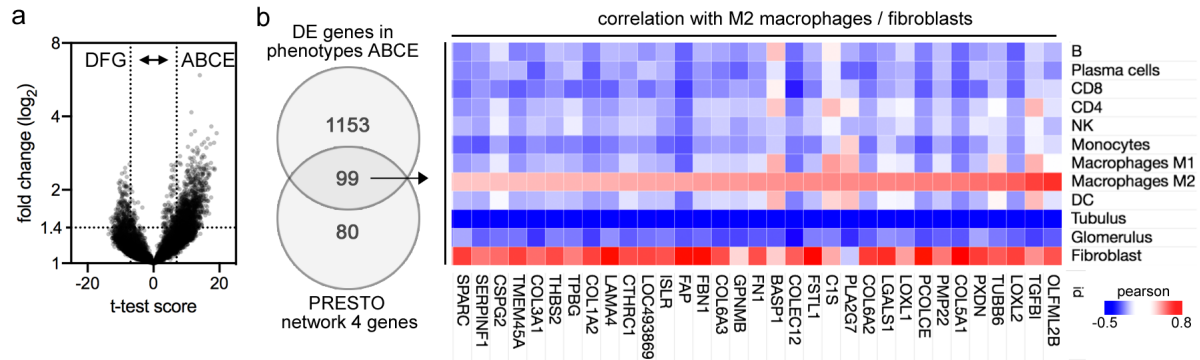


**Supplemental figure 9: Immune phenotypes in pristine biopsies.** Unsupervised phenotype analysis of pristine kidney transplant biopsies. 11 pristine biopsies from GSE30718 were analyzed similar to our main analysis in the manuscript. a) PRESTO gene network analysis of 1914 genes was performed similar to main figure 2a and resulted in 3 main networks. Enriched pathways for the three networks are shown. b) KID9plus3 deconvolution analysis of pristine biopsies similar to main figure 1c. c) Correlation of leukocyte enumeration data and gene network activity similar to main figure 2d. d) Deconvolution of phenotypes using the 7 phenotypes A-G developed in main figure 3. Phenotypes F and D are annotated as “stable”, C and B as “fibrosis-related”, and A and E as “rejection-related” according to main figure 3.

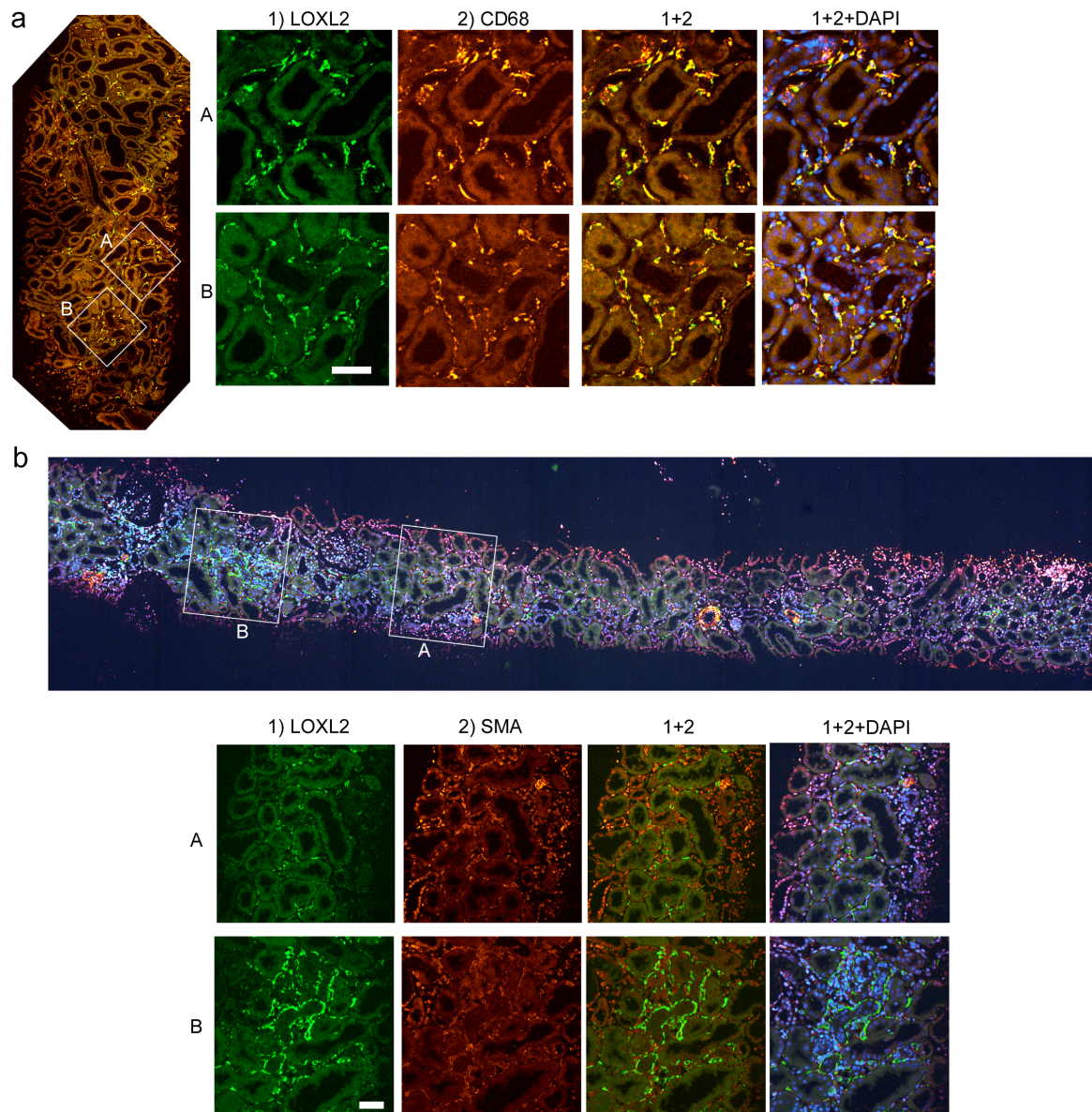


**Supplemental figure 10: Graft survival analysis in the independent data set GSE21374.** n=282. Related to figure 4. a) The enrichment of each phenotype in each sample was calculated and two groups (high and low expression) were defined based on the enrichment score as described in materials and methods. Hazard ratio (HR) and log rank p-value are indicated. b) Graft survival stratified by histology (rejection versus non-rejection by Banff) versus molecular phenotypes (low survival phenotypes DFG versus high survival phenotypes BCE). The lower panel shows graft survival by molecular phenotypes only in a subcohort of non-rejection biopsies (histology, n=206)

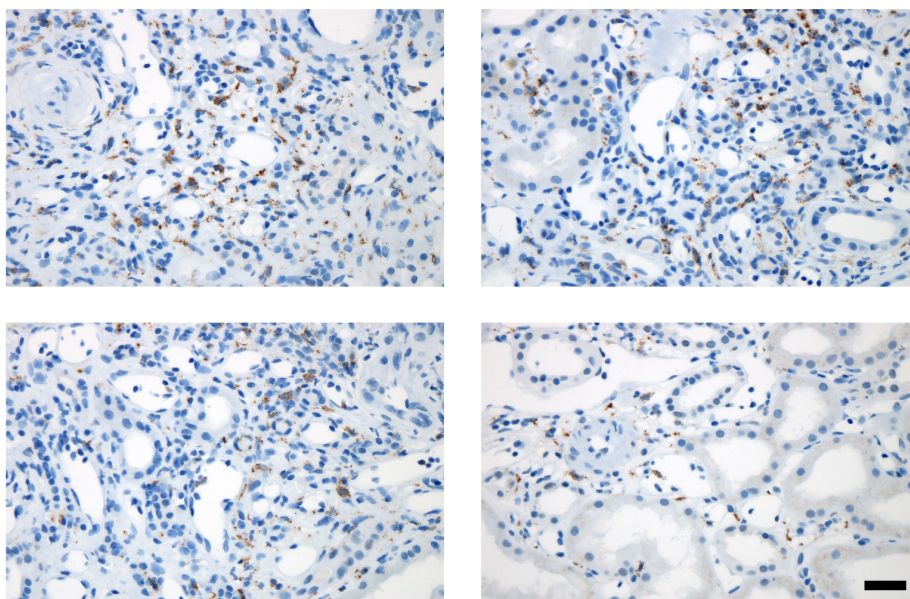




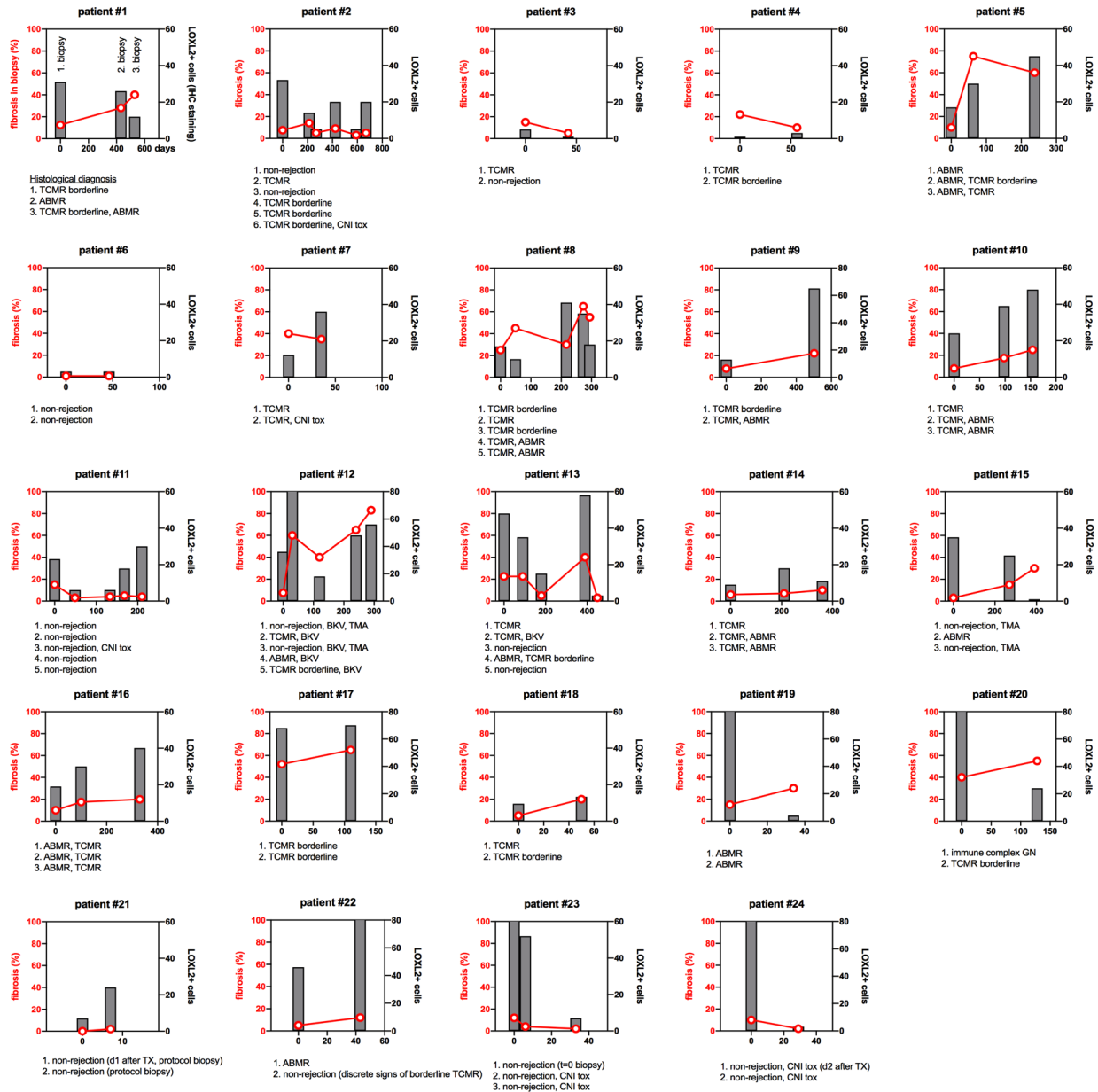
**Supplemental figure 11: Molecular candidate markers of dysfunctional transplant phenotypes.** a) Differentially expressed genes (DE) were calculated for phenotypes with reduced graft survival (A,B,C,E) versus phenotypes with normal graft survival (D,F,G) (see main figure 3) as described in materials and methods. b) The overlap of DE genes and PRESTO gene network 4 are 99 genes. They were correlated with the calculated cell type abundance and ranked by the KID9plus3 M2 macrophage enrichment score. Highly correlating genes are macrophage/fibroblast-expressed molecular candidates for graft dysfunction.



**Supplemental figure 12: Immunofluorescence microscopy of LOXL2 in human kidney biopsies.** A stitched image of multiple high resolution and multi color confocal micrographs is acquired using clinical kidney transplant biopsies. Nuclear staining with DAPI. Two fields of view are shown in detail. a) LOXL2 and CD68 staining. b) LOXL2 and smooth muscle actin (SMA) staining. Scale bar indicates 100  $\mu$ m.



**Supplemental figure 13: Immunohistochemical staining of LOXL2 in kidney transplant biopsies.** Representative IHC stainings of clinical transplant biopsies are shown. Scale bar 100um.



**Supplemental figure 14: Quantification of LOXL2+ peritubular macrophages and fibrosis in longitudinal biopsies.** 24 kidney transplanted patients were selected that underwent sequential indication biopsies (n=70). For each biopsy the histological fibrosis score (red line) and the LOXL2+ macrophage cell abundance (gray bar) was determined, and the BANFF diagnosis is indicated.

**Supplemental table 1:** Overview of independent validation data sets.

**Supplemental table 2: Gene network composition.** For each gene network 1-13 the gene names are indicated.

**Supplemental table 3: Functional annotation of gene networks.** The functional annotation list of gene networks 1-13 are shown.

**Supplemental table 4:** Clinical characteristics of the patients studied in figure 6.