Supplemental Data for

Kidney single-cell atlas reveals myeloid heterogeneity

in progression and regression of kidney disease

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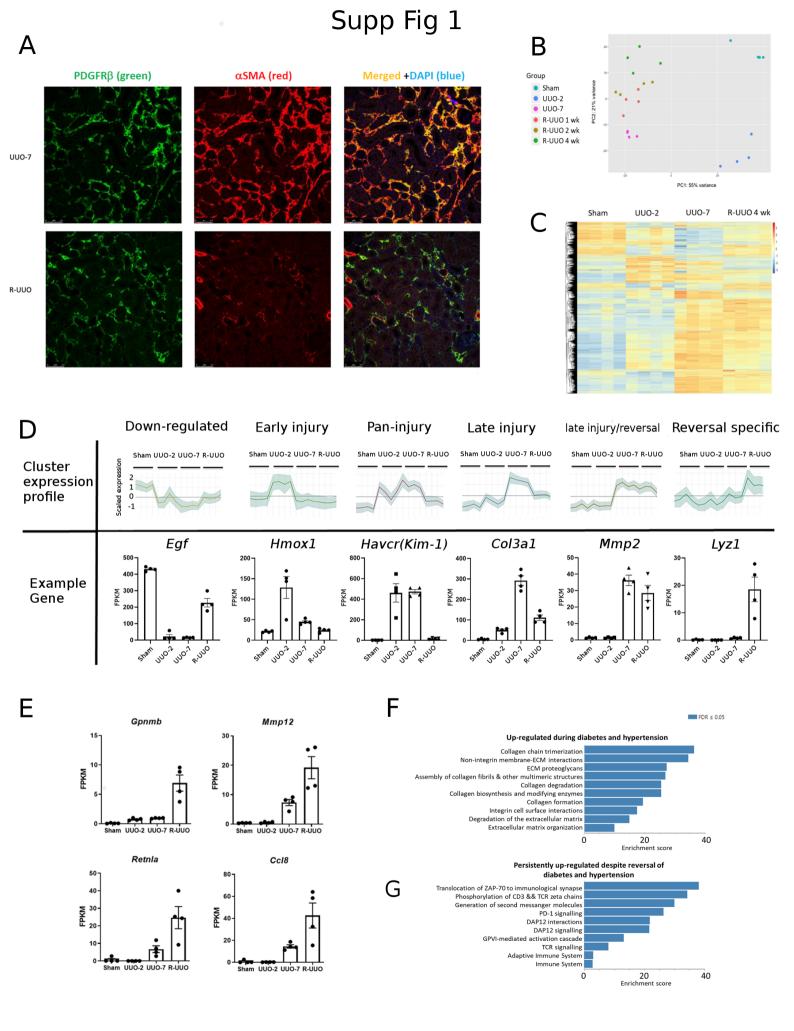
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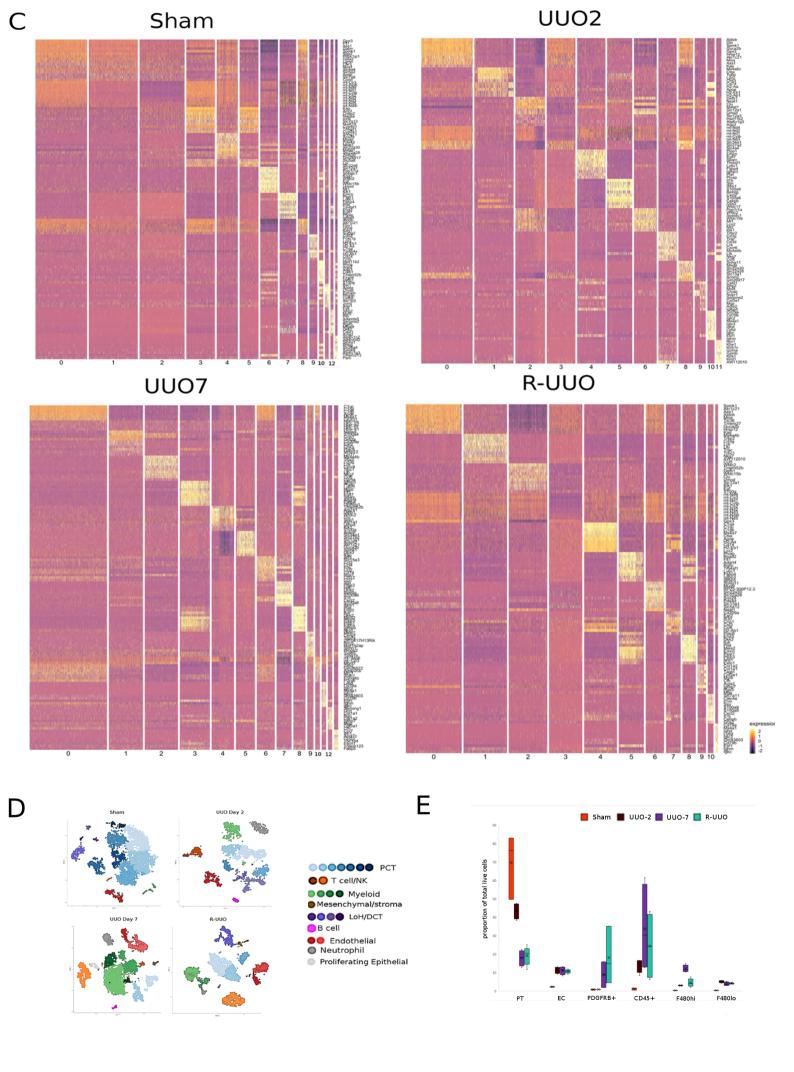
Short running title: Myeloid cells in kidney injury and repair

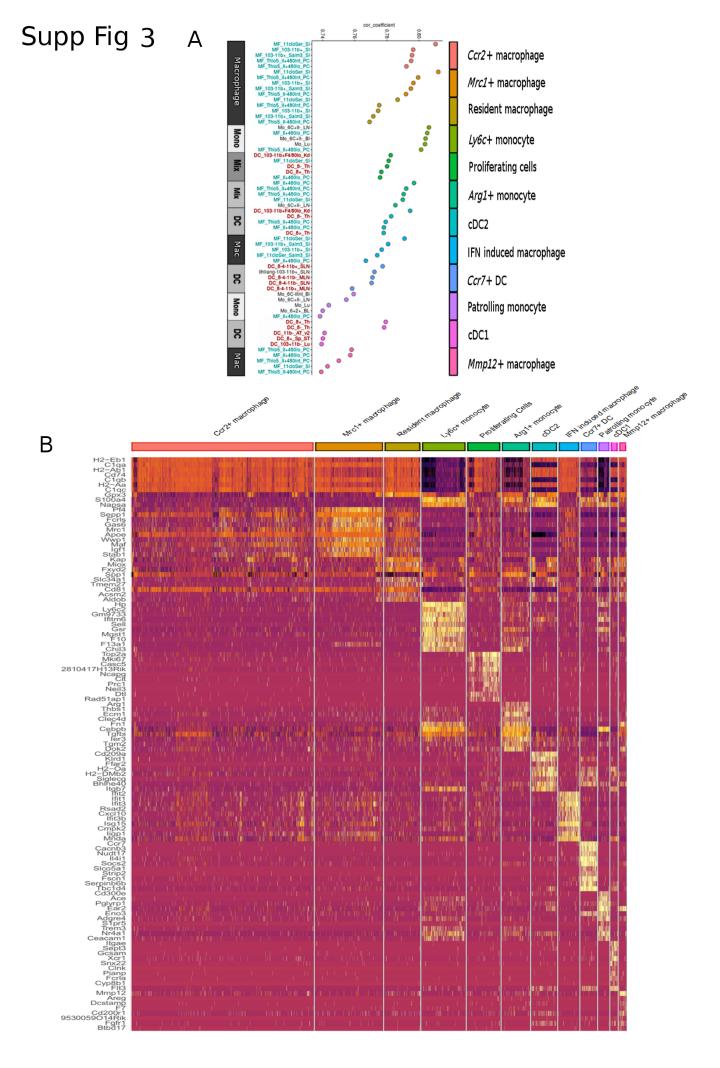
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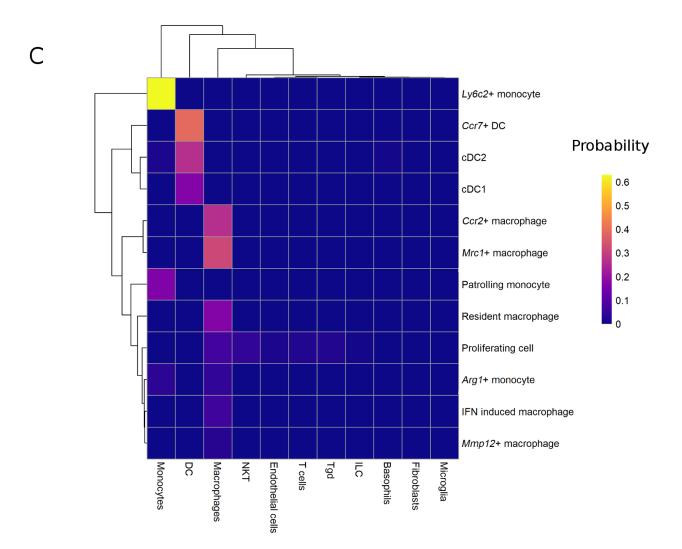
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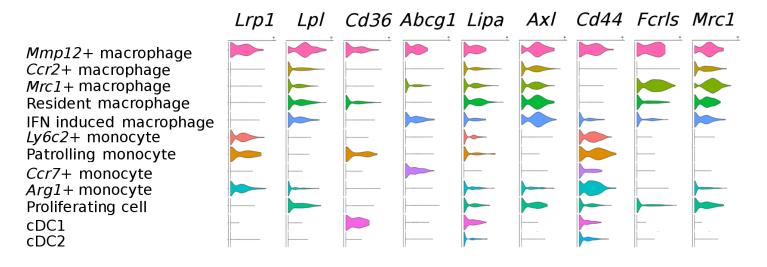
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String Part
String Part Myeloid: Proliferating
B cell PCT PCT a PCT b PCT S1 Endothelial b Collecting Duct LoH/DCT NK/T cell Endothelial a Mesencymal/Stroma Neutrophil Myeloid: Mac Myeloid: DC/Mono expression 2 1 0 -1 -2 ²⁸ S100a8 S100a9 Collal Serpine2 C Plat Ehd3 Aqp2 Fxyd4 Top2a Tubalb Clqa Adgre1 Umod Slc12a1 Cd3g Slc34a1 Gpx3 В PCT a Myeloid: Mac PCT b LoH/DCT PCT c NK/T cell Myeloid: DC/Mono Endothelial a Endothelial b Neutrophil **Collecting Duct** Mesencymal/Stroma Myeloid: Proliferating



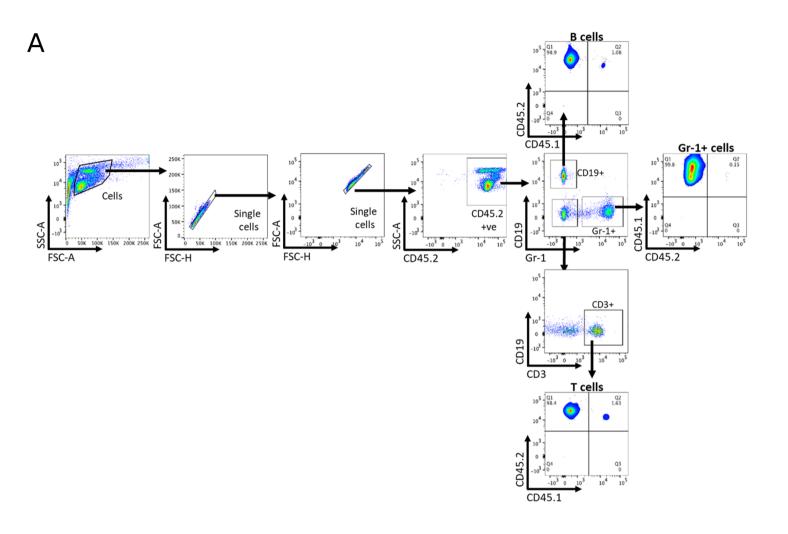


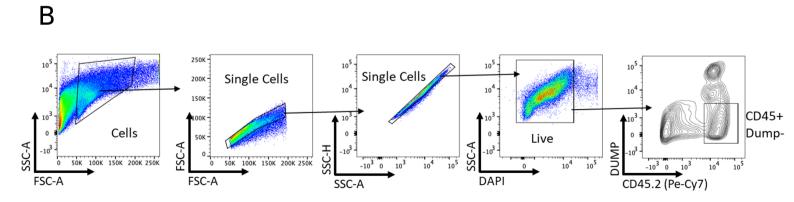






Supp Fig 4





Antibody	Company/Cat Num/Dilution
Collagen III	Southern Biotech/1330-08/1:200
PDGFR-β	Abcam/ab32570/1:500
α-SMA	Abcam/ab5694/1:400
F4/80	Abcam/ab16911/1:100
YM1 (Chil3)	Stemcell/60130/1:500
Mannose Receptor	R&D/AF2535/1:200
Mmp12	Santa Cruz/sc390863/1:100
Anti-rabbit biotinylated	Vector Labs/BA1000/1:300
Anti-goat biotinylated	Vector Labs/BA5000/1:300
Anti-rat biotinylated	Vector Labs/BA9400/1:300
Anti-rabbit AF555	ThermoFisher/A-11034/1:200
Anti-goat AF488	ThermoFisher/A-11055/1:200
Anti-mouse AF488	ThermoFisher/A-11029/1:200
Anti-rat AF488	ThermoFisher/A-21434/1:200

 $\begin{array}{lll} \textbf{Supp Table 1.} & \textbf{Antibodies utilised in immunohistochemistry/immunoflourescence studies} \end{array}$

Gene	Probe/Assay ID
Mmp12	Mm00500554_m1
Gpnmb	Mm01328587_m1
Lyz1	Mm00657323_m1
Hprt	Mm00446968_m1

Supp Table 2. Taqman probes used for qRT-PCR in studies

Antibody	Clone/Flurochrome/final concentration
Live	DAPI / 1:1000
CD45	30-F11/BV650 or APC/1:100
CD45.1	A20/FITC/1:200 K or 1:100 blood
CD45.2	104/Pe-Cy7/1:200 K or 1:100 blood
F4/80	BM8/Pe-Cy7 or APC / 1:100 or 1:200 K or 1:100
	blood
MHCII	M5-114.15.2/APC-Cy7 / 1:400
Ly6G	1A8/BV421 / 1:200
Ly6C	HK1.4/AF700 / 1:200
CD11b	M1_70/PE Dazzle / 1:1000
CD11c	N418/BV605 / 1:100
CD64	X54-5/7.1/PerCp5.5/1:200
CD24	30-F1/PE/1:200
Siglec-F	E50-2440/BV421/1:200
TCRβ	H57-597/BV421/1:200
CD19	6D5/BV421/1:200
CD3	17A2/PerCp5.5 1:100
CD206	MR5D3/APC / 1:200
GR-1	RB6-8C5/PE/1:200
CCR2	SA203G11/BV605/1:200
PDGFRβ	APB5/PE/1:50
CD31	390/BV605/1:100
LTL	FL-1321/FITC/1:200

Supp Table 3. Antibodies used in flow cytometry

Supplemental Figure 1

A Immunofluorescence for PDGFR- $β^+$ and α-SMA⁺ indicates that a proportion of PDGFR- $β^+$ cells co-express α-SMA⁺ at UUO-7 indicating activation to myofibroblasts; 1 week following R-UUO, the interstitial PDGFR- $β^+$ cells no longer express α-SMA⁺, which is now restricted to the arterial smooth muscle walls. **B** Principal component (PC) analysis of RNA sequencing data from bulk cortical kidney tissue from animals undergoing sham surgery, or at UUO-2, UUO-7, or 1-2- or 4-weeks post R-UUO (n=4/time-point). **C** Heatmap of differentially expressed genes created using Pheatmap R package. Rows were clustered via unsupervised hierarchical clustering. **D** Expression of exemplar genes aligning to each cluster **E** Expression of *Gpnmb*, *Mmp12*, *Retnla* and *Ccl8* genes in RNAseq dataset **F** Pathways enriched for genes that are induced in the kidney of *Cyp1a1mRen2* rats during diabetes and hypertension, but which revert towards baseline following tight glycaemic and blood pressure control (52) **G** Pathways enriched for genes that are persistently elevated in the kidney of *Cyp1a1mRen2* rats despite onset of tight glycaemic and blood pressure control (52).

Supplemental Figure 2

A Top 10 differentially expressed genes by fold change in each cluster, calculated using Wilcoxon signed-rank test. The colour scheme is based on z-score distribution. **B** Violin plots showing the expression levels of selected marker genes in each cluster. The x axis shows the log-scale normalized read count. **C** Top 10 differentially expressed genes by fold change in each cluster across the sham, UUO-2, UUO-7 and R-UUO (2 weeks) libraries analysed individually using Wilcoxon signed-rank test. The colour scheme is based on z-score distribution **D** Separate tSNE plots restricted to cells from each separate time-point coloured by shared nearest neighbour (SNN) allocated cluster, annotated by cell type **E** Flow cytometry was employed to determine the proportion of total kidney cells at each time point comprised by proximal tubular cells (PT), endothelial cells (EC), fibroblasts (PDGFR-β), immune cells (CD45), and F4/80^{Hi} or F4/80^{Lo} macrophages.

Supplemental Figure 3

A Top 5 Immunological Genome Project (ImmGen) reference cell types per cluster ranked by Spearman's correlation coefficients with full dataset details following a cluster-to-references analysis using cluster identify predictor v2. B Top 10 differentially expressed genes by fold change per cluster, calculated using Wilcoxon signed-rank test. The colour scheme is based on z-score distribution C Consensus matrix of ImmGen reference cell types using SingleR generated labels (bottom) with final classifications D Violin plots of efferocytosis genes across the myeloid cell clusters. The y-axis shows the log-scale normalized read count.

Supplemental Figure 4

A Flow cytometry gating strategy for characterizing circulating blood immune cells into B-cells (CD19 $^+$), monocyte/neutrophils (Gr-1 $^+$), and T-cells (CD3 $^+$) using the D2 blood sample as an exemplar. **B** Flow cytometry gating strategy for sorting myeloid cells in the kidney. Lineage dump gate includes TCR β , CD19, Ly6G and SiglecF all on the same fluorophore to exclude T-cells, B-cells, neutrophils and eosinophils respectively.

Supplemental Table 1

Antibodies utilised in immunohistochemistry/immunofluorescence studies.

Supplemental Table 2

Tagman probes used for qRT-PCR in studies.

Supplemental Table 3

Antibodies utilised in Flow Cytometry

Supplemental Table 4

Summary of 10x sequencing statistics

Supplemental Table 5

Bulk RNA-Seq matrix of differentially expressed genes in each gene cluster, expressed in FKPM.

Supplemental Table 6

Single cell RNA-Seq table of differentially expressed genes in the global data as described in Fig. 2, expressed per cluster, where avg_logFC is the average log fold change between the cluster in question and the remaining clusters, pct.1 and pct.2 is the percentage of cells expressing the gene in question in the cluster in question the remaining clusters, and p_val_adj is the adjusted P-value following Bonferroni correction for multiple hypothesis testing.

Supplemental Table 7

Single cell RNA-Seq table of differentially expressed genes in the myeloid only data as described in Fig. 3, expressed per cluster, where avg_logFC is the average log fold change between the cluster in question and the remaining clusters, pct.1 and pct.2 is the percentage of cells expressing the gene in question in the cluster in question the remaining clusters, and p_val_adj is the adjusted P-value following Bonferroni correction for multiple hypothesis testing and rank is the assigned rank for gene set enrichment analysis calculation.