

Supplementary Data  
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**Supplementary Table 1:** Clinical Study Participants Characteristics.

	<b>Uninfected study participants</b>	<b>HIV-1 infected study participants</b>
<b>Number of subjects</b>	14	24
<b>Age (yrs)</b>	31 (23-54)	33.5 (22-58)
<b>Male/Female Ratio</b>	9/5	18/6
<b>CD4 count (cells/<math>\mu</math>l)*</b>	724 (468-1071)	445 (221-1248)
<b>Plasma viral load (HIV-1 RNA copies/ml)</b>	-	51,350 (2880-207,000)
<b>Race:</b>		
White/Caucasian	10 (71.4%)	17 (70.8%)
Black/African American	2 (14.3%)	6 (25%)
Asian	2 (14.3%)	1 (4.2%)

Values are shown as median (range) or the number (percentage) of each cohort. Statistical analysis performed using Mann-Whitney test for comparisons between uninfected and HIV-1 infected study participants and the Fisher Exact test or Chi-square test for comparison of categorical data. \*p=0.001.

**Supplementary Table 2:** Study Participants Characteristics (RUMC IRB gastrointestinal repository)

	<b>Uninfected study participants</b>	<b>HIV-1 infected study participants</b>
<b>Number of subjects</b>	15	15
<b>Age (yrs)</b>	52 (39-62)	51 (42-59)
<b>Male/Female Ratio</b>	11/4	11/4
<b>CD4 count (cells/<math>\mu</math>l)</b>	n/d	508 (178-1415)
<b>Plasma viral load (HIV-1 RNA copies/ml)</b>	-	<40
<b>Race:</b>		
White/Caucasian	2 (13.3%)	2 (13.3%)
Black/African American	13 (86.7%)	13 (86.7%)

Values are shown as median (range) or the number (percentage) of each cohort. Statistical analysis performed using Mann-Whitney test for comparisons between uninfected and HIV-1 infected study participants and the Fisher Exact test or Chi-square test for comparison of categorical data. n/d: not determined.

**Supplementary Table 3:** PBMC and colon biopsies with appropriate RNA or GAPDH levels for accurate Type I IFN and ISG gene measurements.

		UC-AMC Cohorts	
		Uninfected study participants (Total N=14)	HIV-infected study participants (Total N=24)
		<i>PBMC</i>	
<b>IFNα</b>		14	24
<b>IFNα subtypes</b>		8	13
<b>IFNβ</b>		14	24
		<i>Colon biopsies</i>	
<b>IFNα</b>		12	18
<b>IFNα subtypes</b>		7	11
<b>IFNβ</b>		12	18
<b>Tetherin</b>		11	16
<b>APOBEC3G</b>		11	16
<b>MX2</b>		13	18
		RUMC Cohorts	
		Uninfected study participants (Total N=11)	HIV-infected study participants (Total N=15)
		<i>Colon biopsies</i>	
<b>IFNα</b>		11	15
<b>IFNβ</b>		11	15
<b>Tetherin</b>		11	15
<b>APOBEC3G</b>		11	15
<b>MX2</b>		11	15

Samples were excluded due to insufficient RNA or when GAPDH levels <200 copies/ng.

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**Supplementary Table 4.** Correlations between colonic IFN $\alpha$  and IFN $\beta$  gene transcripts and clinical, virological and immunological parameters.<sup>#</sup>

	<b>IFN<math>\alpha</math></b>	<b>IFN<math>\beta</math></b>
<i>Clinical Parameters</i>		
Blood CD4 T cell count	R=-0.23, P=0.36	R=-0.17, P=0.51
Plasma HIV-1 viral load	R=-0.10, P=0.70	R=-0.02, P=0.95
<i>Systemic inflammation, immune activation, microbial translocation and epithelial barrier disruption parameters</i>		
Plasma IL-6	R=-0.42, P=0.08	R=0.38, P=0.12
Plasma C-reactive protein (CRP)	R=-0.27, P=0.28	R=0.27, P=0.29
Plasma TNF $\alpha$	R=-0.50, P=0.09	R=0.40, P=0.18
Plasma IFN $\gamma$	R=-0.09, P=0.77	<b>R=0.61, P=0.03</b>
Plasma IL-10	R=-0.24, P=0.44	<b>R=0.58, P=0.04</b>
Activated blood CD4 T cells	R=-0.07, P=0.77	R=-0.18, P=0.48
Activated blood CD8 T cells	R=0.07, P=0.77	R=-0.03, P=0.89
Plasma soluble CD27 (sCD27)	R=0.02, P=0.94	R=0.03, P=0.92
Plasma sCD14	R=-0.45, P=0.09	R=0.31, P=0.26
Plasma lipopolysaccharide (LPS)	R=-0.08, P=0.76	R=-0.18, P=0.51
Plasma lipoteichoic acid (LTA)	R=-0.10, P=0.71	R=-0.30, P=0.27
Plasma intestinal fatty acid binding protein (iFABP)	R=0.05, P=0.83	R=0.09, P=0.74
<i>Colonic immunity parameters</i>		
HIV-1 RNA levels	R=0.12, P=0.63	R=0.03, P=0.88
Number of CD1c $^+$ mDC	R=0.12, P=0.64	R=0.38, P=0.14
Number of pDC	R=-0.12, P=0.66	R=0.04, P=0.88
CD40 expression levels on CD1c $^+$ mDC	R=0.07, P=0.79	R=0.09, P=0.72
CD40 expression levels on pDC	R=0.04, P=0.87	R=0.00, P=1.0
Percent of CD83-expressing CD1c $^+$ mDC	R=0.40, P=0.12	R=0.29, P=0.26
Percent of CD83-expressing pDC	R=0.24, P=0.36	R=0.35, P=0.17
Number of CD4 T cells	R=-0.17, P=0.49	R=0.00, P=1.0
Number of CD8 T cells	R=-0.08, P=0.75	R=0.23, P=0.35
Number of activated CD4 T cells	R=-0.08, P=0.76	R=0.003, P=0.99
Number of activated CD8 T cells	R=0.11, P=0.67	R=0.22, P=0.38
Number of IFN $\gamma$ -expressing CD4 T cells	R=-0.20, P=0.44	R=0.43, P=0.08
Number of IL-17-expressing CD4 T cells	R=-0.22, P=0.40	R=0.44, P=0.08
Number of IL-22-expressing CD4 T cells	R=0.22, P=0.40	R=0.28, P=0.27
Number of IFN $\gamma$ -expressing CD8 T cells	R=0.07, P=0.79	R=0.12, P=0.65
Constitutive IL-12p70 production	R=0.13, P=0.67	R=0.30, P=0.30
Constitutive IL-23 production	R=-0.06, P=0.83	R=0.24, P=0.40
Constitutive IL-10 production	R=0.27, P=0.35	R=-0.08, P=0.80
Constitutive IL-1 $\beta$ production	R=-0.19, P=0.51	R=-0.06, P=0.83
Constitutive TNF $\alpha$ production	R=-0.13, P=0.65	R=-0.10, P=0.72
Constitutive IL-6 production	R=-0.19, P=0.51	R=-0.04, P=0.89
Constitutive IFN $\gamma$ production	R=-0.14, P=0.60	R=0.06, P=0.83
Constitutive IL-17 production	R=-0.18, P=0.50	R=0.02, P=0.95

Statistical analysis performed using the Spearman test. <sup>#</sup>Bold font indicates statistically significant correlation.

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**Supplementary Table 5.** Correlations between colonic Type I IFN gene transcripts.\*

	IFN $\alpha$	IFN $\beta$	Tetherin	APOBEC3G	MX2
IFN $\alpha$	---				
IFN $\beta$	R=-0.10, P=0.70	---			
Tetherin	R=-0.03, P=0.93	R=0.43, P=0.11	---		
APOBEC3G	R=0.10, P=0.93	R=0.41, P=0.13	<b>R=0.81, P=0.0002</b>	---	
MX2	R=-0.12, P=0.66	R=0.26, P=0.30	<b>R=0.84, P=0.0004</b>	<b>R=0.58, P=0.03</b>	---

Statistical analysis was performed using the Spearman test. \*Bold font indicates statistically significant correlation.